

Aalborg Universitet

Human and computational models of atopic dermatitis

A review and perspectives by an expert panel of the International Eczema Council

Eyerich, Kilian; Brown, Sara J.; Perez White, Bethany E.; Tanaka, Reiko J.; Bissonette, Robert; Dhar, Sandipan; Bieber, Thomas; Hijnen, Dirk J.; Guttman-Yassky, Emma; Irvine, Alan; Thyssen, Jacob P.; Vestergaard, Christian; Werfel, Thomas; Wollenberg, Andreas; Paller, Amy S.; Reynolds, Nick J.

Published in: Journal of Allergy and Clinical Immunology

DOI (link to publication from Publisher): 10.1016/j.jaci.2018.10.033

Creative Commons License CC BY-NC-ND 4.0

Publication date: 2019

Document Version
Publisher's PDF, also known as Version of record

Link to publication from Aalborg University

Citation for published version (APA):

Eyerich, K., Brown, S. J., Perez White, B. E., Tanaka, R. J., Bissonette, R., Dhar, S., Bieber, T., Hijnen, D. J., Guttman-Yassky, E., Irvine, A., Thyssen, J. P., Vestergaard, C., Werfel, T., Wollenberg, A., Paller, A. S., & Reynolds, N. J. (2019). Human and computational models of atopic dermatitis: A review and perspectives by an expert panel of the International Eczema Council. *Journal of Allergy and Clinical Immunology*, *143*(1), 36-45. https://doi.org/10.1016/j.jaci.2018.10.033

General rights

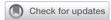
Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal -

Take down policy
If you believe that this document breaches copyright please contact us at vbn@aub.aau.dk providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from vbn.aau.dk on: December 04, 2025

Human and computational models of atopic dermatitis: A review and perspectives by an expert panel of the International Eczema Council



Newcastle upon Tyne, United Kingdom; Montreal, Quebec, Canada; Kolkata, India; Davos, Switzerland; Rotterdam, The Netherlands; New York, NY; Dublin, Ireland; and Copenhagen and Aalborg, Denmark

Atopic dermatitis (AD) is a prevalent disease worldwide and is associated with systemic comorbidities representing a significant burden on patients, their families, and society. Therapeutic options for AD remain limited, in part because of a lack of well-characterized animal models. There has been increasing interest in developing experimental approaches to study the

pathogenesis of human AD *in vivo*, *in vitro*, and *in silico* to better define pathophysiologic mechanisms and identify novel therapeutic targets and biomarkers that predict therapeutic response. This review critically appraises a range of models, including genetic mutations relevant to AD, experimental challenge of human skin *in vivo*, tissue culture models,

From athe Department of Dermatology and Allergy, Technical University of Munich; bthe Center of Allergy and Environment (ZAUM), HMGU and Technical University of Munich; cthe Skin Research Group, School of Medicine, University of Dundee and dthe Department of Dermatology, Ninewells Hospital and Medical School, Dundee; ethe Department of Dermatology and Skin Tissue Engineering Core, Feinberg School of Medicine, Northwestern University, Chicago; fthe Department of Bioengineering, Imperial College London; gInnovaderm Research, Montreal; hthe Department of Pediatric Dermatology, Institute of Child Health, Kolkata; the Department of Dermatology and Allergy, University of Bonn; the Christine Kühne-Center for Allergy Research and Education, Davos; kthe Department of Dermatology, Erasmus University Medical Center (Erasmus MC), Rotterdam; 1the Icahn School of Medicine at Mount Sinai Medical Center, New York; ^mTrinity College Dublin, National Children's Research Centre, Paediatric Dermatology Our Lady's Children's Hospital, Dublin; "the Department of Dermatology and Allergy, National Allergy Research Centre, Herlev and Gentofte Hospital, University of Copenhagen; ^othe Department of Dermatology, Aalborg University Hospital, Aalborg; ^pMedizinische Hochschule Hannover; ^qthe Department of Dermatology and Allergy, Ludwig-Maximilians-Universität Munich; rthe Departments of Dermatology and Pediatrics and the Skin Disease Research Center, Northwestern University Feinberg School of Medicine, Chicago; Spermatological Sciences, Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne; and the Department of Dermatology, Royal Victoria Infirmary, Newcastle Hospitals NHS Foundation Trust, Newcastle upon Tyne.

*These authors contributed equally to this work.

‡These authors contributed equally to this work.

K.E. is funded by an ERC grant (IMCIS, 676858) and the German Research Foundation (EY97/3-1). S.J.B. holds a Wellcome Trust Senior Research Fellowship in Clinical Science (106865/Z/15/Z). B.E.P.W. is supported by the Dermatology Foundation and the National Institutes of Health (NIH)/National Institute of Arthritis and Musculoskeletal and Skin Diseases (NIAMS; P30AR057216 and 1K01AR072773-01A1). N.J.R.'s research/laboratory is funded in part by the Newcastle National Institute of Health Research (NIHR) Biomedical Research Centre, the Newcastle NIHR Medtech and *In vitro* diagnostic Co-operative, and the Newcastle MRC/EPSRC Molecular Pathology Node. J.P.T. is funded by an unrestricted grant from the Lundbeck Foundation.

Disclosure of potential conflict of interest: K. Eyerich is funded by an ERC grant (IMCIS, 676858) and the German Research Foundation (EY97/3-2) and is a consultant and advisor/board member for Abbvie, Almirall, Berlin Chemie, Celgene, Hexal, Janssen, Leo, Lilly, Novartis, and Sanofi. S. J. Brown holds a Wellcome Trust Senior Research Fellowship in Clinical Science (106865/Z/15/Z) and reports honorarium from the British Society for Paediatric Dermatology and other support from the British Association of Dermatologists. B. E. Perez White reports grants from the Dermatology Foundation Research Career Development Award and from the National Institute of Arthritis, Musculoskeletal and Skin Diseases K01 Mentored Research Career

Development Award and P30 Skin Disease Research Center Grant. R. J. Tanaka reports grants from the Engineering and Physical Sciences Research Council (EPSRC), the Royal Society, and the British Skin Foundation. R. Bissonette is an investigator, consultant, advisory board member, and speaker for and/or receives honoraria from Aguinox Pharma, Antiobix, Asana, Astellas, Brickell Biotech, Dermavant, Dermira, Dignity Sciences, Eli Lilly, Galderma, Glenmark, GlaxoSmithKline-Stiefel, Hoffman-LaRoche Ltd, Kiniksa, Leo Pharma, Neokera, Pfizer, Regeneron, Sienna, and Vitae and is also Shareholder of Innovaderm Research. T. Bieber is a consultant for Dermavant, AbbVie, Kymab, and Glenmark and a lecturer and consultant for Sanofi, Novartis, Lilly, Pfizer, and Almirall. E. Guttman-Yassky is a consultant and/or advisory board member for and/or received grants and/or personal fees from Novartis, Pfizer, Regeneron, Asnan, Dermira, Sanofi, Eli Lilly, Asana Bioscience, Kyowa Kirin, Allergan, Escalier, AbbVie, Celgene, Gladerma, Glenmark, LEO Pharmaceuticals, Novartis, Pfizer, Regeneron, DS Biopharma, Janssen Biotech, Innovaderm, Ralexar, Novan, Dermavant, Mitsubishi Tanabe, Concert, Amgen, and DBV. J. P. Thyssen is funded by an unrestricted grant from the Lundbeck Foundation. A. Wollenberg reports personal fees and/or grants and/or nonfinancial support from Almirall, Anacor, Astellas, Beiersdorf, Bioderma, Celgene, Chugai, Galderma, GlaxoSmithKline, Hans Karrer, Leo Pharma, L'Oreal, MEDA, MSD, Novartis, Pierre Fabre, Pfizer, Regeneron, and Sanofi. A. A. Paller is an investigator or consultant with and receives honoraria from AbbVie, Anaptysbio, Eli Lilly, Galderma, Incyte, Leo, Janssen, Novartis, Sanofi-Regeneron, Amgen, Asana, Dermavant, Dermira, Galderma, Forte, Matrisys, Menlo, Morphosys/Galapagos, and Pfizer. N. J. Reynolds has received grant support through Newcastle University from AstraZeneca, Bristol-Myers Squibb, Genentech, and GlaxoSmithKline and his research/laboratory is funded in part by the Newcastle National Institute of Health Research (NIHR) Biomedical Research Centre, the Newcastle NIHR Medtech and In vitro diagnostic Co-operative, and the Newcastle MRC/EPSRC Molecular Pathology Node. The rest of the authors declare that they have no relevant conflicts of interest.

Received for publication August 15, 2018; revised October 10, 2018; accepted for publication October 30, 2018.

Available online November 7, 2018.

Corresponding authors: Kilian Eyerich, MD, PhD, Biedersteiner Strasse 29, 80802 Munich, Germany. E-mail: kilian.eyerich@tum.de. Or: Nick J. Reynolds, BSc, MB BS, MD, FRCP, Newcastle upon Tyne, NE2 4HH United Kingdom. E-mail: nick. reynolds@ncl.ac.uk.

The CrossMark symbol notifies online readers when updates have been made to the article such as errata or minor corrections
0001.6740

© 2018 The Authors. Published by Elsevier Inc. on behalf of the American Academy of Allergy, Asthma & Immunology. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

https://doi.org/10.1016/j.jaci.2018.10.033

integration of "omics" data sets, and development of predictive computational models. Although no one individual model recapitulates the complex AD pathophysiology, our review highlights insights gained into key elements of cutaneous biology, molecular pathways, and therapeutic target identification through each approach. Recent developments in computational analysis, including application of machine learning and a systems approach to data integration and predictive modeling, highlight the applicability of these methods to AD subclassification (endotyping), therapy development, and precision medicine. Such predictive modeling will highlight knowledge gaps, further inform refinement of biological models, and support new experimental and systems approaches to AD. (J Allergy Clin Immunol 2019;143:36-45.)

Key words: Atopic dermatitis, atopic eczema, endotype, human models, machine learning, mechanistic models, precision medicine, tissue culture models, skin equivalents, systems biology

Atopic dermatitis (AD; synonym atopic eczema) has a complex etiology involving multiple genetic and environmental factors.^{1,2} With its very high incidence in childhood, chronicity, devastating effect on quality of life for affected patients and their families, enormous socioeconomic costs, and limited therapeutic options to date, AD represents a major challenge. Furthermore, there is clear evidence that AD represents a systemic inflammatory disease with multiple comorbidities extending beyond the well-recognized atopic associations.³ Consequently, a number of animal models have been developed and used by investigators and the pharmaceutical industry to better understand the disease and consider new pathways to target.⁴ However, as recently reviewed, mouse models do not adequately reflect the transcriptomic and gene pathways activated in human AD skin, and the intrinsic difference between mouse and human skin represents a barrier to direct translation of findings from animals into human disease. Consequently, there has been increasing interest in experimental studies in human subjects (in part facilitated by technological and "omics" developments), cell culture models using human tissue, and use of computational or mathematical models that are developed by integrating these data.

In this review article we have used the term human AD model to define representations of the disease state and interventions that enable scientific insight into disease pathogenesis, disease course, and response to therapy. We delineate and critically appraise these AD modeling approaches that range from experimental study of human skin in vivo (including challenge studies and detailed phenotyping and investigation of patients harboring specific genetic mutations) to generation of AD-relevant models by using immunologic, genetic, and molecular methods in 2-dimensional and 3-dimensional human tissue culture to development of in silico computational models using a systems biology approach. Although by definition a reductionist approach cannot recapitulate the full spectrum of AD, these models have greatly increased our understanding of the molecular drivers of AD and provide a powerful tool for preclinical drug development and target validation. However, just as the etiology, clinical expression, and severity of AD range broadly among patients, in vitro and in silico models of AD vary widely both in how the AD phenotype is induced and how the models are evaluated. Therefore we invited members of the International Eczema Council (www.eczemacouncil.org), a group of experts in AD, Abbreviations used

ACD: Allergic contact dermatitis

AD: Atopic dermatitis
APT: Atopy patch test
FLG: Gene encoding filaggrin
LC: Langerhans cell

and associated authorities in the field to contribute to a scoping and development meeting and subsequently to evaluate and critically appraise the breadth of human AD and computational models to determine their strengths and weaknesses in how they recapitulate the pathophysiology of AD and enable therapeutics to be tested and validated.

IN VIVO MODELS OF AD

Two general approaches using human *in vivo* models have been followed to dissect the pathogenesis of AD: (1) the study of rare genetic variants with AD-like phenotypes and (2) the experimental challenge of patients with or without AD with allergens or irritants. Regarding the first approach, numerous studies have characterized genetic disorders that display skin barrier function abnormalities. Most often, these studies characterized ichthyosis vulgaris, a disease that allowed insight into the function of the epidermal differentiation gene filaggrin (*FLG*), in which mutations show the strongest association to AD development of all known genes (Fig 1).

Other studies have focused on disorders characterized by systemic inflammation³ and immunodeficiency with AD-like skin manifestations (Fig 1). One example is immunodysregulation, polyendocrinopathy, enteropathy, X-linked syndrome, which serves as a model to study how systemic imbalances in the regulatory T-cell population can drive cutaneous AD-like inflammation.⁷ In addition, the link between type 2 immunity; transcription factors, such as Janus kinase or signal transducer and activator of transcription; and high levels of IgE was investigated in immunodeficiency syndromes, such as signal transducer and activator of transcription 3 and dedicator of cytokinesis 8 hyper-IgE syndromes or combined immunodeficiency disorders.^{8,9}

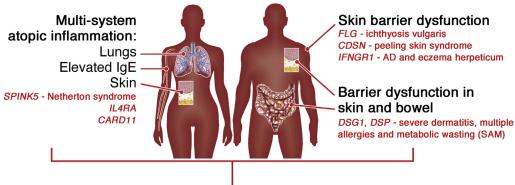
Table E1 in this article's Online Repository at www.jacionline. org lists the main genetic conditions that have provided insight into AD pathogenesis to date. Although the study of rare variants offers the opportunity to delineate distinct molecular mechanisms and control pathways of a particular phenotype and thus can be regarded as "human models of AD," a limitation of this approach is that not all observed phenomena are relevant in patients with AD, which is more complex and heterogeneous than monogenic disorders.

The second *in vivo* approach to study the pathogenesis of AD is standardized challenge with allergens or other environmental factors. The most commonly used model is the atopy patch test (APT), an epicutaneous challenge of specific allergens dissolved in vehicle, ¹⁰ which has provided insight into the temporal development of immune phenomena in patients with AD (see Table E2 in this article's Online Repository at www.jacionline. org). ¹¹ Although developed in part to define clinically relevant reactions to aeroallergens, food allergens, and autoantigens, ¹²⁻¹⁴ its validity and predictive value depend on a variety of factors

38 EYERICH ET AL

J ALLERGY CLIN IMMUNOL

JANUARY 2019



Immunodeficiency syndromes

Omenn syndrome e.g. RAG1, RAG2
Wiskott Aldrich syndrome - WAS
Hyper-IgE syndromes e.g. STAT3, DOCK8, PGM3
Immunodysregulation polyendocrinopathy enteropathy X-linked
(IPEX) e.g. FOXP3, MALT1, IL2RA, CTLA-4

FIG 1. Diagrammatic representation of "human knockout" monogenic models, providing insight into the pathomechanisms of AD. Specific genetic variants affecting the structural and/or immune functions of skin or other organs recapitulate features but not the entire phenotype of atopic inflammation and AD. *CARD11*, Caspase recruitment domain-containing protein 11; *CDSN*, corneodesmosin; *CTLA4*, cytotoxic T lymphocyte–associated protein 4; *DOCK8*, dedicator of cytokinesis 8; *DSG1*, desmoglein 1; *DSP*, desmoplakin; *FLG*, filaggrin; *FOXP3*, forkhead box protein 3; *IL2RA*, IL-2 receptor α; *IL4RA*, IL-4 receptor α; *IFNGR1*, IFN-γ receptor 1; *MALT1*, mucosa-associated lymphoid tissue lymphoma translocation protein 1; *PGM3*, phosphoglucomutase 3; *RAG1*, Recombination-activating gene 1; *RAG2*, recombination-activating gene 2; *SPINK5*, serine protease inhibitor Kazal type 5; *STAT3*, signal transducer and activator of transcription 3.

in the protocol, ¹⁵ and the APT is not used routinely in clinical practice. Experimentally, the APT has provided insight into the temporal sequence of cutaneous cellular infiltrates. Acute skin lesions show a highly reproducible T_H2-dominant inflitrate, ¹⁶ although other cell types, including T_H17 cells, are also present. ^{17,18} This T_H2 dominance is in sharp contrast to other inflammatory skin diseases, such as psoriasis. 19,20 Time-course studies have shown that additional immune cell subsets, such as T_H1 and T_H22 cells, progressively infiltrate the skin during an ongoing APT reaction, mirroring the cellular composition of acute versus chronic human AD. 17,21 The APT has also been used to characterize dendritic cells within early lesional AD skin, such as inflammatory dendritic epidermal cells. 18 Furthermore, the APT has provided insight into the interaction of microbiota and our immune system, in particular the role of bacteria-derived superantigens acting as an amplifier of the allergen-specific cutaneous response in patients with AD.²¹⁻²³ In all these experimental APT studies, the population of patients with AD were well defined, with specific inclusion and exclusion criteria (although the precise definitions of AD varied); in most studies AD, together with specific IgE to the corresponding allergen used in the APT, was an inclusion criterion.

Hapten challenge to induce classical allergic contact dermatitis (ACD) in patients with AD has also broadened our understanding of AD pathogenesis (see Table E2). Whether patients with AD have an increased risk of ACD remains controversial and might depend on whether they harbor FLG mutations, which might have allowed increased allergen penetration. However, attenuated ACD reactions have been reported in patients with AD compared with control subjects in a severity-dependent manner. ^{24,25} This might be due to the fact that haptens induce distinct immune responses, ²⁶ with fragrances mimicking the $T_{\rm H2}/T_{\rm H22}$ dominance of AD, whereas nickel, 2,4-Dinitrochlorobenzene

(DNCB), or imiquimod²⁷ induced T_H1/T_H17-skewed immune responses. Of note, patients with AD show a T_H2-skewed ACD reaction, ²⁸ and this immune deviation might account for the diminished ACD prevalence in patients with AD. A T_H2 immune reaction profile of patients with AD was also observed in an aerochallenge setup, ²⁹ as well as when challenging patients with AD with physical factors, such as hard water. ^{30,31}

All current challenge models have some limitations (see Table E2) because they only represent acute reactions, and the small areas of application cannot reproduce the intense pruritus and sleep disturbances usually present in patients with AD. Furthermore, to date, they have not stratified for genetic differences/endotypes among patients with AD. For example, comparing APTs in patients with and without *FLG* mutations might be a useful line of future investigation. Moreover, in the future, molecular profiling of lesional skin from standardized challenge models adjusted according to AD endotype can be used in early clinical studies to evaluate the potential of new drugs to improve AD.³²

IN VITRO MODELS

As shown in Table E3 in this article's Online Repository at www.jacionline.org, there are several 2-dimensional cell-culture and 3-dimensional organotypic models for AD that complement each other in addressing specific experimental questions. Although 2-dimensional cell culture models (by definition) do not duplicate the architecture of skin, they are amenable to high-throughput techniques for drug discovery and target validation (2-dimensional model section, see Table E3). Accordingly, Otsuka et al³³ used 2-dimensional cultures to screen a chemical library for compounds that enhance *FLG* transcriptional activation and mRNA expression, suggesting a

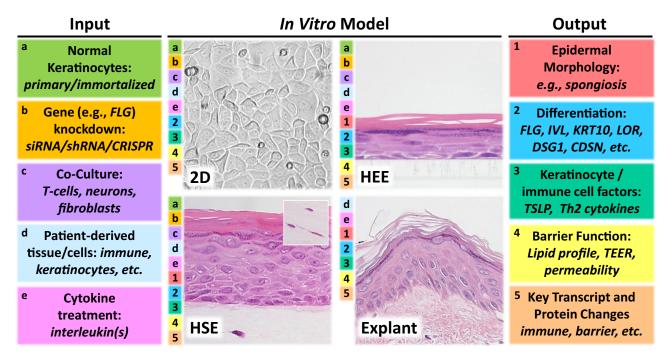


FIG 2. Human *in vitro* models of AD. *In vitro* models can be designed to address specific experimental questions based on the input materials of the cultures. Assessment of the cultures or output depends on the type of culture. *CDSN*, Corneodesmosin; *DSG1*, desmoglein 1; *FLG*, filaggrin; *HEE*, human epidermal equivalent; *HSE*, human skin equivalent (*inset*, fibroblasts in collagen); *IVL*, involucrin; *KRT10*, keratin 10; *shRNA*, short hairpin RNA; *siRNA*, small interfering RNA; *TEER*, transepithelial electrical resistance; *TSLP*, thymic stromal lymphopoietin.

potential novel therapeutic agent for AD. On the other hand, 3-dimensional models replicate the stratified squamous epithelium of epidermis but require specific expertise and are time consuming. Epidermal equivalents consist of keratinocytes without a dermal compartment, whereas skin equivalents have a dermis, such as fibroblast-embedded collagen (3-dimensional model section, see Table E3). Both 2-dimensional and 3-dimensional models are amenable to treatment with disease-relevant cytokines, gene knockdown, use of patient-derived cells, and/or coculture (Fig 2 and see Table E3).

The immune system is a major driver of AD, and in vitro immune modulation with disease-relevant cytokines, such as interleukins, can lead to AD-like phenotypes in normal primary keratinocytes³⁴ and 3-dimensional models (3-dimensional cytokine model section, see Table E3).35-41 Knockdown of filaggrin (FLG) in culture systems can provide insight into the molecular and proteomic changes associated with its loss in patients with AD, 42 and combining FLG knockdown with other perturbations, such as cytokine treatment, can be used to study the multifactorial drivers of AD. For example, Honzke et al⁴ reported that FLG knockdown exacerbated epidermal responses to IL-4 and IL-13, including increased proliferation and keratinocyte-released cytokines in 3-dimensional equivalents. Patient-derived cells for 2-dimensional and 3-dimensional culture or tissue for explant culture are limited by access and availability but might be the most relevant in terms of modeling AD. 44-47

Furthermore, patients' biopsy specimens can be a source of skin cells other than keratinocytes, allowing for coculture models. Given that multiple systems contribute to AD, coculture models that include immune cells, dermal fibroblasts, and neurons can

begin to address their interplay with keratinocytes. For example, Berroth et al⁴⁷ derived keratinocytes and fibroblasts from normal and AD skin and showed that AD-derived fibroblasts are sufficient to decrease *FLG* mRNA in normal-derived keratinocytes in 3-dimensional culture. Moreover, combining *FLG* knockdown with CD4⁺ activated T cells uncovered direct cross-talk between keratinocytes and T cells that resulted in T-cell migration within the dermal compartment toward the epidermis.⁴⁸

These studies highlight the levels of complexity that can be engineered into the 3-dimensional culture models. Three-dimensional culture systems have also been used to understand environmental influences on skin, including air pollution, UV radiation exposure, and bacterial infection. 49-51 Therefore these relevant environmental factors could be incorporated into in vitro models of AD. The 3-dimensional cultures and skin explants can also be used to assess the comparative efficacy and practical applicability of novel drug delivery systems. 52,53 Notably, despite the assorted methodologies applied in developing in vitro models of AD, there is overlap in the AD-like characteristics among the various models: most produce perturbed epidermal morphology, abnormal differentiation, and barrier dysfunction. Most often, disparities in reported phenotypes appear to stem, at least in part, from differences in the methodologies used in evaluating models (not necessarily because of the absence of the phenotype).

Although *in vitro* models might not mimic certain symptomatic and/or subjective aspects of the disease, such as pruritus and pain, they allow monitoring of changes in epidermal morphology and differentiation, gene and protein expression, lipid synthesis, and barrier function. Histologically, AD skin sections and most 3-dimensional models of AD show profound changes in the

epidermal compartment, including hypogranulosis, spongiosis, and increased cellularity because of hyperproliferation (3-dimensional model section, see Table E3). Changes in expression of genes (detected by using microarray, RNA-sequencing, or quantitative PCR) and protein (detected by using liquid chromatography—mass spectrometry, Western blotting, ELISA, or immunohistochemistry) can be used to evaluate disturbances in differentiation and immune response in 2-dimensional and 3-dimensional models. Lipid synthesis, which is required for optimal barrier function, can be monitored by expression of related enzymes or directly by using mass spectrometry. Epidermal barrier function can be monitored in 2-dimensional and 3-dimensional models, depending on the assay.

We recommend that the phenotype of any AD *in vitro* model should be extensively characterized and should include parallel analysis of epidermal morphology, differentiation status, loss or gain of key transcripts/proteins, analysis of immune components, and assessment of functional epidermal barrier parameters. Full characterization of any AD model can inform downstream evaluation of potential therapeutic agents with respect to reversing different aspects of the disease. Testing potential targets or drugs in several model types can add rigor and indicate whether a signaling pathway or protein is central to the diverse manifestations of AD.

IN SILICO COMPUTATIONAL MODELS

A core element of a systems biology approach is development of *in silico* computational models (mechanistic models) by means of integration of different types of experimental and clinical data from multiple studies, including those associated with disease conditions. *In silico* experiments (ie, computer simulations or mathematical analysis of *in silico* models) can test model-specific hypotheses, predict disease prognosis or treatment outcomes, and identify knowledge gaps, guiding future experiments and clinical trials that produce further data. This iterative process refines *in silico* models, providing holistic systems-level mechanistic insight into how perturbations (treatments or risk factors) lead to whole-organism phenotypes.

A mechanistic model describes causative interactions between the system's components involved in the phenomena of interest (eg, disease or treatment outcomes). Existing mechanistic models of AD vary widely, depending on the levels of interaction (tissue, cells, proteins, and genes) included in the model and mathematical methods used to describe the interactions.

Domínguez-Hüttinger et al⁵⁴ developed a multi-scale deterministic model that delineates interactions between the environment, skin barrier integrity, and immune activation using ordinary differential equations (Table I).⁵⁴⁻⁶¹ Two bistable "switches" are described: the first regulating the onset of AD flares and the second controlling progression to severe and persistent disease. The model predicts, for example, that genetic predisposition to barrier dysfunction (eg, *FLG* haploinsufficiency) predisposes to longer flares and more persistent disease and that prophylactic emollient use might be beneficial (Table I).

Application of optimal control theory to the hybrid mathematical model can inform the design of patient-specific optimal strategies for "proactive therapy" to prevent recurrent flares once the disease has been brought under initial control.⁵⁵

For example, this computational model supports the need for a greater topical steroid treatment dose after disease worsening and the potential need for more frequent than 2 to 3 days per week application of topical steroid treatment to maintain remission 62 in patients with FLG haploinsufficiency (Table I), presenting a readily testable stratification treatment regimen based on genotype.

Polak et al⁵⁶ developed a stochastic Petri net model that delineates genetic regulatory mechanisms responsible for immune responses in Langerhans cells (LCs; Table I). The model describes reported interactions between interferon regulatory factors, interferon regulatory factor transcription partners, and DNA sequences in a logic-based diagram. In vitro experiments validated model predictions that the ability of LCs to present a peptide is altered by cytokine milieu and that a phosphoinositide 3-kinase γ inhibitor reduces the ability of LCs to induce T_H1 responses. These smaller-scale and focused mechanistic models can describe detailed interactions that are difficult to include and validate in multiscale models. Inclusion of the detailed interactions would make the multiscale models too complex to interpret and validate because of the current lack of quantitative dynamic data measuring the variables across different scales simultaneously.

Subramanian et al⁵⁷ used a pathway model that included manually curated skin-specific pathways and relevant genes (Table I). Pathway enrichment analysis using transcriptomic data sets of patients with AD provided mechanistic insights into drug actions of topical betamethasone and pimecrolimus. The pathway model would allow *in silico* experiments once the kinetics parameters for pathways are identified to provide quantitative and dynamic predictions of disease progression and treatment outcomes.

Population pharmacokinetic and pharmacodynamic models have also been developed to describe differences and variability in pharmacologic effects observed in large clinical studies for AD treatment. The authors identified the model parameters that best fit the effects of nemolizumab and dupilumab measured in terms of AD severity score or pharmacokinetics (Table I). Population pharmacokinetic and pharmacodynamic models could help achieve mechanistic understanding of pharmacologic effects if combined with mechanistic models.

One of the challenges in developing mechanistic models is identification of the components and pathways relevant to the model-specific hypothesis to be tested. This can be achieved by using unbiased multivariate analyses of a collection of large-scale data, for example by using machine learning data analysis. Application of machine learning methods to AD-related data is relatively limited at present, but some relevant works have been already published. Thijs et al⁶⁰ developed a piecewise linear mixed model to predict AD severity scores after different treatments, and Kiiski et al⁶¹ developed a multivariate logistic regression model to predict a "good treatment response." A sufficient level of cross-validation is crucial to reduce bias and ensure the general applicability of models with predictive power beyond mere data description.

All the models presented above were developed based on the published data derived from studies in which the inclusion and exclusion criteria for AD were specified. Although the majority of studies used the Hanifin and Rajka criteria and specified further clinical (including comorbidities) and demographic details, it is clear that patients with AD present with a wide spectrum of

TABLE I. In silico computational models of AD

Model type	Scientific merits	Clinical utility	Limitations	Key features	Key findings/ predictions	References
Multiscale mechanistic model	Mechanistic understanding of system-level effects of potential triggers and processes on disease state	Identification of therapeutic targets and their mechanisms for further clinical investigation Prediction of dynamic effects of therapeutics, leading to patient stratification	Models developed based on hypothesized relationships that were previously described experimentally	A hybrid ordinary differential equation model of the dynamic interplay between skin barrier function, immune responses, and environmental stressors that determines AD pathogenesis	Preventive effects of emollients against AD progression (shown by clinical trials) Synergistic effects of environmental (eg, microbiome) and genetic (eg, FLG) risk factors on AD progression (shown by mouse experiments with ovalbumin challenge or dose-dependent effects of FLG deficiency)	Dominguez- Huttinger et al ⁵⁴
				A hybrid model of treatment effects of corticosteroids and emollients on AD pathogenesis and exploration of optimal regimens for induction of remission and maintenance of remission	Poor adherence to the suggested optimal treatment schedule leads to higher treatment doses. Application of corticosteroids for 2 consecutive days per week is optimal for the maintenance period.	Christodoulides et al ⁵⁵
Gene regulatory network model	Understanding of gene regulatory mechanisms behind disease processes	Identification of therapeutic targets and their mechanisms at the gene regulation level	Models developed based on published genetic interactions	Stochastic Petri net model of interferon regulatory factor gene regulatory network in response to <i>in vitro</i> treatment of LCs with TNF-α and TSLP	In vitro experiments validated predictions that the ability of LCs to present a peptide is altered by cytokine milieu and that PI3Kγ inhibitor reduces the ability of LCs to induce T _H 1 responses.	Polak et al ⁵⁶
Pathway models	Understanding of disease mechanisms	Identification of therapeutic targets and their mechanisms	Models developed based on published pathways	A pathway model including 35 manually curated skin-specific pathways and >2600 genes	Pathway enrichment analysis using transcriptomic data sets of 10 patients with AD treated with betamethasone valerate and pimecrolimus predicted mechanism of action of both drugs on human skin.	Subramanian et al ⁵⁷

42 EYERICH ET AL

J ALLERGY CLIN IMMUNOL

JANUARY 2019

TABLE I. (Continued)

Model type	Scientific merits	Clinical utility	Limitations	Key features	Key findings/ predictions	References
Population PK/PD models	Understanding of differences and variability in pharmacologic effects among a target population from clinical trial	Prediction of optimal dose regimen Testing effects of weight, sex, etc	Requires large clinical data to have sufficient predictive power	PK/PD model for serum nemolizumab and pruritus VAS developed from 299 patients' time course data	An appropriate flat dose regimen that is independent of body weights is used.	Saito et al ⁵⁸
	data			Two-compartment PK model for dupilumab developed from data of 197 healthy volunteers and patients with AD from 6 studies	Production rate of IL-4Ra is similar for patients with AD and healthy volunteers and does not change over time.	Kovalenko et al ⁵⁹
Machine learning predictive models	Unbiased analyses of differences between disease and nondisease (including treated) tissue/ patients and prediction of clinical outcomes (prognostic and therapeutic)	Identification of disease and therapeutic targets Findings can feed into mechanistic models	Causative mechanisms remain largely unknown Machine learning applications to atopic eczema relatively limited at present	Piecewise linear mixed models to predict EASI scores at 3 future time points from baseline biomarkers Developed from data of 150 serum biomarkers measured in 193 patients with AD	Combination of TARC, IL-22, and sIL-2R provides a good predictor for future EASI score.	Thijs et al ⁶⁰
				Multivariate logistic regression model to identify predictors of long-term response to topical maintenance treatment in AD on 169 patients	Serum total IgE (rather than the initial severity) is the most important factor predicting a good long-term treatment outcome.	Kiiski et al ⁶¹

EASI, Eczema Area and Severity Index; IL-4Ra, IL-4 receptor antagonist; PI3Kγ, phosphoinositide 3-kinase γ; PK/PD, pharmacokinetics/pharmacodynamics; sIL-2R, soluble IL-2 receptor; VAS, visual analog scale.

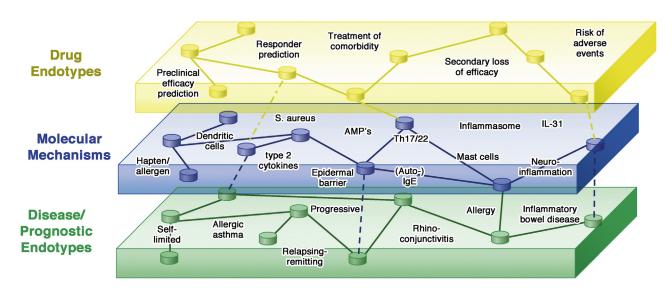
clinical and molecular features (including, for example, a greater heterogeneity in transcriptomic profile of lesional skin compared with psoriasis). ⁶³

FUTURE DEVELOPMENTS

The development of more sophisticated human and computational models of AD that integrate large-scale clinical and "omics" data offer the potential for a deeper understanding of disease endotypes, molecular mechanisms underlying key pathogenic events and clinical hallmarks of AD, as well as prediction of therapeutic outcomes, including comorbidity at the level of an individual patient. Accepting that, by definition, these models are based on a reductionist approach, they need to reflect the complexity of AD pathogenesis, including epidermal barrier dysfunction, altered penetration of chemicals and allergens, host/environment interaction, type 2 immunity, and tissue remodeling. We have illustrated in this review that the main

approaches available today are *in vitro* models, identification and characterization of human inherited syndromes resembling AD, *in vivo* challenges of patients with AD, and *in silico* models. Here we speculate how the future of AD research will likely inform the development of more refined models of AD.

Refinement is likely to depend, at least in part, on methodological advances in the field and the additional information generated by novel approaches. For example, single-cell sequencing has recently identified novel rare but important immunologic subsets, ⁶⁴ and intravital photon microscopy has enabled visualization of cell-cell communication during inflammation. ^{65,66} Application of this technology to AD is likely to inform the inclusion of distinct epithelial and immune cell types ⁶⁴ and/or genetically modified primary human cells. ⁶⁷ Furthermore, small-scale spheroid organoids can enhance high-throughput approaches in the field. ⁶⁸ Finally, we expect that a technological breakthrough in the development of 3-dimensional skin models will be facilitated by cell printers. ^{69,70}



Integration of data by systems biology and machine learning

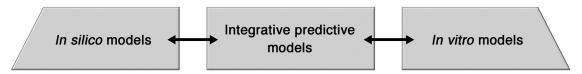


FIG 3. Interconnected multilayer networks: the future of human AD modeling. A combination of innovative *in vitro* and *in silico* models obtained by a systems biology approach and machine learning algorithms will be needed to answer clinically relevant questions, such as identification of distinct disease endotypes, elucidation of molecular pathomechanisms, or prediction of therapeutic response.

Deep neural networks are being applied as artificial intelligence tools to facilitate physician interpretation in the field of melanoma diagnostics⁷¹ and increasingly as methods to enable large data set integration. The first examples of disease classifiers⁷² and prediction of disease severity from biomarker sets^{61,73,74} have recently been published, and we expect this line of development to continue while ensuring a sufficient level of validation. We anticipate that refinement of these methods, in combination with *in silico* models, can lead to computational approaches and predictive models applied to diagnostics and therapeutic stratification.

The descriptive disease ontology of inflammatory skin diseases will need to be revised by shifting to pathogenesis-oriented structure and, in the future, by better definition of disease endotypes based on integration of multiomics data, clinical features, and clinical response to therapy in light of *in silico* models as assessed in large-scale and longitudinal cohorts. These advances are likely to inform the development of many of the current models.

However, to achieve a substantial breakthrough, we expect that different approaches will need to be combined, integrated, standardized, and performed at larger scale (Fig 3). For example, observations made in rare human disease variants or through specific challenge models in patients with AD can be validated in vitro and mapped to disease signatures in silico. Validation of functional hypotheses will increasingly depend on cross-referencing of data derived from clinical samples with outputs from in vitro models. Integration of clinical, biomarker, pharmacokinetic and pharmacodynamic (topical and/or systemic), and clinical outcome data will inform therapy

development and precision medicine. Notably, all of our models depend on how precisely a particular question is asked and the quality of the clinical input, including the clinical metadata and integration with "omics" data derived from clinical samples. Finally, advanced statistical and machine learning analysis combined with *in silico* predictive modeling will be required to integrate information throughout all described layers and data sets to elucidate underlying mechanisms (and endotypes), further highlighting the importance of data standardization and scientific networking.

We acknowledge the following International Eczema Council (IEC) associates and councilors for their contributions to the concepts outlined in this article: Lisa Beck, Rochester, NY; Carle Paul, Toulouse, France; Georg Stingl, Vienna, Austria; and Stephan Weidinger, Kiel, Germany. We thank Margaret Jung, IEC Executive Director, for organizing telephone conferences and collating responses from IEC associates and councilors.

REFERENCES

- Paternoster L, Standl M, Waage J, Baurecht H, Hotze M, Strachan DP, et al. Multi-ancestry genome-wide association study of 21,000 cases and 95,000 controls identifies new risk loci for atopic dermatitis. Nat Genet 2015;47: 1449-56.
- 2. Weidinger S, Novak N. Atopic dermatitis. Lancet 2016;387:1109-22.
- Brunner PM, Silverberg JI, Guttman-Yassky E, Paller AS, Kabashima K, Amagai M, et al. Increasing comorbidities suggest that atopic dermatitis is a systemic disorder. J Invest Dermatol 2017;137:18-25.
- Jin H, He R, Oyoshi M, Geha RS. Animal models of atopic dermatitis. J Invest Dermatol 2009;129:31-40.
- Ewald DA, Noda S, Oliva M, Litman T, Nakajima S, Li X, et al. Major differences between human atopic dermatitis and murine models, as determined by using global transcriptomic profiling. J Allergy Clin Immunol 2017;139:562-71.

- Irvine AD, McLean WH, Leung DY. Filaggrin mutations associated with skin and allergic diseases. N Engl J Med 2011;365:1315-27.
- Barzaghi F, Amaya Hernandez LC, Neven B, Ricci S, Kucuk ZY, Bleesing JJ, et al. Long-term follow-up of IPEX syndrome patients after different therapeutic strategies: an international multicenter retrospective study. J Allergy Clin Immunol 2018:141:1036-49.e5.
- Abolhassani H, Chou J, Bainter W, Platt CD, Tavassoli M, Momen T, et al. Clinical, immunologic, and genetic spectrum of 696 patients with combined immunodeficiency. J Allergy Clin Immunol 2018;141:1450-8.
- Boos AC, Hagl B, Schlesinger A, Halm BE, Ballenberger N, Pinarci M, et al. Atopic dermatitis, STAT3- and DOCK8-hyper-IgE syndromes differ in IgE-based sensitization pattern. Allergy 2014;69:943-53.
- Turjanmaa K, Darsow U, Niggemann B, Rance F, Vanto T, Werfel T. EAACI/GA2LEN position paper: present status of the atopy patch test. Allergy 2006;61:1377-84.
- Werfel T, Allam JP, Biedermann T, Eyerich K, Gilles S, Guttman-Yassky E, et al. Cellular and molecular immunologic mechanisms in patients with atopic dermatitis. J Allergy Clin Immunol 2016;138:336-49.
- Darsow U, Vieluf D, Ring J. Evaluating the relevance of aeroallergen sensitization in atopic eczema with the atopy patch test: a randomized, double-blind multicenter study. Atopy Patch Test Study Group. J Am Acad Dermatol 1999;40:187-93.
- Schmid-Grendelmeier P, Fluckiger S, Disch R, Trautmann A, Wuthrich B, Blaser K, et al. IgE-mediated and T cell-mediated autoimmunity against manganese superoxide dismutase in atopic dermatitis. J Allergy Clin Immunol 2005;115:1068-75.
- Ungar B, Correa da Rosa J, Shemer A, Czarnowicki T, Estrada YD, Fuentes-Duculan J, et al. Patch testing of food allergens promotes Th17 and Th2 responses with increased IL-33: a pilot study. Exp Dermatol 2017;26:272-5.
- Darsow U, Vieluf D, Ring J. Atopy patch test with different vehicles and allergen concentrations: an approach to standardization. J Allergy Clin Immunol 1995;95: 677-84.
- Sager N, Feldmann A, Schilling G, Kreitsch P, Neumann C. House dust mite-specific T cells in the skin of subjects with atopic dermatitis: frequency and lymphokine profile in the allergen patch test. J Allergy Clin Immunol 1992;89:801-10.
- Gittler JK, Shemer A, Suarez-Farinas M, Fuentes-Duculan J, Gulewicz KJ, Wang CQ, et al. Progressive activation of T(H)2/T(H)22 cytokines and selective epidermal proteins characterizes acute and chronic atopic dermatitis. J Allergy Clin Immunol 2012;130:1344-54.
- 18. Kerschenlohr K, Decard S, Przybilla B, Wollenberg A. Atopy patch test reactions show a rapid influx of inflammatory dendritic epidermal cells in patients with extrinsic atopic dermatitis and patients with intrinsic atopic dermatitis. J Allergy Clin Immunol 2003;111:869-74.
- Guttman-Yassky E, Nograles KE, Krueger JG. Contrasting pathogenesis of atopic dermatitis and psoriasis—part II: immune cell subsets and therapeutic concepts. J Allerey Clin Immunol 2011;127:1420-32.
- Eyerich S, Onken AT, Weidinger S, Franke A, Nasorri F, Pennino D, et al. Mutual antagonism of T cells causing psoriasis and atopic eczema. N Engl J Med 2011; 365:231-8.
- Eyerich K, Pennino D, Scarponi C, Foerster S, Nasorri F, Behrendt H, et al. IL-17 in atopic eczema: linking allergen-specific adaptive and microbial-triggered innate immune response. J Allergy Clin Immunol 2009;123:59-66.e4.
- Niebuhr M, Scharonow H, Gathmann M, Mamerow D, Werfel T. Staphylococcal exotoxins are strong inducers of IL-22: a potential role in atopic dermatitis. J Allergy Clin Immunol 2010;126:1176-83.e4.
- Langer K, Breuer K, Kapp A, Werfel T. Staphylococcus aureus-derived enterotoxins enhance house dust mite-induced patch test reactions in atopic dermatitis. Exp Dermatol 2007;16:124-9.
- Hamann CR, Hamann D, Egeberg A, Johansen JD, Silverberg J, Thyssen JP. Association between atopic dermatitis and contact sensitization: a systematic review and meta-analysis. J Am Acad Dermatol 2017;77:70-8.
- Correa da Rosa J, Malajian D, Shemer A, Rozenblit M, Dhingra N, Czarnowicki T, et al. Patients with atopic dermatitis have attenuated and distinct contact hypersensitivity responses to common allergens in skin. J Allergy Clin Immunol 2015;135:712-20
- Dhingra N, Shemer A, Correa da Rosa J, Rozenblit M, Fuentes-Duculan J, Gittler JK, et al. Molecular profiling of contact dermatitis skin identifies allergen-dependent differences in immune response. J Allergy Clin Immunol 2014;134:362-72.
- Garzorz-Stark N, Lauffer F, Krause L, Thomas J, Atenhan A, Franz R, et al. Toll-like receptor 7/8 agonists stimulate plasmacytoid dendritic cells to initiate TH17-deviated acute contact dermatitis in human subjects. J Allergy Clin Immunol 2018;141:1320-33.e11.

 Newell L, Polak ME, Perera J, Owen C, Boyd P, Pickard C, et al. Sensitization via healthy skin programs Th2 responses in individuals with atopic dermatitis. J Invest Dermatol 2013;133:2372-80.

- 29. Werfel T, Heratizadeh A, Niebuhr M, Kapp A, Roesner LM, Karch A, et al. Exacerbation of atopic dermatitis on grass pollen exposure in an environmental challenge chamber. J Allergy Clin Immunol 2015;136:96-103.e9.
- Engebretsen KA, Bager P, Wohlfahrt J, Skov L, Zachariae C, Nybo Andersen AM, et al. Prevalence of atopic dermatitis in infants by domestic water hardness and season of birth: cohort study. J Allergy Clin Immunol 2017;139: 1568-74 e1.
- Engebretsen KA, Kezic S, Jakasa I, Hedengran A, Linneberg A, Skov L, et al. Effect of atopic skin stressors on natural moisturizing factors and cytokines in healthy adult epidermis. Br J Dermatol 2018;179:679-88.
- Guttman-Yassky E, Ungar B, Malik K, Dickstein D, Suprun M, Estrada YD, et al. Molecular signatures order the potency of topically applied anti-inflammatory drugs in patients with atopic dermatitis. J Allergy Clin Immunol 2017;140: 1032-42.e13.
- Otsuka A, Doi H, Egawa G, Maekawa A, Fujita T, Nakamizo S, et al. Possible new therapeutic strategy to regulate atopic dermatitis through upregulating filaggrin expression. J Allergy Clin Immunol 2014;133:139-46, e1-10.
- Howell MD, Kim BE, Gao P, Grant AV, Boguniewicz M, DeBenedetto A, et al. Cytokine modulation of atopic dermatitis filaggrin skin expression. J Allergy Clin Immunol 2009;124:R7-12.
- 35. Kamsteeg M, Bergers M, de Boer R, Zeeuwen PL, Hato SV, Schalkwijk J, et al. Type 2 helper T-cell cytokines induce morphologic and molecular characteristics of atopic dermatitis in human skin equivalent. Am J Pathol 2011;178:2091-9.
- Yuki T, Tobiishi M, Kusaka-Kikushima A, Ota Y, Tokura Y. Impaired tight junctions in atopic dermatitis skin and in a skin-equivalent model treated with interleukin-17. PLoS One 2016;11:e0161759.
- Hanel KH, Pfaff CM, Cornelissen C, Amann PM, Marquardt Y, Czaja K, et al. Control of the physical and antimicrobial skin barrier by an IL-31-IL-1 signaling network. J Immunol 2016;196:3233-44.
- Rouaud-Tinguely P, Boudier D, Marchand L, Barruche V, Bordes S, Coppin H, et al. From the morphological to the transcriptomic characterization of a compromised three-dimensional in vitro model mimicking atopic dermatitis. Br J Dermatol 2015;173:1006-14.
- 39. De Vuyst E, Giltaire S, Lambert de Rouvroit C, Malaisse J, Mound A, Bourtembourg M, et al. Methyl-beta-cyclodextrin concurs with interleukin (IL)-4, IL-13 and IL-25 to induce alterations reminiscent of atopic dermatitis in reconstructed human epidermis. Exp Dermatol 2018;27:435-7.
- 40. Danso MO, van Drongelen V, Mulder A, van Esch J, Scott H, van Smeden J, et al. TNF-alpha and Th2 cytokines induce atopic dermatitis-like features on epidermal differentiation proteins and stratum corneum lipids in human skin equivalents. J Invest Dermatol 2014;134:1941-50.
- Nygaard U, van den Bogaard EH, Niehues H, Hvid M, Deleuran M, Johansen C, et al. The "alarmins" HMBG1 and IL-33 downregulate structural skin barrier proteins and impair epidermal growth. Acta Derm Venereol 2017;97:305-12.
- Elias MS, Long HA, Newman CF, Wilson PA, West A, McGill PJ, et al. Proteomic analysis of filaggrin deficiency identifies molecular signatures characteristic of atopic eczema. J Allergy Clin Immunol 2017;140:1299-309.
- 43. Honzke S, Wallmeyer L, Ostrowski A, Radbruch M, Mundhenk L, Schafer-Korting M, et al. Influence of Th2 cytokines on the cornified envelope, tight junction proteins, and ss-defensins in filaggrin-deficient skin equivalents. J Invest Dermatol 2016;136:631-9.
- 44. Pastore S, Fanales-Belasio E, Albanesi C, Chinni LM, Giannetti A, Girolomoni G. Granulocyte macrophage colony-stimulating factor is overproduced by keratinocytes in atopic dermatitis. Implications for sustained dendritic cell activation in the skin. J Clin Invest 1997;99:3009-17.
- van Drongelen V, Danso MO, Out JJ, Mulder A, Lavrijsen AP, Bouwstra JA, et al. Explant cultures of atopic dermatitis biopsies maintain their epidermal characteristics in vitro. Cell Tissue Res 2015;361:789-97.
- 46. Bogiatzi SI, Fernandez I, Bichet JC, Marloie-Provost MA, Volpe E, Sastre X, et al. Cutting edge: proinflammatory and Th2 cytokines synergize to induce thymic stromal lymphopoietin production by human skin keratinocytes. J Immunol 2007;178:3373-7.
- Berroth A, Kuhnl J, Kurschat N, Schwarz A, Stab F, Schwarz T, et al. Role of fibroblasts in the pathogenesis of atopic dermatitis. J Allergy Clin Immunol 2013;131:1547-54.
- Wallmeyer L, Dietert K, Sochorova M, Gruber AD, Kleuser B, Vavrova K, et al. TSLP is a direct trigger for T cell migration in filaggrin-deficient skin equivalents. Sci Rep 2017;7:774.
- Lecas S, Boursier E, Fitoussi R, Vie K, Momas I, Seta N, et al. In vitro model adapted to the study of skin ageing induced by air pollution. Toxicol Lett 2016;259:60-8.

- Maboni G, Davenport R, Sessford K, Baiker K, Jensen TK, Blanchard AM, et al. A novel 3D skin explant model to study anaerobic bacterial infection. Front Cell Infect Microbiol 2017;7:404.
- Marionnet C, Pierrard C, Lejeune F, Sok J, Thomas M, Bernerd F. Different oxidative stress response in keratinocytes and fibroblasts of reconstructed skin exposed to non extreme daily-ultraviolet radiation. PLoS One 2010;5:e12059.
- Abaci HE, Guo Z, Doucet Y, Jackow J, Christiano A. Next generation human skin constructs as advanced tools for drug development. Exp Biol Med (Maywood) 2017;242:1657-68.
- Castex-Rizzi N, Galliano MF, Aries MF, Hernandez-Pigeon H, Vaissiere C, Delga H, et al. In vitro approaches to pharmacological screening in the field of atopic dermatitis. Br J Dermatol 2014;170(suppl 1):12-8.
- 54. Dominguez-Huttinger E, Christodoulides P, Miyauchi K, Irvine AD, Okada-Hatakeyama M, Kubo M, et al. Mathematical modeling of atopic dermatitis reveals "double-switch" mechanisms underlying 4 common disease phenotypes. J Allergy Clin Immunol 2017;139:1861-72.e7.
- Christodoulides P, Hirata Y, Dominguez-Huttinger E, Danby SG, Cork MJ, Williams HC, et al. Computational design of treatment strategies for proactive therapy on atopic dermatitis using optimal control theory. Philos Trans A Math Phys Eng Sci 2017;375.
- Polak ME, Ung CY, Masapust J, Freeman TC, Ardern-Jones MR. Petri Net computational modelling of Langerhans cell Interferon Regulatory Factor Network predicts their role in T cell activation. Sci Rep 2017;7:668.
- Subramanian I, Singh VK, Jere A. Elucidating mechanistic insights into drug action for atopic dermatitis: a systems biology approach. BMC Dermatol 2018; 18:3
- Saito T, Iida S, Terao K, Kumagai Y. Dosage optimization of nemolizumab using population pharmacokinetic and pharmacokinetic-pharmacodynamic modeling and simulation. J Clin Pharmacol 2017;57:1564-72.
- 59. Kovalenko P, DiCioccio AT, Davis JD, Li M, Ardeleanu M, Graham N, et al. Exploratory population PK analysis of dupilumab, a fully human monoclonal antibody against IL-4Ralpha, in atopic dermatitis patients and normal volunteers. CPT Pharmacometrics Syst Pharmacol 2016;5:617-24.
- Thijs JL, Drylewicz J, Fiechter R, Strickland I, Sleeman MA, Herath A, et al. EASI p-EASI: utilizing a combination of serum biomarkers offers an objective measurement tool for disease severity in atopic dermatitis patients. J Allergy Clin Immunol 2017:140:1703-5.
- Kiiski V, Karlsson O, Remitz A, Reitamo S. High serum total IgE predicts poor long-term outcome in atopic dermatitis. Acta Derm Venereol 2015;95:943-7.
- Schmitt J, von Kobyletzki L, Svensson A, Apfelbacher C. Efficacy and tolerability of proactive treatment with topical corticosteroids and calcineurin

- inhibitors for atopic eczema: systematic review and meta-analysis of randomized controlled trials. Br J Dermatol 2011;164:415-28.
- Guttman-Yassky E, Krueger JG. Atopic dermatitis and psoriasis: two different immune diseases or one spectrum? Curr Opin Immunol 2017;48:68-73.
- 64. Villani AC, Satija R, Reynolds G, Sarkizova S, Shekhar K, Fletcher J, et al. Single-cell RNA-seq reveals new types of human blood dendritic cells, monocytes, and progenitors. Science 2017;356.
- Reber LL, Sibilano R, Starkl P, Roers A, Grimbaldeston MA, Tsai M, et al. Imaging protective mast cells in living mice during severe contact hypersensitivity. JCI Insight 2017;2.
- Dudeck J, Medyukhina A, Frobel J, Svensson CM, Kotrba J, Gerlach M, et al. Mast cells acquire MHCII from dendritic cells during skin inflammation. J Exp Med 2017;214:3791-811.
- 67. Niehues H, Bouwstra JA, El Ghalbzouri A, Brandner JM, Zeeuwen P, van den Bogaard EH. 3D skin models for 3R research: the potential of 3D reconstructed skin models to study skin barrier function. Exp Dermatol 2018;27:501-11.
- Fatehullah A, Tan SH, Barker N. Organoids as an in vitro model of human development and disease. Nat Cell Biol 2016;18:246-54.
- 69. Kim BS, Kwon YW, Kong JS, Park GT, Gao G, Han W, et al. 3D cell printing of in vitro stabilized skin model and in vivo pre-vascularized skin patch using tissue-specific extracellular matrix bioink: a step towards advanced skin tissue engineering. Biomaterials 2018;168:38-53.
- Pourchet LJ, Thepot A, Albouy M, Courtial EJ, Boher A, Blum LJ, et al. Human skin 3D bioprinting using scaffold-free approach. Adv Healthc Mater 2017;6.
- Esteva A, Kuprel B, Novoa RA, Ko J, Swetter SM, Blau HM, et al. Dermatologist-level classification of skin cancer with deep neural networks. Nature 2017;542:115-8.
- Quaranta M, Knapp B, Garzorz N, Mattii M, Pullabhatla V, Pennino D, et al. Intraindividual genome expression analysis reveals a specific molecular signature of psoriasis and eczema. Sci Transl Med 2014;6:244ra90.
- Ungar B, Garcet S, Gonzalez J, Dhingra N, Correa da Rosa J, Shemer A, et al. An
 integrated model of atopic dermatitis biomarkers highlights the systemic nature of
 the disease. J Invest Dermatol 2017;137:603-13.
- Krause L, Mourantchanian V, Brockow K, Theis FJ, Schmidt-Weber CB, Knapp B, et al. A computational model to predict severity of atopic eczema from 30 serum proteins. J Allergy Clin Immunol 2016;138:1207-10.e2.
- Eyerich K, Eyerich S. Immune response patterns in non-communicable inflammatory skin diseases. J Eur Acad Dermatol Venereol 2018;32:692-703.
- Paternoster L, Savenije OEM, Heron J, Evans DM, Vonk JM, Brunekreef B, et al. Identification of atopic dermatitis subgroups in children from 2 longitudinal birth cohorts. J Allergy Clin Immunol 2018;141:964-71.