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Published in:
Medical Physics

Publication date:
2006

Document Version
Publisher's PDF, also known as Version of record

Link to publication from Aalborg University

Citation for published version (APA):

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Automated detection of a prostate Ni–Ti stent in electronic portal images

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(Received 23 December 2005; revised 26 September 2006; accepted for publication 28 September 2006; published 20 November 2006)

Planning target volumes (PTV) in fractionated radiotherapy still have to be outlined with wide margins to the clinical target volume due to uncertainties arising from daily shift of the prostate position. A recently proposed new method of visualization of the prostate is based on insertion of a thermo-expandable Ni–Ti stent. The current study proposes a new detection algorithm for automated detection of the Ni–Ti stent in electronic portal images. The algorithm is based on the Ni–Ti stent having a cylindrical shape with a fixed diameter, which was used as the basis for an automated detection algorithm. The automated method uses enhancement of lines combined with a grayscale morphology operation that looks for enhanced pixels separated with a distance similar to the diameter of the stent. The images in this study are all from prostate cancer patients treated with radiotherapy in a previous study. Images of a stent inserted in a humanoid phantom demonstrated a localization accuracy of 0.4–0.7 mm which equals the pixel size in the image. The automated detection of the stent was compared to manual detection in 71 pairs of orthogonal images taken in nine patients. The algorithm was successful in 67 of 71 pairs of images. The method is fast, has a high success rate, good accuracy, and has a potential for unsupervised localization of the prostate before radiotherapy, which would enable automated repositioning before treatment and allow for the use of very tight PTV margins. © 2006 American Association of Physicists in Medicine. [DOI: 10.1118/1.2369466]

I. INTRODUCTION

Increased doses in external beam radiotherapy (EBRT) of localized prostate cancer increase local control although increased survival rate has not yet been shown.1 Organs at risk in prostate cancer patients treated with EBRT are bladder and rectum. The use of modern imaging techniques and three-dimensional (3D) conformal treatment planning may reduce the volume of normal tissue irradiated.2–4 When normal tissue volume is reduced, evidence of consequently reduced toxicity exists, even with increased dose to the prostate.5 Even further reduction of normal tissue volume in the high dose region may be possible using intensity modulated radiotherapy.6 Planning target volume (PTV) in fractionated EBRT, however, still has to be outlined with wide margins to the Clinical Target Volume due to geometrical uncertainties. Uncertainties arise from shift in daily prostate position due to variation in bladder and rectal filling7,8 and setup errors. Daily variations in the position of the prostate are in the range 10–20 mm.9 Errors may be even greater for patients in prone position due to movements caused by respiration.10,11 Various methods have been developed to reduce PTV margins. These methods are based on indirect visualization of the prostate using x-ray or electronic portal imaging devices (EPIDs). Visualization can be done using radio opaque implanted 2–5 mm gold (Au) markers.9,12–15 Automated detection of the markers will facilitate daily clinical use. Several algorithms such as template matching,16 marker extraction kernels17 and Mexican hat filtering18 have been investigated for automated detection of gold markers. Detection success rate is between 85% and 100%.17–19 and positioning accuracy within 1–2 mm.17,19 Image noise, bony structures and air cavities reduce the detection success rate of gold markers.17,19 Recently a new method of visualization of the prostate has been proposed, which is based on insertion of a thermo-expandable Ni–Ti stent (Memokath®, Engineers and Doctors) in the prostatic urethra. The Memokath stent was developed for treatment of bladder outlet obstruction in patients with benign prostate hyperplasia.20 The stent is visible on electronic portal images from an amorphous silicon imager (Varian as500) and can be positioned with the same accuracy as gold markers. In the present study a new detec-
tion algorithm suited for automated positioning of the Ni–Ti stent is proposed based on a fixed 3D shape and dimension of an inserted stent. The algorithm is using a gray scale morphology operation that looks for enhanced pixels within a given distance equal to the stent diameter. The present study investigated the feasibility of automated detection using this new algorithm to look for a Ni–Ti stent in portal images.

II. MATERIAL AND METHODS

The images in this study are all from patients included in a previous study. All patients presented with histological verified local or locally advanced prostate cancer (stage pT2b to pT3b, N0, M0; UICC 1992 classification was used) were all candidates for intended curative radiotherapy. The stent used in the study is shown in Fig. 1. The stent is made of a coiled Ni–Ti wire and has a cylindrical shape. External and internal diameters of the cylinder are 8.0 and 6.7 mm respectively, i.e., a wall thickness of 0.65 mm. The collar has an expanded maximal external diameter of 13 mm. Because of the cylindrical shape of the stent, the distance of Ni–Ti wire traversed by a beam orthogonal to the cylinder axis is rising from 0 at the external diameter to 4.3 mm when the beam is tangential to the internal diameter as shown in Fig. 2. The average distance traversed by the beam is around 3 mm. This is equivalent to an approximately 4% contrast enhancement. Consequently, the cylindrical shape is imaged as two parallel lines with higher than background absorption. The physical pixel size of the Varian as500 imager is 0.784 mm/pixel. This was scaled to the isocenter using the accelerator source to axis distance and source to detector distance, which are read from the dicom header in each individual image. This gave a pixel size of approximately 0.56 mm in the isocenter, but dependent on the actual position of the imager. Treatment was given with an isocenter 3D conformal treatment plan including three multi leave collimator fields (one anterior and two lateral wedged fields). Pairs of orthogonal 15 cm × 15 cm isocenter setup fields (anterior and lateral projections) were used to obtain electronic portal images in eight treatment sessions in each patient (session numbers 1, 2, 3, 6, 11, 21, 22 and 23). Electronic portal images were recorded using a Varian As500 electronic portal imaging device (EPID). The acquisition protocol was the Varian standard protocol, i.e., the image was averaged over four frames taken with 6 MV using in total between 3 and 5 MU. The orthogonal images were used to determine the 3D reference position of the stent. The stent position was calculated manually using the orthogonal reconstruction in the Brachyvision® program which is part of the Eclipse® dose planning system from
Varian. All images were analyzed with both the manual and the new automated method. The manually determined 3D position of the stent was used as reference for the automated algorithm. Precision of the manual method was also estimated using double readings. In order to estimate localization accuracy a number of images were taken of a Ni–Ti stent placed in a humanoid phantom (Alderson). The stent was inserted in cranial-caudal (CC) direction as a hole was drilled in consecutive slices of the Alderson phantom. The phantom was computer tomography scanned and a treatment plan including setup fields analogous to that used in real patients was made. The phantom was placed on the accelerator couch. The couch was then moved to predefined positions using the readout of couch position. The position of the stent was subsequently determined using the automated method on sets of orthogonal images taken after each couch movement.

A. Automated detection algorithm

The automated algorithm is based on enhancement of vertical lines, followed by a morphology erosion, binary threshold and finally extraction of the largest particle. All image processing is done in ImageJ.22 Before enhancement of vertical lines the portal image was rotated until the cylindrical axis of the stent was vertical. The angle of rotation was initially determined using double readings. In order to estimate localization accuracy a number of images were taken of a Ni–Ti stent placed in a humanoid phantom (Alderson). The stent was inserted in cranial-caudal (CC) direction as a hole was drilled in consecutive slices of the Alderson phantom. The phantom was computer tomography scanned and a treatment plan including setup fields analogous to that used in real patients was made. The phantom was placed on the accelerator couch. The couch was then moved to predefined positions using the readout of couch position. The position of the stent was subsequently determined using the automated method on sets of orthogonal images taken after each couch movement.

for each portal image followed by a convolution with a 5 × 5 kernel to enhance vertical lines. The 5 × 5 kernel used is

\[
\begin{bmatrix}
-1 & 0 & 2 & 0 & -1 \\
-1 & 0 & 2 & 0 & -1 \\
-1 & 0 & 2 & 0 & -1 \\
-1 & 0 & 2 & 0 & -1 \\
-1 & 0 & 2 & 0 & -1 \\
\end{bmatrix}
\]

This convolution enhances line structures that are vertical, including the stent. The stent will show up in the images as two parallel enhanced lines typically 12–13 pixels apart, and each of the lines 1–2 pixels wide. Following the convolution a grayscale morphology operation is performed using the grayscale morphology plug in ImageJ. The plug in performs a gray level erosion using a structure element consisting of two points with a horizontal distance equal to the fixed diameter of the stent [see inset in Fig. 4(b)]. Subsequently a binary threshold of the eroded image is done using maximum entropy in the multi threshold plug in written for ImageJ. Finally a particle analysis is run on the threshold image. The particle analysis looks for eight neighbor pixels, all to be part of the same particle. All particles touching the edge of the image are excluded. The largest particle is then extracted. Center of gravity for the largest particle is used as representation point of the stent. The automated detection is done separately for both images in each pair of orthogonal images. The coordinate of the stent representation point is calculated relative to the image center (beam axis). The beam axis is also determined in each portal image. A binary threshold value equal to 20% the maximal grayscale value in the portal images is set. This value is chosen to make the beam portal appear white and the remaining detector area black. The beam axis is set equal to the center of gravity of the white area. The 3D coordinates are then calculated from the two two-dimensional projections using a previously published method for localization of implants from biplane radiographs.23 The automated method is characterized as being successful in an image pair if the difference between stent position coordinates deviates less than 2 mm from the same coordinates determined using the manual method in both images. All image processing is done using Java plug in’s and macros in the image processing toolbox ImageJ from National Institute of Health.

III. RESULTS

Localization accuracy was tested placing a Ni–Ti stent in a humanoid phantom (Alderson). The results from this test are shown in Table I. The difference between the couch position and the position determined with the algorithm was not statistically different from 0 for either the left-right (LR) or the anterior-posterior (AP) positions. For the cranial-caudal (CC) coordinate there was a significant difference. In general the test demonstrated a localization accuracy of 0.4–0.7 mm (one standard deviation), which was approximately equal to the EPID detector pixel size in the isocenter. A systematic difference of 0.8 mm was, however, observed in the CC di-
rection. Precision of the manual method was determined from double readings and demonstrated to be less than 0.4 mm. Patient images from nine patients were analyzed using the automated method. The images were acquired in eight different treatment sessions for each patient. In total, 71 pairs of orthogonal images were acquired; one pair was missing. Initially automated detection was attempted applying template match analysis (data not shown). The template match analysis was, however, critically dependent on a correct shape, alignment and scaling of template and image. Thus the template match analysis did not provide the same success rate as the grayscale morphology algorithm. The various steps in the automated method are demonstrated Fig. 3. The automated method on average took less than 0.5 s to determine the position of the stent in an image. Comparison of the 3D coordinates determined with both the manual and automated method is shown in Fig. 5, and a good correspondence between the (LR), (AP) and (CC) coordinates is seen.

From the calculated differences in Table II it can be seen that no statistical difference between the manual and automated method was observed. Localization accuracy of the automated method was shown to be in the range of 0.4–0.7 mm. Analogously the differences between the manual and automated method in patient images demonstrated standard deviations of 0.3–0.8 mm, equal to 1–2 pixels. Apparently the LR and AP coordinates demonstrate the smallest differences. From Table II it can be seen that the automated method failed in four cases, i.e., the differences between manual and automated detection were larger than the success criteria of 2 mm. However, differences larger than 5 mm were only seen in one case (highly deteriorated image).

**IV. DISCUSSION**

A new automated detection of a Ni–Ti stent inserted into the prostate was developed. The method was successfully applied to detect the position of the Ni–Ti stent in portal images. Detection of a thin metal marker in portal images is difficult. The detection is made difficult by high noise level, low contrast and background structures such as bony edges or air cavities. This new method takes advantage of the stents cylindrical shape with a fixed diameter to overcome the low signal and high noise ratio in portal images. Failure was characterized as the difference between the manual and automated detection of more than 2 mm. The automated localization method failed in four cases. The detection failed completely in one lateral image of very poor quality (portal

| Table I. Localization of Ni–Ti stent placed in Alderson humanoid phantom. The table shows mean and standard deviation of difference between couch coordinate and coordinate determined by the algorithm. |
|---|---|---|
| | LR | AP | CC |
| Mean (mm) | 0.1 | 0.3 | 0.8 |
| StdErr (mm) | 0.4 | 0.7 | 0.4 |

**Fig. 4.** The figure demonstrates the various steps in the automated detection algorithm used: (a) The original electronic portal image enhanced using histogram equalizations; (b) line enhancement using $5 \times 5$ filtering. Small inset is an enlarged diagram of the morphology structure element. White dots are one pixel in the structure elements; (c) Grayscale morphology operation leaving only bright pixels with a distance equal to the diameter of the stent; (d) binary threshold of image using maximum entropy; (e) result from eight neighbor particle analysis selecting the largest particle; (f) Center of gravity from the largest particle in image; (e) is marked with a red cross on image (a).
imager defect) giving a difference larger than 5 mm. In total the method has yielded acceptable results in 67 of 71 image pairs (94%). The algorithm is susceptible to errors in the angle of rotation. Deviation from a vertical position of the stent may introduce variation in line enhancement. This in turn can lead to discontinuities of the gray values representing the stent and breakup of the particle during threshold. Another source of error can be the carbon fiber net of the linac treatment couch. The net may be enhanced as well as the stent. Part of the enhanced net can erroneously combine with part of the stent collar and be detected as part of the cylindrical shape. Because the couch net is a regular structure it may theoretically be removed using fact Fourier transform to filter out the frequency of the net. But in other cases bony structures combined with part of the stent collar. In such cases filtering in the Fourier domain would not solve the problem. In total, however, the algorithm was shown to be robust. Even when large variations in exposure were seen and different imagers were used. Differences between the manual and automated method larger than 2 mm do occur in images from several patients but only in the CC direction. Thus a lower precision in the CC coordinate seems to be a general weakness of the automated method and is observed in both phantom and patient images. It seems that the method has a low precision detecting the ends of the stent. As a consequence, extension or reduction of the detected stent may occur in one or both ends of the stent. This may lead to a shift primarily in the CC coordinate. This shift is, however, minimized because the CC coordinate is averaged over coordinates from two orthogonal images. The automated method may improve if both ends of the stent can be detected with greater precision. One way to check whether both stent ends have been detected correctly would be to calculate the total length of the stent from the detected end points. Checking the length of the stent would be important as criteria for stopping the algorithm, if the automated method is allowed to run unsupervised. The localization accuracy was demonstrated in the phantom pictures of this study to be equal to the size of 1 pixel. The accuracy of the algorithm was observed to be similar in patient images, even though this also includes the accuracy of the manual method as well. The present study is consistent with other studies using gold markers, which report an accuracy of 0.2–0.6 mm. Another study reports an accuracy similar to the present study. Here analysis was done on a large number of clinical images and with a resolution of the imag-

![Fig. 5. Results of localization of a Ni–Ti stent using both a manual and the new algorithm (automated method). Results are from 71 image pairs taken in different treatment sessions of nine different patients. Each patient is represented by a symbol. The position of the stent is shown in three separate plots defined using the three directions: (a) LR (left-right) (b) AP (anterior-posterior) and (c) CC (cranial-caudal). Also shown in each plot are the regression line and its 95% confidence limits.](image)

<table>
<thead>
<tr>
<th></th>
<th>LR</th>
<th>AP</th>
<th>CC</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean (mm)</td>
<td>0.0</td>
<td>0.0</td>
<td>0.0</td>
</tr>
<tr>
<td>StdDev (mm)</td>
<td>0.3</td>
<td>0.4</td>
<td>0.8</td>
</tr>
<tr>
<td>&gt;2 mm</td>
<td>0</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>&gt;5 mm</td>
<td>0</td>
<td>1</td>
<td>1</td>
</tr>
</tbody>
</table>

Table II. Mean and standard deviation of difference between manual and automated detection of stent coordinates. The three coordinates are determined from 71 pairs of orthogonal portal images. The last two rows are number of image pairs in which the difference is greater than 2 mm, but less than 5 mm and greater than 5 mm, respectively. The AP and CC differences in the last row occurred in the same image. The image was taken with a defect imager.
The method has a future potential for real time tracking of respiration movements.


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