

Phenolic Compounds in Salicornia spp. and Their Potential Therapeutic Effects on H1N1, HBV, HCV, and HIV

A Review

Fredsgaard, Malthé; Kaniki, Samba Eveline Kabemba; Antonopoulou, Io; Chaturvedi, Tanmay; Thomsen, Mette Hedegaard

Published in:
Molecules

DOI (link to publication from Publisher):
[10.3390/molecules28145312](https://doi.org/10.3390/molecules28145312)

Creative Commons License
CC BY 4.0

Publication date:
2023

Document Version
Publisher's PDF, also known as Version of record

[Link to publication from Aalborg University](#)

Citation for published version (APA):

Fredsgaard, M., Kaniki, S. E. K., Antonopoulou, I., Chaturvedi, T., & Thomsen, M. H. (2023). Phenolic Compounds in Salicornia spp. and Their Potential Therapeutic Effects on H1N1, HBV, HCV, and HIV: A Review. *Molecules*, 28(14), Article 5312. <https://doi.org/10.3390/molecules28145312>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal -

Take down policy

If you believe that this document breaches copyright please contact us at vbn@aub.aau.dk providing details, and we will remove access to the work immediately and investigate your claim.

Review

Phenolic Compounds in *Salicornia* spp. and Their Potential Therapeutic Effects on H1N1, HBV, HCV, and HIV: A Review

Malthe Fredsgaard ^{1,*} , Samba Evelyne Kabemba Kaniki ¹, Io Antonopoulou ² , Tanmay Chaturvedi ¹ and Mette Hedegaard Thomsen ¹

¹ AAU Energy, Aalborg University, 6700 Esbjerg, Denmark

² Biochemical Process Engineering, Division of Chemical Engineering, Department of Civil, Environmental and Natural Resources Engineering, Luleå University of Technology, SE-97187 Luleå, Sweden

* Correspondence: mfre@energy.aau.dk; Tel.: +45-42667941

Abstract: Despite public health risk mitigation measures and regulation efforts by many countries, regions, and sectors, viral outbreaks remind the world of our vulnerability to biological hazards and the importance of mitigation actions. The saltwater-tolerant plants in the *Salicornia* genus belonging to the Amaranthaceae family are widely recognized and researched as producers of clinically applicable phytochemicals. The plants in the *Salicornia* genus contain flavonoids, flavonoid glycosides, and hydroxycinnamic acids, including caffeic acid, ferulic acid, chlorogenic acid, apigenin, kaempferol, quercetin, isorhamnetin, myricetin, isoquercitrin, and myricitrin, which have all been shown to support the antiviral, virucidal, and symptom-suppressing activities. Their potential pharmacological usefulness as therapeutic medicine against viral infections has been suggested in many studies, where recent studies suggest these phenolic compounds may have pharmacological potential as therapeutic medicine against viral infections. This study reviews the antiviral effects, the mechanisms of action, and the potential as antiviral agents of the aforementioned phenolic compounds found in *Salicornia* spp. against an influenza A strain (H1N1), hepatitis B and C (HBV/HCV), and human immunodeficiency virus 1 (HIV-1), as no other literature has described these effects from the *Salicornia* genus at the time of publication. This review has the potential to have a significant societal impact by proposing the development of new antiviral nutraceuticals and pharmaceuticals derived from phenolic-rich formulations found in the edible *Salicornia* spp. These formulations could be utilized as a novel strategy by which to combat viral pandemics caused by H1N1, HBV, HCV, and HIV-1. The findings of this review indicate that isoquercitrin, myricetin, and myricitrin from *Salicornia* spp. have the potential to exhibit high efficiency in inhibiting viral infections. Myricetin exhibits inhibition of H1N1 plaque formation and reverse transcriptase, as well as integrase integration and cleavage. Isoquercitrin shows excellent neuraminidase inhibition. Myricitrin inhibits HIV-1 in infected cells. Extracts of biomass in the *Salicornia* genus could contribute to the development of more effective and efficient measures against viral infections and, ultimately, improve public health.

Keywords: *Salicornia* spp.; phenolic compounds; antiviral nutraceuticals; flavonoids; H1N1; HBV; HCV; HIV; enzymatic inhibition



Citation: Fredsgaard, M.; Kaniki, S.E.K.; Antonopoulou, I.; Chaturvedi, T.; Thomsen, M.H. Phenolic Compounds in *Salicornia* spp. and Their Potential Therapeutic Effects on H1N1, HBV, HCV, and HIV: A Review. *Molecules* **2023**, *28*, 5312. <https://doi.org/10.3390/molecules28145312>

Academic Editors: Maria Lopes, Fernando Ramos and Francesco Cacciola

Received: 22 May 2023

Revised: 7 July 2023

Accepted: 7 July 2023

Published: 10 July 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The increasing focus on the interplay between public health and bioactive compounds extracted from biomass guides researchers to shift to the most prominent groups of bioactive compounds, phenolic compounds [1]. A group of secondary metabolites known as phenolic compounds are characterized by the presence of at least one (-OH) groups attached to a benzene ring, and they often possess additional functional groups such as carboxyl, methyl, ethyl, and ketone groups. These compounds can form various structures such as homodimers, heterodimers, or polymers, including diferulic acid, chlorogenic

acid, or condensed tannins [2]. Phenolic compounds have been found to possess different in vitro and in vivo properties; e.g., anti-inflammatory, anticarcinogenic, antidiabetic, pain relieving, and neuroprotective [3–6]. Though the concentrations and the presence of phenolic compounds have been studied and documented in various fruits [7], nuts [8], and berries [9,10], the scientific interest in phenolic compounds from halophytic biomasses is new. Utilizing the residual biomass at the end of the annual cultivation cycle of edible halophytes could introduce a novel, holistic, circular economy view of biobased nutraceuticals, as compounds found in end-of-cycle halophyte fractions are known for their pharmacological properties [11–14].

Salicornia spp. are edible plants in the Amaranthaceae family and are otherwise also known as glasswort and marsh samphire, amongst others, and are known to contain a broad array of phenolic compounds [15–19]. Ferulic acid is one of the many phenolic compounds found in *Salicornia* spp., and other selected phenolic compounds found include caffeic acid, chlorogenic acid, apigenin, kaempferol, quercetin, isorhamnetin, myricetin, isoquercitrin, and myricitrin [15,20–24]. Species belonging to the *Salicornia* genus have been underutilized as medicinal herbs in recent decades, even though they have been traditionally consumed for their flavorful aerial parts in salads and seafood dishes. Additionally, they have been used as a green salt alternative, which results in a lower risk of cardiovascular diseases compared to regular salt [25–32]. The average person's daily intake of fruit and vegetables has decreased with the introduction of processed foods containing high amounts of simple sugars, starch, sodium, and fat. As fruits, vegetables, and herbs are the primary sources of phenolic compounds, this change in diet towards heavily processed food rich in macronutrients also decreased the intake, and thereby the nutraceutical effects, of phenolic compounds, thus potentially increasing the risk of lifestyle-related diseases and severe viral infections [14,33–36]. Limongelli et al. reviewed phenolic compounds from the *Salicornia* genus and their health-promoting effects. In their review, they stated that the species in the *Salicornia* genus possess antitumor, antihypertensive, antibacterial, neuroprotective, and antidiabetic activities. However, antiviral effects were not mentioned [37]. High sodium diets have been proven to induce vascular dysfunction, and in a study by Panth et al., a diet with a similar high sodium concentration provided from extracts of *Salicornia europaea* L. did not induce vascular dysfunction in rats. This effect was thought to be due to ferulic acid, which was associated with nitric oxide regulation in a study by Suzuki et al. [26,38]. *Salicornia* spp. possess many medicinal properties attributed to the phenolic content, many of which contain multiple inhibitory pathways towards enzymes related to pathogenesis, some completely unrelated to the antiviral protein inhibitory effect [39,40]. Among the many pharmacological applications of *Salicornia* spp., these are plants of great interest as traditional medicine against hepatitis, constipation, nephropathy, and diarrhea, and are proven to possess antihyperglycemic and antihyperlipidemic activities, thus making *Salicornia* spp. effective as a potential glucose- and lipid-regulating agent to subjects fed with high-glucose and high-lipid diets [27,41,42]. Of the phenolic compounds reviewed are two known and generally recognized as safe (GRAS) by The United States Food and Drug Administration (USFDA) and can be administered in concentrations of 500 mg/serving (quercetin); 150 mg/kg and 1500 mg/kg in food and chewing gum, respectively (isoquercitrin). USFDA is an administrative entity that discusses chemical stability, manufacturing procedures, metabolism, toxicology, clinical studies, and the reason for granting GRAS status to specific substances. At the time of publication, 1040 chemical compounds, extracts, fiber residues, proteins, etc., are registered as GRAS, and 68 with a pending decision by USFDA [43]. The other phenolic compounds in this review are not yet GRAS compounds but have been used by independent researchers worldwide for in vitro and in vivo experiments in less than toxic concentrations for their beneficial nutraceutical and pharmaceutical properties and have been deemed safe in amounts obtained through dietary consumption [44,45].

Oliveira-Alves et al. [28] attributed the variations in phenolic compound concentrations observed in *Salicornia ramosissima* J. Woods from Table 1 to several factors. These

factors include the maturity of the biomass, physical pretreatment methods such as milling or shredding, drying pretreatment techniques such as oven-drying or freeze-drying, and the choice of extraction methods. These factors may account for the significant differences in the measured concentrations of ferulic acid and chlorogenic acid, as presented by Oliveira-Alves et al. and Silva et al. [18,28].

Table 1. Selected phenolic compounds detected in *Salicornia* spp. reviewed for their antiviral properties. Data are presented as mean values and standard deviations measured by triplicates or more samples.

Name	Synonym	Biomass Origin	Concentration mg/kg DM	Ref.
Caffeic acid	3,4-Dihydroxycinnamic acid	<i>S. europaea</i> L.	30.4	[17]
		<i>S. europaea</i> L.	28.6	[46]
		<i>S. fruticosa</i> L.	31.0 ± 0.8	[16]
		<i>S. ramosissima</i> J. Woods	14.4 ± 0.7	[18]
Ferulic acid	3-Methoxy-4-hydroxycinnamic acid	<i>S. europaea</i> L.	58.2	[17]
		<i>S. fruticosa</i> L.	67.9 ± 1.9	[16]
		<i>S. ramosissima</i> J. Woods	28.5 ± 1.4	[18]
		<i>S. ramosissima</i> J. Woods	35.7 ± 3.4	[28]
Chlorogenic acid	3-(3,4-Dihydroxycinnamoyl)quinic acid	<i>S. europaea</i> L.	45.1	[17]
		<i>S. europaea</i> L.	391.7	[46]
		<i>S. europaea</i> L.	840.0	[37]
		<i>S. fruticosa</i> L.	85.2 ± 1.5	[16]
		<i>S. ramosissima</i> J. Woods	16.7 ± 0.8	[18]
		<i>S. ramosissima</i> J. Woods	450.8 ± 115.2	[28]
Apigenin	4',5,7-Trihydroxyflavone	<i>S. fruticosa</i> L.	<QL	[16]
		<i>S. ramosissima</i> J. Woods	4.5 ± 0.2	[18]
Kaempferol	3,4',5,7-Tetrahydroxyflavone	<i>S. fruticosa</i> L.	<QL	[16]
		<i>S. ramosissima</i> J. Woods	4.5 ± 0.2	[18]
Quercetin	3,3',4',5,7-Pentahydroxyflavone	<i>S. europaea</i> L.	8.0	[17]
		<i>S. europaea</i> L.	14.8	[47]
		<i>S. fruticosa</i> L.	15.6 ± 0.3	[16]
		<i>S. ramosissima</i> J. Woods	34.0 ± 1.7	[18]
Isorhamnetin	3,4',5,7-Tetrahydroxy-3'-methoxyflavone	<i>S. europaea</i> L.	58.9	[17]
Myricetin	3,3',4',5,5',7-Hexahydroxyflavone	<i>S. fruticosa</i> L.	137.0 ± 1.0	[16]
		<i>S. ramosissima</i> J. Woods	465.5 ± 23.3	[18]
Isoquercitrin	Quercetin 3-O-glucoside	<i>S. europaea</i> L.	10.9	[17]
		<i>S. fruticosa</i> L.	531.4 ± 9.4	[16]
		<i>S. ramosissima</i> J. Woods	453.7 ± 68.8	[28]
Myricitrin	Myricetin 3-O-rhamnoside	<i>S. fruticosa</i> L.	3314.5 ± 94.8	[16]

DM: Dry matter. <QL: Below quantification level.

Phenolic compounds are often not very soluble in water, and the scalability of conventional extraction methods using flammable organic solvents is complex, making extracting naturally derived phenolic compounds expensive. Chaturvedi et al. reviewed the extraction of phenolic compounds from lignocellulosic biomasses, hereunder *Salicornia* spp. They concluded sub-critical water extraction to be a cost-effective extraction for phenolic compounds on an industrial scale, for which the compounds have also been proven valuable for biorefinery purposes, as concluded by other authors [13,48,49]. The group of deep eutectic solvents (DES) has emerged as a promising solvent group for the extraction of phenolic compounds. Qin et al. [50] used DES to greatly increase the solubility of phenolic compounds in the solvent while still being a harmless solvent for humans and the environment. Similar effects have been seen by using an ionic liquid, closely related to DES, for the extraction of phenolic compounds from a *Salicornia europaea* matrix, recently performed by Wang and Wang, who found a significantly higher extraction efficiency using

1-butyl-3-methylimidazolium chloride salt in water than using pure water or 70% ethanol as extraction solvents [51]. As the extract obtained after any extraction of phenolics is significantly diluted, as seen in Table 1, isolation and purification are necessary to yield a pure and concentrated fraction of phenolics. Industrial-scale isolation poses a challenge, as many laboratory-scale isolations from aqueous solutions use aqueous-organic liquid-liquid extractions. However, scaling of liquid-liquid extractions can be of concern due to energy-intensive solvent recovery, flammability, and toxicity [48]. Soto et al. reviewed the flexibility and options for resin adsorption and desorption of phenolic compounds, and a common finding across all the studies reviewed was a high concentration of phenolics in the desorbate, which can be selectively desorbed from the resin by altering the desorption solvent, allowing for the separation of different classes of phenolics, hence using far less solvent than a liquid-liquid extraction [52].

While vaccines play a crucial role in preventing the spread of many viruses, antiviral drugs are used to treat viral infections once they have occurred. Currently, available antiviral drugs are limited in number and effectiveness, and the development of new antiviral agents is challenging due to the rapid evolution of viruses and the development of drug resistance by viruses [53,54]. Epidemic and pandemic outbreaks of viruses have been shown to kill millions. The influenza A (H1N1)-induced Spanish Flu (1918–1920) has various estimations regarding the death burden, but a realistic estimate was 17.4 million [55]. Furthermore, despite the attempts of worldwide influenza vaccinations, 1 billion cases are estimated yearly, resulting in between 290,000 and 650,000 deaths [53,56]. Estimates of the amount of hepatitis virus carriers globally vary greatly due to long incubation times for the virus, with ranges from 257–290 million for hepatitis B virus (HBV) and 71–300 million for hepatitis C virus (HCV) [57,58]. Of this amount, only 15 million people worldwide have been diagnosed with HCV; 439,000–1,000,000 people were under treatment for HCV in 2015 [58,59]. An estimated annual 887,000 and 385,000 deaths were attributed to HBV and HCV infections, respectively, where the majority of the deaths could have been avoided by timely detection and treatment [60]. This review will only cover HBV and HCV infections, as these account for most infections and deaths of hepatitis infections, even though the worldwide contribution towards treating chronic HCV has more than 10-fold increased from 439,000–1,000,000 to 9,400,000 from 2015 to 2021 [58]. Infections and treatment vary as infected patients might carry the viruses in inactive states for decades, unknowingly spreading the disease to sexual partners and, for women, their newborns. A vaccine has been developed for HBV but one has yet to be developed for HCV. However, once infected by HCV, an oral treatment for 8–24 weeks is usually enough to cure the patient, but combination direct-acting drug therapy can decrease the treatment period [58,59]. Chronic symptoms usually develop after many years of infection, with the fully established virus in the patient's system. Symptoms can vary from none to cirrhosis and hepatocellular carcinoma (HCC), and many patients are unaware of being carriers of the virus, risking infecting people that will show symptoms [57–59,61,62]. Antiviral resistance has been found in up to 76% of HBV patients receiving the reverse transcriptase (RT) inhibiting the nucleoside analog lamivudine for five years or more [63]. This, amongst other factors, has led to the development of more potent synthetic antiviral drugs. Such newer drugs, e.g., tenofovir and entecavir, have proven much lower antiviral resistance of <1% in 2 years of treatment in treatment-naïve patients, as at least three RT substitutions induced by HBV-DNA mutation are required to develop antiviral resistance, implying the importance of continuous research in antiviral therapeutics for early-stage infected or prevention [63,64].

Phenolic compounds have been shown to possess antiviral activity against a range of viruses, which serves as an inspiration for developing new antiviral drugs. The potential antiviral properties of phenolic compounds found in *Salicornia* spp. have been the subject of recent research. Studies have shown that *Salicornia* spp. contain a range of phenolic compounds, which have been individually proven to exhibit antiviral activity through inhibition of H1N1, HBV and HCV, and human immunodeficiency virus 1 (HIV-1), see

Table 1. The antiviral activity of phenolic compounds is due to their ability to interact and inhibit viral and host proteins and enzymes, preventing viral replication and infectivity [65–68]. Among the many research groups investigating the viral inhibitory effect of phenolic compounds, Loaiza-Cano et al., Xu et al., Besednova et al., Ono et al., Badshah et al., and Nagarajan researched the anti-viral properties of more than 100 selected phenolic compounds, and concluded that phenolic compounds with a high number of (-OH) groups and certain specific positions obtained the highest inhibitory effect. Many of the compounds with virus-inhibitory effects were found in plants known as traditional medicinal plants or foods [69–74]. Chen et al. reviewed the bioavailability of phenolic compounds and found that free phenolic compounds, e.g., quercetin, are presumed to partly degrade through dehydration reactions to hydroxybenzoic acids through gastric absorption, as the environment of the gastrointestinal tract possesses high bacterial conjugative enzyme activity [75]. By maintaining the phenolic compounds in a food matrix as bound phenolic compounds, many of the phenolic compounds can be delivered through the colon, hence avoiding degradation in the small intestine, and higher concentrations can be detected in the blood plasma. However, this adsorption method only delivers low concentrations of phenolic compounds in an untargeted manner [76,77]. The pharmacokinetics plasma concentration-time profiles for the adsorption of phenolic compounds from *Echinacea purpurea* extract were studied in a rat model by Gan et al., who found that adsorption of chlorogenic and caffeic acid through ingestion occurs in 15 and 360 min with associated disappearance half-times of 7.72 and 6.00 h, respectively [78]. This indicates that after the adsorption of phenolic compounds in rats, 10% of the initial adsorption concentration will be available after 25.6 h for chlorogenic acid and 20 h for caffeic acid. If this is also the case in humans, administering phenolic compounds towards a viral infection to obtain optimal response from the phenolic compounds should be precisely timed, and possibly administered multiple times daily during the infection period. Serra et al. investigated the distribution and accumulation of luteolin, a flavonoid closely related to quercetin, in a rat model. They found a quick distribution of luteolin into the kidneys, testicles, and heart, and no luteolin was detected in the liver, brain, thymus, and spleen, which might indicate a poor effect from luteolin to inhibit hepatitis infections in the liver. However, very low concentrations of luteolin were detected compared to the phenolic acids and their concentrations in the extract provided for ingestion, potentially indicating that the flavonoids degraded to other phenolic compounds [79]. If the research by Serra et al. also holds for other flavonoids, this could indicate the necessity for stabilizing flavonoids by encapsulation, nanoemulsion, or as a glucan conjugate for targeted delivery of flavonoids.

This study reviews phenolic compounds of *Salicornia* spp. and their potential antiviral effect on H1N1, HBV, HCV, and HIV, as these viruses continue to have serious effects on human health globally and could induce pandemic outbreaks if strains develop multi-resistance towards available treatments.

2. H1N1

The influenza virus is a part of the *Orthomyxovirus* family that contains four types; influenza A to influenza D, with only influenza A to influenza C being able to infect humans, with the H1N1 strain of influenza A being the most predominant of all. H1N1 contains the active site proteins, as seen on the H1N1 virion in Figure 1, hemagglutinin (HA), neuraminidase (NA), and M2 ion channels, which antiviral therapeutics can inhibit. Multiple subtypes of influenza have been described, including 18 different subtypes of HA and 11 subtypes of NA, classified in 162 influenza A subtypes, with a theoretical amount of 198 being possible [56,80]. NA and HA are surface glycoprotein spikes, usually in a ratio of 4 NA to 1 HA.

Liu et al. concluded that the inhibitory effects of flavonoids towards NA were highly structure-dependent and established a general order of potency: aurones > flavonoles > isoflavones > flavanones and flavanols. For the optimal inhibitory effect of NA, specific hydroxyl group placements were established at positions 4'-OH and 7-OH, the carbonyl

group on C4=O, and a set of double-bond carbon C2=C3; see Figure 2 for reference. Compounds from *Salicornia* spp. in Table 1 that have these properties are kaempferol, apigenin, myricetin, quercetin, myricitrin, and isoquercitrin. The most potent NA inhibitory effects from the study of 25 flavonoids by Liu et al. were from the flavones apigenin, luteolin, not having the 3-OH hydroxyl group, and the aurones flavonoid group [65].

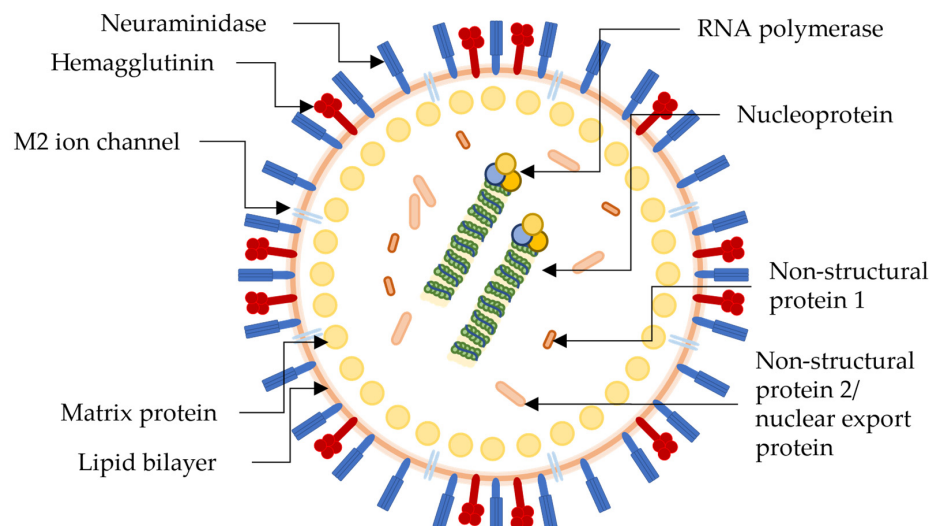


Figure 1. Pictorial representation of the structure and biology of a generic influenza virion with antiretroviral drug targets. Not specific to the H1N1 virus.

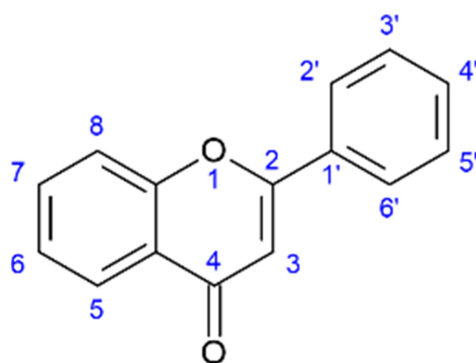


Figure 2. General structure of the flavone backbone (2-phenyl-1,4-benzopyrone) and numbered sites for attaching, amongst others, hydroxyl, methyl, and sugar groups.

Isoquercitrin is the 3-*O*-glucoside form of quercetin, making the compound more polar than the aglycone and with higher bioavailability [40]. Even though the structural differences between glycosylated flavonoids can be minor, a clear difference between the antiviral effects of the compounds when a green fluorescent protein (GFP) reporter was inserted into H1N1, and the modified virus was mixed with the flavonoids at 25 μ M and compared to a negative assay control (mock). An insignificant effect was seen between the effect from the mock and hyperoside, but isoquercitrin resulted in almost complete inhibition of H1N1-infected chicken embryos. The H1N1 inhibiting effect provided by isoquercitrin was likely the primary reason for the H1N1 inhibiting effect seen by a water extract of *Nelumbo nucifera* Gaertn. [81]. As isoquercitrin has been found in great concentrations in *S. fruticosa* L., see Table 1, a purified fraction of isoquercitrin from the *Salicornia* species could also possess high inhibition of H1N1 in chicken embryos. Isoquercitrin has been shown to exhibit the suppression of hemagglutinin (HA) and NA and inhibit cytopathic effects on both H1N1 and H3N2, making isoquercitrin a multi-acting therapeutic agent [82]. The anti-H1N1 effect of isoquercitrin has also been linked to the suppression of reactive oxy-

gen species (ROS), acidic vesicular organelle formation, and the polymerase basic protein 2 (PB2) protein [83]. Kim et al. observed a significant synergistic effect when isoquercitrin was combined with amantadine in low concentrations and did not induce resistant virus. Isoquercitrin showed a higher inhibition rate of the H1N1 virus than (-)-epigallocatechin gallate and resveratrol, which have previously been known to have a high inhibition rate of the H1N1 virus when compared to other antiviral phenolic compounds [54].

During the past few years, ferulic acid has been determined to play an essential role as a nutraceutical and against viral infections, including H1N1 with an IC_{50} inhibition at 140 μ M towards NA [84]. In addition to low cytotoxicity, ferulic acid and caffeic acid show utility for regulating the activity of the immune system [85,86]. Sodium ferulate activated toll-like receptors (TLR) 7 and 9, enhancing survival in H1N1-infected mice, contributing to antiviral defense [87]. This effect could be secondary to the induction of the heme oxygenase 1 gene (HMOX1) and possibly reflect ferulic acid's additional benefits. TLR7 and TLR9 are proteins responsible for detecting viral nucleic acids, initiating swift signaling pathways that play a crucial role in generating interferons, thereby enhancing the body's antiviral defenses [87]. Another potential impact of ferulic acid is its inhibiting effect on functions such as virus replication. During the past few years, ferulic acid has been determined to play an essential role against viral infections, including H1N1, murine coronavirus (MHV-A59), and human rhinovirus 14 (HRV-14), as a component of phenolic extract from plants with exceptional and identified action [84,88–94]. Ferulic acid is also a cytochrome P450 1A2 enzyme (CYP1A2) inhibitor, positively increasing its half-life and averting serious drug interactions [95]. CYP1A2 is involved in the metabolism of xenobiotics in the body involved in drug metabolism. Feruloylated oligosaccharides (FOs) are nutraceuticals derived from arabinoxylans, with ferulic acid esterified to L-arabinofuranosyl side chain oligosaccharides. FOs possess the physiological functions of both ferulic acid and oligosaccharides. Unlike pure oligosaccharides, FOs still release ferulic acid after fermentation by human gut microorganisms, which can introduce prebiotic effects in the gut microbiota [96]. Ferulic acid released in the colon can be absorbed in the body, exerting various physiological functions as free ferulic acid. The anti-inflammatory impact of antioxidant nutraceuticals, such as phenolics from *Salicornia* spp., might also quell the excessive inflammatory reaction within lung parenchyma evoked by viral H1N1 [93,97]. McCarty et al. suggested that a very low daily dosage ranging from 500 to 1000 mg of ferulic acid could aid in the control of H1N1 by decreasing infection response by suppressing viral spread and dampening proinflammatory signaling, which promotes an influx of inflammatory cells [98].

Chlorogenic acids are a group of esters formed between caffeic and quinic acids and contain chlorogenic acid (ChA), neochlorogenic acid (neo-ChA), and cryptochlorogenic acid (crypto-ChA), amongst others. These three compounds are closely related isomers, only differentiated by the orientation of the quinic acid group. ChAs have been recognized as an effective NA inhibitor by Ding et al., who proposed it as a nutraceutical that can decrease the time of the infection and be safely administered through the diet in low concentrations; e.g., by ingesting wild lowbush blueberries [99]. Ding et al. proved that the synthetic medicine oseltamivir at 2 μ M was less inhibiting on H1N1 than 100 μ M ChA, and the cellular toxicity of ChA was 0% at levels below 200 μ M [99].

Dayem et al. investigated the in vivo pharmaceutical effect of five flavonoids and their effect on H1N1-infected mice. Trials were set up with the administration of flavonoids as pre-treatment before virus inoculation, co-treatment with simultaneous viral inoculation and flavonoid administration, and post-treatment flavonoid administration after virus inoculation [100]. It was found that isorhamnetin suppressed the covalent binding of lipids to peptides and the lipidation of microtubule-associated protein 1 light chain 3 beta (MAP1LC3B), hence decreasing virus-induced overexpression of autophagy-related genes of healthy cells (autophagy-related gene proteins 5 and 7 (atg-5 and atg-7)) protecting the infected host. Isorhamnetin was proven to inhibit both NA and HA at a concentration of 50 μ M, indicating the potential use of isorhamnetin as an anti-H1N1 medication [100].

As seen in Table 2, many of the phenolic compounds selected for the review have been tested for various inhibition mechanisms *in silico* and *in vitro*. Typical for many of the selected phenolic compounds is the ability to inhibit the strain A/PR/8/34 (H1N1) in an infected MDCK cell line, and the NAI of that strain has also been investigated in multiple studies. From this, the inhibition efficiency of the compounds, tested on the MDCK cell line with A/PR/8/34 (H1N1), was shown to be isoquercitrin > caffeic acid > quercetin, and the NAI of the selected compounds on A/PR/8/34 (H1N1) was shown to be apigenin > kaempferol > quercetin > myricetin. Molecular docking simulations also proved the effect on NAI with similar results to the *in vitro* NAI results, showing the most potent inhibitors to be isoquercitrin > ferulic acid > kaempferol > quercetin. Furthermore, ChA is seen to possess good NAI and could, together with isoquercitrin, be viable options for NAI.

Table 2. Selected phenolic compounds detected in *Salicornia* spp. reviewed for their H1N1 inhibitory properties. Data are presented as mean values and standard deviations measured by triplicates.

Compound	H1N1 Inhibition Target	Inhibition Activity	Ref.
Caffeic acid	A/PR/8/34 (H1N1) infected MDCK	IC ₅₀ inhibition: 81.6 ± 8.3 µM	[101]
	A/Osaka/2024/09 (H1N1) infected MDCK	IC ₅₀ inhibition: 98.8 µM	[101]
	A/Osaka/71/11 (H1N1) infected MDCK	IC ₅₀ inhibition: 97.1 µM	[101]
Ferulic acid	NA	Docking energy: −7.1 kcal/mol	[84]
	A/Malaysia/Muar/33/09 NA (H1N1) inhibition (NAI) <i>in vitro</i> assay	IC ₅₀ inhibition: 140 µM	[84]
	EC ₅₀ of A/Malaysia/Muar/33/09 (H1N1) in infected MDCK cells	EC ₅₀ inhibition: 1.32 ± 0.08 µM	[84]
Chlorogenic acid	EC ₅₀ of A/PR/8/34 (H1N1) in infected MDCK cells	EC ₅₀ inhibition: 44.87 µM	[99]
	NAI by fluorescence-based <i>in vitro</i> assay	IC ₅₀ inhibition: 22.13 ± 1.07 µM	[99]
Apigenin	EC ₅₀ of A/PR/8/34 (H1N1) in infected MDCK cells	EC ₅₀ inhibition: 56.7 ± 11.1 µM	[102]
	EC ₅₀ of A/Toyama/129/11 (H1N1) in infected MDCK cells	EC ₅₀ inhibition: 65.9 ± 32.2 µM	[102]
	EC ₅₀ of A/Toyama/26/11 (H1N1) in infected MDCK cells	EC ₅₀ inhibition: 30.0 ± 17.4 µM	[102]
Kaempferol	A/PR/8/34 (H1N1) NAI <i>in vitro</i> assay	IC ₅₀ inhibition: 31.6 ± 0.9 µM	[65]
	NA	Docking energy: −6.8 kcal/mol	[103]
	NAI by RT-PCR <i>in vitro</i> assay	Significant inhibition at 50 µM	[100]
Quercetin	A/PR/8/34 (H1N1) NAI <i>in vitro</i> assay	IC ₅₀ inhibition: 58.6 ± 0.6 µM	[65]
	A/PR/8/34 (H1N1) infected MDCK	IC ₅₀ inhibition: 274.6 ± 3.3 µM	[101]
	A/PR/8/34 (H1N1) plaque formation inhibition <i>in vitro</i> assay	IC ₅₀ inhibition: 1.40 µM	[104]
Isorhamnetin	NAI by RT-PCR <i>in vitro</i> assay	Significant inhibition at 50 µM	[100]
	NA	Docking energy: −6.8 kcal/mol	[103]
	A/PR/8/34 (H1N1) NAI <i>in vitro</i> assay	IC ₅₀ inhibition: 58.4 ± 3.8 µM	[65]
Myricetin	HI by RT-PCR <i>in vitro</i> assay	Significant inhibition at 50 µM	[100]
	NAI by RT-PCR <i>in vitro</i> assay	Significant inhibition at 50 µM	[100]
	A/PR/8/34 (H1N1) NAI <i>in vitro</i> assay	IC ₅₀ inhibition: 82.6 ± 8.9 µM	[65]
Isoquercitrin	A/PR/8/34 (H1N1) plaque formation inhibition <i>in vitro</i> assay	IC ₅₀ inhibition: 0.90 µM	[104]
	HI RAW 264.7 cell line <i>in vitro</i> assay	Complete inhibition: 5 µM	[82]
	NAI by fluorescence-based <i>in vitro</i> assay	IC ₅₀ inhibition: 37.1 ± 0.6 µM	[82]
Myricitrin	PB2	Docking energy: −8.0 kcal/mol	[83]
	NA	Docking energy: −8.6 kcal/mol	[105]
	A/PR/8/34 (H1N1) infected MDCK	IC ₅₀ inhibition: 10.6 ± 0.4 µM	[83]
Myricitrin	A/WS/33 (H1N1) infected MDCK	IC ₅₀ inhibition: 21.4 ± 2.4 µM	[83]
	A/PR/8/34 (H1N1) infected MDCK	255.6% increase in cell viability at 224.0 µM	[106]

MDCK: Madin-Darby Canine Kidney cells, IC₅₀: Half maximal inhibitory concentration, EC₅₀: Half maximal effective concentration, HI: Hemagglutination inhibition, NA: Neuraminidase, NAI: Neuraminidase inhibition, RT-PCR: Reverse transcription polymerase chain reaction, PB2: polymerase basic protein 2.

3. Hepatitis

Many phenolic compounds have been found to possess direct hepatoprotective properties and inhibit HBV and HCV reproduction; e.g., ellagic acid, isorhamnetin, quercetin, and myricitrin [66,67,107,108]. These hepatoprotective properties might not eliminate the root cause of the problem, the viral infection, but intentional administration of such compounds in elevated concentrations in the diet might offset, delay, or reduce the symptoms related to HBV in a non-vaccinated individual by acting as an agonist, antagonist, or synergistic by inducing or inhibiting protein and enzymatic multi-pathways [107,109]. On the other hand, quercetin has been found to act as an antagonist by inhibiting the multi-enzymatic activity non-structural protein 3 (NS3) protease, NS5B, and envelope glycoprotein 2 protein, hence stopping HCV replication. The flavones closely related to quercetin, luteolin and apigenin, showed similar binding energy through in silico molecular docking results of NS5B compared to the synthetic NS5B-inhibiting drug sofosbuvir [67,68,110]. Although sofosbuvir is a second-generation direct-acting antiviral drug with fewer side effects, it is still known to induce a series of side effects, e.g., headaches, fatigue, arthralgias, diarrhea, nausea, and anemia, in patients for whom it is administered. A drug substitution using a naturally occurring GRAS NS5B-inhibiting nutraceutical, such as quercetin, could eliminate the drug-related side effects and generally increase the wellbeing of the patient, as the majority of the compounds are not only direct-acting but are, in general, beneficial for the entire body [111].

Quercetin and myricitrin have been found to inhibit the HBV surface and e antigens, HBsAg and HBeAg; see Figure 3 [16,66]. Quercetin and myricitrin were isolated from *Guiera senegalensis* J.F. Gmel leaves by 96% ethanolic 72-h maceration, followed by rotary evaporation of the solvent, and were sequentially partitioned using hexane, chloroform, and n-butanol. The lipophobic n-butanol fraction was fractionated using normal phase column chromatography into eight sub-fractions. Secondary column chromatography was performed, and the pure compounds were obtained and verified using 2D NMR spectral analysis by ¹H-NMR and ¹³C-NMR. The isolated compounds, quercetin and myricitrin, were analyzed in silico by molecular docking, and both compounds showed good docking scores towards all analyzed proteins, RT, Na⁺/Taurocholate Cotransporting Polypeptide (NTCP), and HBcAg. NTCP, a protein membrane transporter in healthy liver cells, has also been proven to be a receptor for HBV entry, and therapeutics can act as host receptor restrictors and block HBV entry [112]. Quercetin and myricitrin were tested by a human hepatoblastoma cell line cytotoxicity assay and determined to be safe in the liver up to 165 µM and 107 µM, respectively, and these concentrations were therefore used in the HBsAg and HBeAg enzyme bioassays. Quercetin exhibited excellent HBsAg inhibition and HBeAg downregulation of 60% and 62%, respectively, compared to myricitrin by 44% and 35%, respectively. The effect of quercetin was close to that of the HBeAg-targeting synthetic lamivudine (69% at 2 µM) [66]. The aglycone of myricitrin, myricetin, also shows anti-hepatitis and hepatoprotective properties in acute hepatitis and more severe fulminant hepatitis. Myricetin was shown to exert anti-programmed cell death as antiapoptosis of healthy liver cells, together with the antioxidant and anti-inflammatory effects, further potentially decreasing the risk of symptoms. The antiapoptosis effect was explained by the regulation of the cell metabolism enzymes cysteine-dependent aspartate-directed proteases 3 (caspase-3), caspase-9, protein p53, and the inhibition of nuclear factor-kappa B (NF-κB) activation [113]. Caspase-3 and caspase-9 are responsible for executing and initiating apoptosis, respectively, for healthy cells, and regulation hereof is essential for killing infected cells and avoiding or postponing severe hepatitis symptoms, hence ensuring natural cell metabolism [114]. During an HBV infection, the protein responsible for caspase-3 regulation is the virus-encoded hepatitis B virus protein X (HBx), produced by the host cell upon infection as a gene product [115]. HBx is, together with other proteins, responsible for activating the transcription of viral genes and HCC, and can induce or block regular metabolism apoptosis, and was shown to do so through caspase-3 inhibition [116,117]. HCC is a common type of primary liver cancer that can, amongst other things, be caused by

the down-expression or inactivation of phosphatase and tensin homolog (PTEN), a tumor suppressor gene. HCC caused by HBV is one of the most common cancer variants, and the mortality rate is projected to increase by 40% from 2019 to 2030 [118,119]. Multiple phenolic compounds, e.g., gossypetin and naringenin, have been shown to attenuate the metabolic changes directly caused by the HBx protein [120]. This decreases apoptosis, hepatitis-related hepatic steatosis, HCC, and cirrhosis. The antiapoptosis effect, as seen by myricetin, might be expected to be similar, as some of the compounds responsible for the inhibition are close isomers, even though it has not yet been described [107,120].

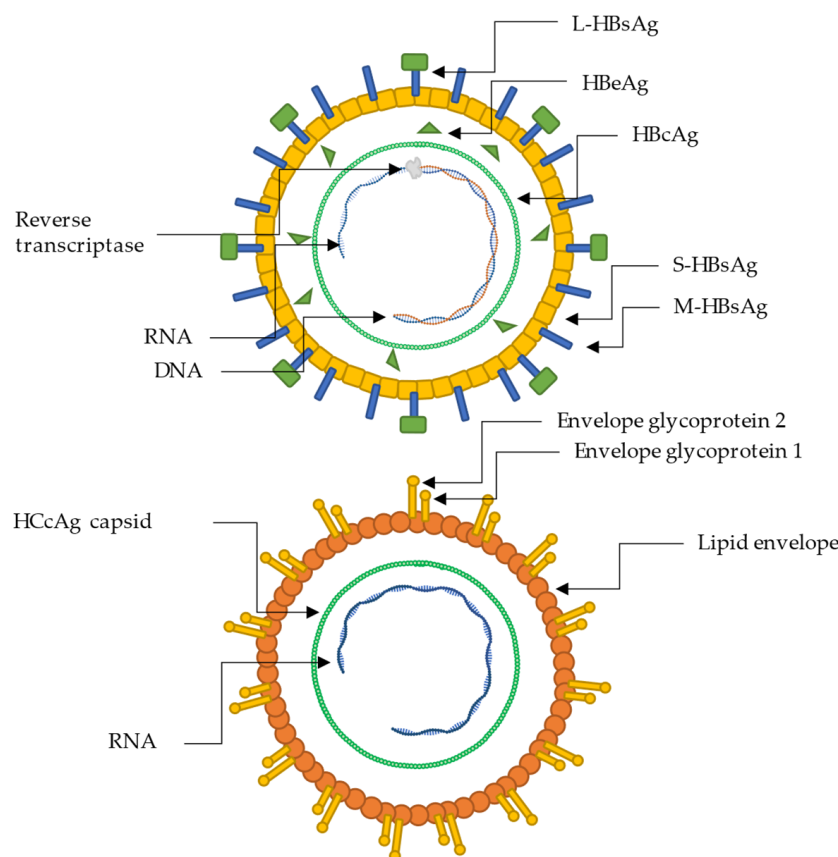


Figure 3. Pictorial representation of the structure and biology of hepatitis B and C virions. The upper virion represents hepatitis B, and the lower virion represents hepatitis C.

Zhang et al. investigated the inhibitory effect of phenolic compounds on proteins and enzymes involved in HBV replication, inflammatory response, and interaction with other proteins and enzymes [121]. Zhang et al. introduced a target network of 13 potential targets for treating HBV, and among these were the signal transducer and activator of transcription 3 (STAT3) and euchromatic histone-lysine N-methyltransferase 2 (EHMT2). These two proteins are responsible for taking place in the virus replication of HBV and down-regulating the host's immune response to the virus. ChA was seen, through molecular docking simulations, to partly inhibit both STAT3 and EHMT2, whereas caffeic acid partly inhibited EHMT2, ensuring optimal host immune defense upon infection [121].

With hepatitis viruses being necroinflammatory and hepatotropic, the anti-inflammatory effect of nutraceuticals on the liver's vital functions in the late stage of the infection can be highly beneficial to attenuate the virus-induced inflammatory response [122,123]. Proinflammatory enzymes and cytokines, such as interleukin-1 beta (IL-1 β), inducible nitric oxide synthase (iNOS), and cyclooxygenase-2 (COX-2), are produced by HCV gene expression upon infection, and the suppression of these specific enzymes and cytokines are found from phenolic compounds found in green tea, many of which are also present in the plants of the *Salicornia* genus [123]. Ma et al. investigated the protective effect of quercetin

against CCl₄-induced inflammation by significantly decreasing the production of IL-1 β , iNOS, and COX-2 [124]. The anti-inflammatory response from quercetin by Ma et al. was achieved by inhibiting TLR2, TLR4, and NF- κ B. TLR2 and TLR4 have both been found during infection by HCV and HCV/HIV-1 co-infection, indicating that the inhibitory effect of quercetin towards TLR2 and TLR4 could also apply during viral infections, lowering the inflammatory response of the patient [124,125].

Both HBV and HCV can trigger ROS production in the mitochondria of the infected hepatocytes of the host cells, hence causing mitochondrial dysfunction. Neoplastic cell transformation of the said host cell can cause oxidative stress by increased ROS levels, possibly due to the interference with the host cell's DNA repair systems for the removal of oxidized DNA, increasing the risk of HCC. ROS also promotes proinflammatory cytokine production, increasing the liver's inflammatory response [126]. When infected with HBV, the intracellular produced HBx will identify the mitochondria and increase ROS levels towards 10,000-fold of normal levels, which can cause 20 types of DNA mutations [127–129]. Administering phenolics found in *Salicornia* spp., natural antioxidants, and hence ROS scavengers, to HBV and HCV patients, intracellular hepatic ROS production can be limited and delay HBV- and HCV-related liver carcinogenesis, HCC, and hepatic fibrogenesis [130,131].

As seen in Table 3, the literature available shows sparse data regarding the interactions between HBV or HCV and the phenolic compounds selected for this review from *Salicornia* spp. Some molecular docking studies, which tend to be the first indication of potentially new drug candidates, have been performed on the proteins and enzymes responsible for viral entry and replication and the phenolic compounds of interest. From this data, the most promising compounds for HBcAg inhibition are kaempferol > quercetin > myricitrin. The most promising HBV-RT inhibition compounds are kaempferol > quercetin > myricitrin. In addition to these inhibitions, ChA shows good HBV-DNA inhibitory effects at low concentrations compared to caffeic acid, but poorer inhibition of HBsAg and HBeAg, two crucial targets for anti-HBV therapy.

Table 3. Selected phenolic compounds detected in *Salicornia* spp. reviewed for their HBV/HCV inhibitory properties. Data are presented as mean values and standard deviations measured by triplicates.

Name	HBV/HCV Inhibition Target	Inhibition Activity	Ref.
Caffeic acid	HCV-1 infected Huh7.5.1 cells	IC ₅₀ inhibition: 100 \pm 20 μ M	[132]
	EHMT2	Docking energy: −5.9 kcal/mol	[121]
	HBV-DNA	IC ₅₀ inhibition: 3.9 \pm 1.1 μ M	[133]
	HBsAg in vitro assay in infected HepG2.2.2.15 cells	IC ₅₀ inhibition: 12.7 \pm 9.9 μ M	[133]
	HBeAg in vitro assay in infected HepG2.2.2.15 cells	IC ₅₀ inhibition: 109.3 \pm 56.0 μ M	[133]
Ferulic acid	No data		
Chlorogenic acid	EHMT2	Docking energy: −7.8 kcal/mol	[121]
	STAT3	Docking energy: −7.2 kcal/mol	[121]
	HBV-DNA	IC ₅₀ inhibition: 1.2 \pm 0.4 μ M	[133]
	HBsAg in vitro assay in infected HepG2.2.2.15 cells	IC ₅₀ inhibition: 241.5 \pm 198.2 μ M	[133]
	HBeAg in vitro assay in infected HepG2.2.2.15 cells	IC ₅₀ inhibition: >1000 μ M	[133]
Apigenin	EC ₅₀ of HBsAg	EC ₅₀ inhibition: 26.3 \pm 5.6 μ M	[134]
	EC ₅₀ of HBeAg	EC ₅₀ inhibition: 47.4 \pm 3.3 μ M	[134]
Kaempferol	HBV-RT	Docking energy: −9.0 kcal/mol	[66]
	HBcAg	Docking energy: −9.1 kcal/mol	[66]

Table 3. Cont.

Name	HBV/HCV Inhibition Target	Inhibition Activity	Ref.
Quercetin	HBV-RT	Docking energy: −8.3 kcal/mol	[66]
	HBcAg	Docking energy: −8.7 kcal/mol	[66]
	NTCP	Docking energy: −5.4 kcal/mol	[66]
	HBsAg in vitro assay in infected HepG2.2.2.15 cells	60% inhibition	[66]
	HBeAg in vitro assay in infected HepG2.2.2.15 cells	62% inhibition	[66]
Isorhamnetin	No data		
Myricetin	Infected fulminant hepatitis mice	66% increased survival rate	[113]
Isoquercitrin	No data		
Myricitrin	HBV-RT	Docking energy: −7.7 kcal/mol	[66]
	HBcAg	Docking energy: −7.1 kcal/mol	[66]
	NTCP	Docking energy: −6.7 kcal/mol	[66]
	HBsAg in vitro assay in infected HepG2.2.2.15 cells	44% inhibition	[66]
	HBeAg in vitro assay in infected HepG2.2.2.15 cells	35% inhibition	[66]

IC₅₀: Half maximal inhibitory concentration, EC₅₀: Half maximal effective concentration, EHMT2: Euchromatic histone-lysine *N*-methyltransferase 2, HBsAg: Hepatitis B surface antigen, HBeAg: Hepatitis B e antigen, STAT3: Signal Transducer and Activator of Transcription 3, RT: Reverse transcriptase, NTCP: Sodium Taurocholate Cotransporting Polypeptide.

4. HIV

HIV-1 is a lentivirus the virion of which is a highly complex structure enveloped by a lipid bilayer with two glycoproteins attached (GP41 and GP120) and contains a multitude of proteins including protease (PR), integrase (IN) and RT, see Figure 4, all being actively targeted for anti-HIV-1 mechanisms by therapeutic antagonists. During virus replication and infection, a polyprotein consisting of RT, PR, and IN is split by the PR. The single-stranded RNA genome of the virus then relies on the RT and IN to catalyze the viral DNA integration into the cluster of differentiation 4 (CD4) host cells. After integration, GP41 and GP120 progressively mature, resulting in a stronger viral envelope over time. As GP120 and GP41 are glycoproteins, they have been proven to be inhibited by glucosidase through cleavage [135]. HIV-1 antiretroviral therapy often targets RT, PR, or glycoproteins using pharmacophore compatibility, such as protein-ligand docking simulations [135,136]. Polyhydroxylated compounds such as caffeic acid, myricitrin, quercetin, or kaempferol have been reported as natural pharmacophores for HIV-1 [69,89,136]. IN's 3'-P central core domain contains a metal-binding active site with three amino acid residues specific for Mn and Mg cations. During infection, the IN-DNA complex is formed and later stabilized by IN. The high affinity of viral polyproteins against polyhydroxylated phenolic compounds reveals their ability to compete with various co-factors and form protein-ligand complexes [136,137].

A study by Bailly et al. found that caffeic acid interacts with the viral envelope GP120, which is essential for the virus to bind and enter the host cell [138]. GP120 binds and fuses with the T lymphocytes using its variable regions V2, V3, and V4. These glycoprotein regions form loops to neutralize the antibodies and are crucial in the virus-host interaction [139]. Repeated passages of the virus in the presence of caffeic acid resulted in the inhibition of the V2-V4 regions of GP120. Caffeic acid-induced inhibition can alter the virus's ability to bind and penetrate past the host cell receptors [138]. The HIV-1 IN has the ability to perform the G140S mutation, in which glycine (G) is replaced with serine (S) at 140 base pair. This effect was found to be inhibited by caffeic acid, and hence the stops the mutation from happening. The mutation gives antiviral resistance against pharmaceutical IN inhibitors, creating a negative feedback loop for the infected patient [140,141]. However, the catechol group of caffeic acid makes it an efficient cleavage inhibitor for IN despite

the mutation [137,142]. The results suggest that caffeic acid and its derivatives could be promising candidates for developing anti-HIV-1 therapeutic agents.

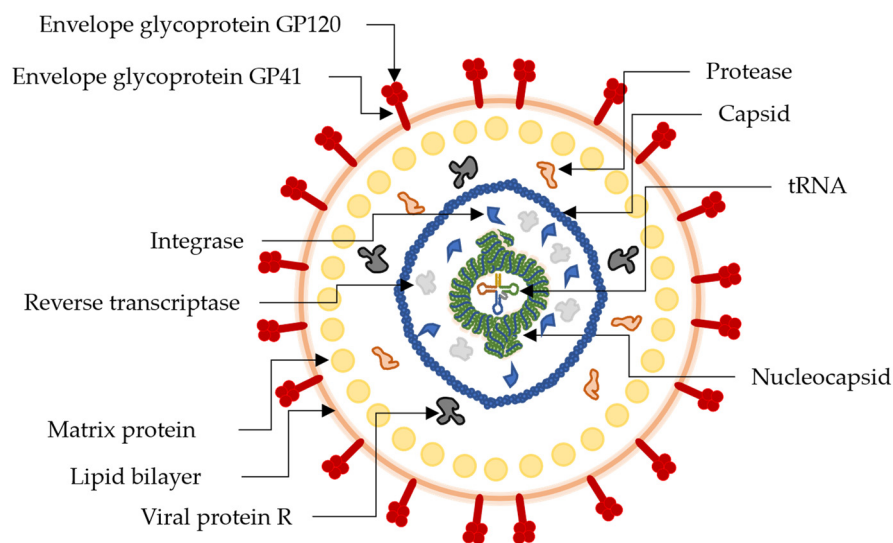


Figure 4. Pictorial representation of the structure and biology of an HIV-1 virion with antiretroviral drug targets.

In a study by Yang et al., leukaemia cells, C11 Jurkat, were infected with the HIV-1 provirus flanked and controlled by the long terminal repeat (LTR) regions [143]. The aim was to quantify the provirus transcription with a GFP as a selection marker. After 72 h of constant exposure to quercetin at 20 μM , the HIV-1 transcription in latently infected cells was increased to 14.5%. A positive correlation between the concentration of quercetin and the HIV-1 expression was observed. The expression of the provirus was observed by the presence of fluorescence from the GFP in fluorescence microscopy. GFP-positive cells increased over time to 14.6% until 72 h and with increased concentration of quercetin until 20 μM [144]. In the attempt to understand which transcription factor activates the provirus expression, the transcription factor NF- κB was silenced before inducing transcription. The induction of HIV-1 LTR ($\Delta\kappa\text{B}$) with the tumor necrosis factor- α (TNF- α) failed to show any HIV-1 expression. This finding indicates that quercetin uses the NF- κB signaling pathway to activate the viral transcription. Quercetin's ability to activate the expression of retroviral genes in latent cells is extremely useful in the development of therapy against HIV-1, as latency of HIV-1-infected cells continue to be a risk for affected patients. Therefore, it is valuable to reactivate the dormant cells with a safe natural agent such as quercetin. It could make an effective supplement for co-treatment with a High Active Antiviral Therapy (HAART) drug in order to reactivate the latent infected cells and introduce the HAART drug, in a 'shock-and-kill' strategy [144]. Neither closely related isomers of quercetin or quercetin derivatives saw high inhibition of HIV-1, and if the 3' hydroxyl group was placed on the 2' position, yielding the isomer morin, the inhibitory effect was not present [69]. On the other hand, adding a hydroxyl group on the 5' position, yielding myricetin, an increased inhibitory effect against RT was observed [69,135]. The mechanism of this flavonoid relies on its ability to compete with the template primer of RT [145]. Myricetin was found by Fesen et al. to possess high IN cleavage efficiency at 67.8%. Not only was myricetin found to be a good RT inhibitor, but also a good inhibitor of certain ATP-dependent enzymes, such as the plasma membrane Ca^{2+} pump, and could therefore increase the potential for pharmacological applications [146]. Engineered TZM-bl cells sampled from the uterine epithelial tissue were genetically manipulated to express the CD4 co-receptor, were exposed to myricetin and showed >87% inhibition of HIV-1 at a concentration of 100 μM . Those findings were supported in another study by Ortega et al., where myricetin showed HIV-1 inhibition of >80% with an IC_{50} of 7.6 μM [147].

This proves isolated myricetin from *Salicornia* spp. to be a compatible flavonoid for developing anti-HIV-1 topical microbicides [147].

Apigenin showed inhibition of HIV-1 expression in contaminated H9 cells from the T cell line [147,148]. Similar to quercetin, apigenin interacts with the transcription factor NF- κ B at a concentration as low as 10 μ M [148]. This is crucial to regulate viral transcription and replication by binding to the HIV-1 promoter region in the host cell's DNA. Apigenin competes with NF- κ B to stop the viral transcription while keeping relatively low toxicity.

As seen from Table 4, isorhamnetin, myricetin, and quercetin show promising preliminary results and should be further investigated for other anti-HIV-1 properties in clinical trials through extraction and isolation from *Salicornia* spp. to investigate their safety, efficiency, and potential widespread use in treating HIV-1. These three phenolic compounds, administered as single compounds or in a mix for potential synergistic effects, could have the potential to achieve optimal inhibitory effects against HIV-1 by targeting IN, RT, TNF- α , and NF- κ B. The synergistic interaction of these compounds, due to high inhibitory activities on various target mechanisms, could lead to greater inhibition of the virus than using a single compound alone. Despite the various in vitro cytotoxicity and inhibition analyses of the single compounds, further research and clinical trials are needed to fully evaluate the safety and inhibition efficiency of the combination of flavonoids from *Salicornia* spp., but it holds promise as a potential treatment for HIV-1 infections.

Table 4. Selected phenolic compounds detected in *Salicornia* spp. reviewed for their HIV-inhibitory properties. Data are presented as mean values and standard deviations measured by triplicates.

Name	HIV-1 Inhibition Target	Inhibition Activity	Reference
Caffeic acid	IN assay in infected MT-2 cells	IC ₅₀ inhibition: >278 μ M	[149]
	Syncytia formation in Molt-3 cells	93% inhibition at 255 μ M	[150]
	Matrix protein by immunofluorescence analysis	50% inhibition at 255 μ M	[150]
	Capsid protein by immunofluorescence analysis	57% inhibition at 255 μ M	[150]
	RT assay	32% inhibition at 255 μ M	[150]
Ferulic acid	No inhibitory effect	N/A	[151,152]
Chlorogenic acid	IN assay in infected MT-2 cells	IC ₅₀ inhibition: >142 μ M	[149]
	RT by colorimetric enzyme immunoassay	IC ₅₀ inhibition: 374 μ M	[153]
Apigenin	RT by PCR assay	IC ₅₀ : >37.03 μ M	[69]
	HIV-1 in infected OM-10.1 cells	IC ₅₀ inhibition: 12 μ M	[148]
Kaempferol	IN integration	IC ₅₀ inhibition: 64.7 \pm 18.1 μ M	[146]
	IN cleavage	IC ₅₀ inhibition: 97.8 \pm 9.2 μ M	[146]
	RT by PCR assay	IC ₅₀ inhibition: >34.94 μ M	[69]
	PR by fluorescence assay	62.7% inhibition at 174.7 μ M	[154]
Quercetin	RT by ELOSA	IC ₅₀ inhibition: 60 μ M	[135]
	PR by oligopeptide cleavage assay	IC ₅₀ inhibition: >100 μ M	[135]
	α -glucosidase inhibition assay	IC ₅₀ inhibition: >100 μ M	[135]
	IN integration	IC ₅₀ inhibition: 13.6 \pm 3.4 μ M	[146]
	IN cleavage	IC ₅₀ inhibition: 23.6 \pm 6.6 μ M	[146]
	RT by PCR assay	IC ₅₀ inhibition: <1.65 μ M	[69]
	RT by PCR assay	100% inhibition at 6.62 μ M	[69]
	RT by ELISA colorimetric enzyme immunoassay	43.41 \pm 4.56% inhibition at 661.7 μ M	[155]
Isorhamnetin	HIV-1 replication in H9 lymphocyte cells	EC ₅₀ inhibition: 6.6 μ M	[154]
	RT by ELISA colorimetric enzyme immunoassay	56.99 \pm 3.91% inhibition at 632.4 μ M	[155]

Table 4. Cont.

Name	HIV-1 Inhibition Target	Inhibition Activity	Reference
Myricetin	IN integration	IC ₅₀ inhibition: 2.5 ± 1.0 µM	[146]
	IN cleavage	IC ₅₀ inhibition: 7.6 ± 0.6 µM	[146]
	RT by PCR assay	IC ₅₀ inhibition: <1.57 µM	[69]
	RT by PCR assay	100% inhibition at 6.29 µM	[69]
	HIV-1 Blood and lymphocytes infected TZM-bl cells	87% inhibition at 100 µM	[147]
	HIV-1 Blood and lymphocytes infected TZM-bl cells	IC ₅₀ inhibition: 20.43 µM	[147]
	HIV-1 in infected MT4 cells	EC ₅₀ inhibition: 230 µM	[156]
	RT assay	IC ₅₀ inhibition: 7.6 µM	[156]
	RT	Docking energy: −7.0 kcal/mol	[156]
Isoquercitrin	PR by fluorescence assay	64.4% inhibition at 107.7 µM	[154]
Myricitrin	PR by fluorescence assay	50.4% inhibition at 107.7 µM	[154]
	HIV-1 in infected MT4 cells	EC ₅₀ inhibition: 120 µM	[156]
	RT assay	IC ₅₀ inhibition: 10.6 µM	[156]
	RT	Docking energy: −5.0 kcal/mol	[156]

IC₅₀: Half maximal inhibitory concentration, EC₅₀: Half maximal effective concentration, IN: Integrase, RT: Reverse transcriptase, ELOSA: Enzyme-Linked Oligonucleotide Sorbent Assay, PCR: Polymerase Chain Reaction, ELISA: Enzyme-Linked Immunosorbent Assay, PR: Protease.

A large-scale study on the molecular docking energies of all the selected compounds should be conducted on IN, RT, capsid, PR, matrix protein, GP120, GP41, etc., to better understand the HIV-1 protein and enzyme inhibitions.

5. Discussion

To ensure correct dosage and avoid possible toxicity, the hormesis relationships between the dose of a purified *Salicornia* spp. nutraceutical extract and its effect on the targeted virus needs to be established. Some of the hormesis relationships are between the host membrane protein NTCP, other viral host receptors, and the administration of phenolic compounds found in *Salicornia* spp. Optimal function of NTCP is necessary to maintain optimal liver function, so overinhibition of this host membrane protein to exclude viral infection can be damaging for liver cells [66,112,157]. Multiple hormesis effects have also been seen from increased phenolic intakes towards cancer therapy, as excessive intake has been shown to increase the risk of acute myeloid leukemia in infants [158,159]. However, phenolic compounds have been found to exert antitumor functions by inhibiting leukemic cell growth, and in a recent study, quercetin was found to be applicable in leukemic cancer therapy [160]. Hormesis effects from the pure fractions of *Salicornia* spp. should be investigated to avoid an increased risk of adverse effects as an extraordinarily high concentration of flavonoids, e.g., myricetin, have been found [18,44,161]. If extracts of *Salicornia* spp. are produced and fractionated, these possible adverse effects might be avoided if individual phenolic compounds, or other compounds being co-extracted and co-isolated, are discarded in the fractionation process. Greater understanding of phenolic compound-disease and phenolic compound-symptom hormesis relationships should be obtained before using *Salicornia* spp. phenolic compound-rich extracts for administration against the diseases and symptoms discussed in this review [44].

The reviewed phenolic compounds with the highest concentrations, see Table 1, are myricetin (*S. ramosissima* J. Woods, 465.5 ± 23.3 mg/kgDM), isoquercitrin (*S. fruticosa* L., 531.4 ± 9.4 mg/kgDM), and myricitrin (*S. fruticosa* L., 3314.5 ± 94.8 mg/kgDM), and a potential anti-viral therapy agent containing these compounds could be administered against H1N1, HBV, and HIV-1 infections. Inhibition of H1N1 plaque formation,

IN integration and cleavage, and RT are seen from myricetin, NA from isoquercitrin, and HIV-1-infected cells from myricitrin. However, myricitrin has not been well documented for anti-viral effects and has not been well quantified in *Salicornia* spp., so its presence as an antiviral source remains largely undiscovered, unlike the array of other bioactive properties expelled by myricitrin [162–166]. Furthermore, as non-stabilized flavonoid glucosides release glucosides during digestion, the hydrolysis of the ether bond in isoquercitrin, yielding quercetin, will be able to exhibit a great inhibitory effect against HBV-RT and HBcAg due to the release of quercetin [167].

Plant extracts of *Salicornia* spp. are complex mixtures of high and low molecular compounds, and the in vitro analysis for H1N1, HBV, HCV, and HIV-1 are made using different assays by different providers, and with or without different cell lines. It is important to note that individual assays indicate specific antiviral enzymatic inhibition, and are only comparable for the specific virus and with a specific cell line, in specific comparable concentrations of the anti-viral compounds. To better understand the anti-viral therapeutical properties of the compounds isolated from *Salicornia* spp., testing each compound against a standard set of assays in vitro and using in vivo animal trials is suggested. It is also suggested that the combination of various molecule classes, including terpenoids, lectins, and proteins, are involved in the general process of anti-viral activity, such as virus adsorption on host cells or contaminated cell molecule release, as interactions of complex mixtures might give surprising synergistic effects [135]. As the phenolic compounds of *Salicornia* spp. here have been reviewed for their anti-viral effects, these findings can be used for the formulation of natural anti-viral therapeutic agents, as some phenolic compounds generally show similar inhibition activities towards the viral therapeutic targets as the known synthetic agents [82]. To ensure safe handling and processing, the optimized extraction of the selected compounds should be investigated, and a fractionated extract should be stabilized; i.e., through encapsulation with food-grade materials. Phenolic compounds have been proven to have a high destruction rate due to environmental stresses, and the encapsulation of extract is, therefore, essential for stabilization as the compounds, due to high enzymatic activity, are cleaved into smaller, less bioactive compounds [168].

In conclusion, these findings highlight the potential of mono- and polyhydroxylated phenolic compounds in the species of the genus *Salicornia* as a source of anti-H1N1, anti-HBV, anti-HCV, and anti-HIV-1 agents and warrant further investigation into their potential use in clinical settings.

6. Methodology

The review was performed through a literature search on phenolic compounds and H1N1, HBV, HCV, and HIV. The search was performed using the Google Scholar search engine from January 2023 to May 2023. The following keywords were used for targeted searches between viruses and specific phenolic compounds: H1N1, HBV, HCV, HIV, inhibition, virus, M2, reverse transcriptase, integrase, NNRT, protease, neuraminidase, hemagglutinin, envelope glycoprotein, HBsAg, HBeAg, HBcAg, HCCAg, bioactivity, caffeic acid, ferulic acid, chlorogenic acid, apigenin, myricetin, quercetin, kaempferol, isorhamnetin, myricitrin, and isoquercitrin.

Author Contributions: Conceptualization, M.F., I.A. and M.H.T.; writing—original draft preparation, M.F., S.E.K.K. and I.A.; writing—review and editing, T.C. and M.H.T.; supervision, T.C. and M.H.T. All authors have read and agreed to the published version of the manuscript.

Funding: This study has received funding from the European Union’s Horizon 2020 research and innovation program under Grant Agreement No 862834. Any results of this project reflect only this consortium’s view; the European Commission is not responsible for any use that may be made of the information it contains.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Rashmi, H.B.; Negi, P.S. Phenolic Acids from Vegetables: A Review on Processing Stability and Health Benefits. *Food Res. Int.* **2020**, *136*, 109208. [CrossRef] [PubMed]
2. Herrmann, K. Occurrence and Content of Hydroxycinnamic and Hydroxybenzoic Acid Compounds in Foods. *Crit. Rev. Food Sci. Nutr.* **1989**, *28*, 315–347. [CrossRef] [PubMed]
3. Naveed, M.; Hejazi, V.; Abbas, M.; Kamboh, A.A.; Khan, G.J.; Shumzaid, M.; Ahmad, F.; Babazadeh, D.; FangFang, X.; Modarresi-Ghazani, F.; et al. Chlorogenic Acid (CGA): A Pharmacological Review and Call for Further Research. *Biomed. Pharmacother.* **2018**, *97*, 67–74. [CrossRef] [PubMed]
4. Giordano, R.; Aliotta, G.E.; Johannesen, A.S.; Voetmann-Jensen, D.; Laustsen, F.H.; Andersen, L.A.; Rezai, A.; Fredsgaard, M.; Lo Vecchio, S.; Arendt-Nielsen, L.; et al. Effects of *Salicornia*-Based Skin Cream Application on Healthy Humans' Experimental Model of Pain and Itching. *Pharmaceuticals* **2022**, *15*, 150. [CrossRef] [PubMed]
5. Taheri, Y.; Suleria, H.A.R.; Martins, N.; Sytar, O.; Beyatli, A.; Yeskalyeva, B.; Seitimova, G.; Salehi, B.; Semwal, P.; Painuli, S.; et al. Myricetin Bioactive Effects: Moving from Preclinical Evidence to Potential Clinical Applications. *BMC Complement. Med. Ther.* **2020**, *20*, 241. [CrossRef] [PubMed]
6. Wang, Q.; Wei, H.C.; Zhou, S.J.; Li, Y.; Zheng, T.T.; Zhou, C.Z.; Wan, X.H. Hyperoside: A Review on Its Sources, Biological Activities, and Molecular Mechanisms. *Phyther. Res.* **2022**, *36*, 2779–2802. [CrossRef]
7. Haminiuk, C.W.I.; Maciel, G.M.; Plata-Oviedo, M.S.V.; Peralta, R.M. Phenolic Compounds in Fruits—An Overview. *Int. J. Food Sci. Technol.* **2012**, *47*, 2023–2044. [CrossRef]
8. Taş, N.G.; Gökmen, V. Phenolic Compounds in Natural and Roasted Nuts and Their Skins: A Brief Review. *Curr. Opin. Food Sci.* **2017**, *14*, 103–109. [CrossRef]
9. Li, W.; Hydamaka, A.W.; Lowry, L.; Beta, T. Comparison of Antioxidant Capacity and Phenolic Compounds of Berries, Chokecherry and Seabuckthorn. *Cent. Eur. J. Biol.* **2009**, *4*, 499–506. [CrossRef]
10. Puupponen-Pimia, R.; Nohynek, L.; Meier, C.; Kahkonen, M.; Heinonen, M.; Hopia, A.; Oksman-Caldentey, K.-M. Antimicrobial Properties of Phenolic Compounds from Berries. *J. Appl. Microbiol.* **2001**, *90*, 494–507. [CrossRef]
11. Giordano, R.; Saii, Z.; Fredsgaard, M.; Hulkko, L.S.S.; Poulsen, T.B.G.; Thomsen, M.E.; Henneberg, N.; Zucolotto, S.M.; Arendt-Nielsen, L.; Papenbrock, J.; et al. Pharmacological Insights into Halophyte Bioactive Extract Action on Anti-Inflammatory, Pain Relief and Antibiotics-Type Mechanisms. *Molecules* **2021**, *26*, 3140. [CrossRef] [PubMed]
12. Kim, J.Y.; Cho, J.-Y.; Moon, J.-H.; Choi, G.-C.; Lee, K.-D.; Ham, K.-S.; Kim, S.-J. Change of Phenylpropanoic Acid and Flavonol Contents at Different Growth Stage of Glasswort (*Salicornia herbacea* L.). *Food Sci. Biotechnol.* **2014**, *23*, 685–691. [CrossRef]
13. Hulkko, L.S.S.; Rocha, R.M.; Trentin, R.; Fredsgaard, M.; Chaturvedi, T.; Custódio, L.; Thomsen, M.H. Bioactive Extracts from *Salicornia ramosissima* J. Woods Biorefinery as a Source of Ingredients for High-Value Industries. *Plants* **2023**, *12*, 1251. [CrossRef]
14. Pérez-Jiménez, J.; Neveu, V.; Vos, F.; Scalbert, A. Identification of the 100 Richest Dietary Sources of Polyphenols: An Application of the Phenol-Explorer Database. *Eur. J. Clin. Nutr.* **2010**, *64*, S112–S120. [CrossRef] [PubMed]
15. Zengin, G.; Aumeeruddy-Elalfi, Z.; Mollica, A.; Yilmaz, M.A.; Mahomoodally, M.F. In Vitro and in Silico Perspectives on Biological and Phytochemical Profile of Three Halophyte Species—A Source of Innovative Phytopharmaceuticals from Nature. *Phytomedicine* **2018**, *38*, 35–44. [CrossRef] [PubMed]
16. Ben Farhat, M.; Beji-Serairi, R.; Selmi, S.; Saidani-Tounsi, M.; Abdely, C. *Salicornia fruticosa* L. and *Portulaca oleracea* L. Antioxidants as Affected by Domestic Cooking Processes. *Int. J. Gastron. Food Sci.* **2022**, *27*, 100462. [CrossRef]
17. Won, K.J.; Lee, K.P.; Baek, S.; Cui, L.; Kweon, M.H.; Jung, S.H.; Ryu, Y.K.; Hong, J.M.; Cho, E.A.; Shin, H.S.; et al. Desalted *Salicornia europaea* Extract Attenuated Vascular Neointima Formation by Inhibiting the MAPK Pathway-Mediated Migration and Proliferation in Vascular Smooth Muscle Cells. *Biomed. Pharmacother.* **2017**, *94*, 430–438. [CrossRef]
18. Silva, A.M.; Lago, J.P.; Pinto, D.; Moreira, M.M.; Grosso, C.; Cruz Fernandes, V.; Delerue-Matos, C.; Rodrigues, F. *Salicornia ramosissima* Bioactive Composition and Safety: Eco-Friendly Extractions Approach (Microwave-Assisted Extraction vs. Conventional Maceration). *Appl. Sci.* **2021**, *11*, 4744. [CrossRef]
19. Wang, X.; Bai, J.; Wang, W.; Zhang, G.; Yin, S.; Wang, D. A Comparative Metabolomics Analysis of the Halophyte Suaeda Salsa and *Salicornia europaea*. *Environ. Geochem. Health* **2021**, *43*, 1109–1122. [CrossRef]
20. Cybulska, I.; Zembruska, J.; Brudecki, G.; Thomsen, M.H. Optimizing Methods to Characterize Caffeic, Ferulic, and Chlorogenic Acids in *Salicornia sinus-persica* and *Salicornia bigelovii* Extracts by Tandem Mass Spectrometry (LC-MS/MS). *BioResources* **2021**, *16*, 5508–5523. [CrossRef]
21. Lee, D.; Park, J.Y.; Lee, S.; Kang, K.S. In Vitro Studies to Assess the α -Glucosidase Inhibitory Activity and Insulin Secretion Effect of Isorhamnetin 3-O-Glucoside and Quercetin 3-O-Glucoside Isolated from *Salicornia herbacea*. *Processes* **2021**, *9*, 483. [CrossRef]
22. Ameixa, O.M.C.C.; Rebelo, J.; Silva, H.; Pinto, D.C.G.A. Gall Midge *Baldratia salicorniae* Kieffer (Diptera: Cecidomyiidae) Infestation on *Salicornia europaea* L. Induces the Production of Specialized Metabolites with Biotechnological Potential. *Phytochemistry* **2022**, *200*, 113207. [CrossRef] [PubMed]
23. Kim, K.-S.; Park, S.-H. Isolation and Identification of Antioxidant Flavonoids from *Salicornia herbacea* L. *Appl. Biol. Chem.* **2004**, *47*, 120–123.

24. Kong, C.S.; Lee, J.I.; Kim, Y.A.; Kim, J.A.; Bak, S.S.; Hong, J.W.; Park, H.Y.; Yea, S.S.; Seo, Y. Evaluation on Anti-Adipogenic Activity of Flavonoid Glucopyranosides from *Salicornia herbacea*. *Process Biochem.* **2012**, *47*, 1073–1078. [CrossRef]
25. Ekanayake, S.; Egodawatta, C.; Attanayake, R.N.; Perera, D. From Salt Pan to Saucepan: *Salicornia*, a Halophytic Vegetable with an Array of Potential Health Benefits. *Food Front.* **2023**, *4*, 641–676. [CrossRef]
26. Panth, N.; Park, S.-H.; Kim, H.; Kim, D.-H.; Oak, M.-H. Protective Effect of *Salicornia europaea* Extracts on High Salt Intake-Induced Vascular Dysfunction and Hypertension. *Int. J. Mol. Sci.* **2016**, *17*, 1176. [CrossRef]
27. Rhee, M.H.; Park, H.-J.; Cho, J.Y. *Salicornia herbacea*: Botanical, Chemical and Pharmacological Review of Halophyte Marsh Plant. *J. Med. Plants Res.* **2009**, *3*, 548–555.
28. Oliveira-Alves, S.C.; Andrade, F.; Prazeres, I.; Silva, A.B.; Capelo, J.; Duarte, B.; Caçador, I.; Coelho, J.; Serra, A.T.; Bronze, M.R. Impact of Drying Processes on the Nutritional Composition, Volatile Profile, Phytochemical Content and Bioactivity of *Salicornia ramosissima* j. Woods. *Antioxidants* **2021**, *10*, 1312. [CrossRef]
29. Hryniewicz, K.; Patz, S.; Ruppel, S. *Salicornia europaea* L. as an Underutilized Saline-Tolerant Plant Inhabited by Endophytic Diazotrophs. *J. Adv. Res.* **2019**, *19*, 49–56. [CrossRef]
30. Zhang, S.; Wei, M.; Cao, C.; Ju, Y.; Deng, Y.; Ye, T.; Xia, Z.; Chen, M. Effect and Mechanism of *Salicornia bigelovii* Torr. Plant Salt on Blood Pressure in SD Rats. *Food Funct.* **2015**, *6*, 920–926. [CrossRef]
31. Cho, H.D.; Lee, J.H.; Jeong, J.H.; Kim, J.Y.; Yee, S.T.; Park, S.K.; Lee, M.K.; Seo, K.-i. Production of Novel Vinegar Having Antioxidant and Anti-Fatigue Activities from *Salicornia herbacea* L. *J. Sci. Food Agric.* **2016**, *96*, 1085–1092. [CrossRef]
32. Patel, S. *Salicornia*: Evaluating the Halophytic Extremophile as a Food and a Pharmaceutical Candidate. *3 Biotech* **2016**, *6*, 1–10. [CrossRef]
33. Cole, G.M.; Lim, G.P.; Yang, F.; Teter, B.; Begum, A.; Ma, Q.; Harris-White, M.E.; Frautschy, S.A. Prevention of Alzheimer's Disease: Omega-3 Fatty Acid and Phenolic Anti-Oxidant Interventions. *Neurobiol. Aging* **2005**, *26*, 133–136. [CrossRef] [PubMed]
34. Mateos, R.; Pérez-Correa, J.R.; Domínguez, H. Bioactive Properties of Marine Phenolics. *Mar. Drugs* **2020**, *18*, 501. [CrossRef] [PubMed]
35. Grosso, G.; Bella, F.; Godos, J.; Sciacca, S.; Del Rio, D.; Ray, S.; Galvano, F.; Giovannucci, E.L. Possible Role of Diet in Cancer: Systematic Review and Multiple Meta-Analyses of Dietary Patterns, Lifestyle Factors, and Cancer Risk. *Nutr. Rev.* **2017**, *75*, 405–419. [CrossRef] [PubMed]
36. Bertolio, M.L.; Mukamal, K.J.; Cahill, L.E.; Hou, T.; Ludwig, D.S.; Mozaffarian, D.; Willett, W.C.; Hu, F.B.; Rimm, E.B. Changes in Intake of Fruits and Vegetables and Weight Change in United States Men and Women Followed for Up to 24 Years: Analysis from Three Prospective Cohort Studies. *PLOS Med.* **2015**, *12*, e1001878. [CrossRef]
37. Limongelli, F.; Crupi, P.; Clodoveo, M.L.; Corbo, F.; Muraglia, M. Overview of the Polyphenols in *Salicornia*: From Recovery to Health-Promoting Effect. *Molecules* **2022**, *27*, 7954. [CrossRef] [PubMed]
38. Suzuki, A.; Kagawa, D.; Fujii, A.; Ochiai, R.; Tokimitsu, I.; Saito, I. Short- and Long-Term Effects of Ferulic Acid on Blood Pressure in Spontaneously Hypertensive Rats. *Am. J. Hypertens.* **2002**, *15*, 351–357. [CrossRef]
39. Srinivasan, M.; Sudheer, A.R.; Menon, V.P. Ferulic Acid: Therapeutic Potential Through Its Antioxidant Property. *J. Clin. Biochem. Nutr.* **2007**, *40*, 92–100. [CrossRef]
40. Valentová, K.; Vrba, J.; Bancířová, M.; Ulrichová, J.; Křen, V. Isoquercitrin: Pharmacology, Toxicology, and Metabolism. *Food Chem. Toxicol.* **2014**, *68*, 267–282. [CrossRef]
41. Kim, S.; Lee, E.Y.; Hillman, P.F.; Ko, J.; Yang, I.; Nam, S.J. Chemical Structure and Biological Activities of Secondary Metabolites from *Salicornia europaea* L. *Molecules* **2021**, *26*, 2252. [CrossRef]
42. Park, S.H.; Ko, S.K.; Choi, J.G.; Chung, S.H. *Salicornia herbacea* Prevents High Fat Diet-Induced Hyperglycemia and Hyperlipidemia in ICR Mice. *Arch. Pharm. Res.* **2006**, *29*, 256–264. [CrossRef] [PubMed]
43. U.S. Food and Drug Administration. GRAS Notices. Available online: https://www.cfsanappsexternal.fda.gov/scripts/fdcc/index.cfm?set=GRASNotices&sort=FDA_s_Letter&order=DESC&showAll=true&type=basic&search= (accessed on 23 February 2023).
44. Murakami, A. Dose-Dependent Functionality and Toxicity of Green Tea Polyphenols in Experimental Rodents. *Arch. Biochem. Biophys.* **2014**, *557*, 3–10. [CrossRef] [PubMed]
45. Lambert, J.D.; Sang, S.; Yang, C.S. Possible Controversy over Dietary Polyphenols: Benefits vs Risks. *Chem. Res. Toxicol.* **2007**, *20*, 583–585. [CrossRef] [PubMed]
46. Rahmani, R.; El Arbi, K.; Aydi, S.S.; Hżami, A.; Tlahig, S.; Najar, R.; Aydi, S.; Debouba, M. Biochemical Composition and Biological Activities of *Salicornia europaea* L. from Southern Tunisia. *J. Food Meas. Charact.* **2022**, *16*, 4833–4846. [CrossRef]
47. De Souza, M.M.; Mendes, C.R.; Doncato, K.B.; Badiale-Furlong, E.; Costa, C.S. Growth, Phenolics, Photosynthetic Pigments, and Antioxidant Response of Two New Genotypes of Sea Asparagus (*Salicornia neei* Lag.) to Salinity under Greenhouse and Field Conditions. *Agriculture* **2018**, *8*, 115. [CrossRef]
48. Chaturvedi, T.; Hulkko, L.S.S.; Fredsgaard, M.; Thomsen, M.H. Extraction, Isolation, and Purification of Value-Added Chemicals from Lignocellulosic Biomass. *Processes* **2022**, *10*, 1752. [CrossRef]
49. Cárdenas-Pérez, S.; Piernik, A.; Chanona-Pérez, J.J.; Grigore, M.N.; Perea-Flores, M.J. An Overview of the Emerging Trends of the *Salicornia* L. Genus as a Sustainable Crop. *Environ. Exp. Bot.* **2021**, *191*, 104606. [CrossRef]
50. Qin, G.; Lei, J.; Li, S.; Jiang, Y.; Qiao, L.; Ren, M.; Gao, Q.; Song, C.; Fu, S.; Zhou, J.; et al. Efficient, Green Extraction of Two Biflavonoids from *Selaginella uncinata* with Deep Eutectic Solvents. *Microchem. J.* **2022**, *183*, 108085. [CrossRef]

51. Wang, X.H.; Wang, J.P. Ultrasonic-Assisted Extraction and Enrichment of the Flavonoids from *Salicornia europaea* Leaves Using Macroporous Resins and Response Surface Methodology. *Chem. Pap.* **2023**, *77*, 2769–2781. [CrossRef]
52. Soto, M.L.; Moure, A.; Domínguez, H.; Parajó, J.C. Recovery, Concentration and Purification of Phenolic Compounds by Adsorption: A Review. *J. Food Eng.* **2011**, *105*, 1–27. [CrossRef]
53. Lampejo, T. Influenza and Antiviral Resistance: An Overview. *Eur. J. Clin. Microbiol. Infect. Dis.* **2020**, *39*, 1201–1208. [CrossRef]
54. Kim, Y.; Narayanan, S.; Chang, K.O. Inhibition of Influenza Virus Replication by Plant-Derived Isoquercetin. *Antivir. Res.* **2010**, *88*, 227–235. [CrossRef] [PubMed]
55. Spreuwerberg, P.; Kroneman, M.; Paget, J. Reassessing the Global Mortality Burden of the 1918 Influenza Pandemic. *Am. J. Epidemiol.* **2018**, *187*, 2561–2567. [CrossRef]
56. World Health Organization. Influenza (Seasonal). Available online: [https://www.who.int/news-room/fact-sheets/detail/influenza-\(seasonal\)](https://www.who.int/news-room/fact-sheets/detail/influenza-(seasonal)) (accessed on 25 January 2023).
57. Soriano, V.; Barreiro, P.; Cachay, E.; Kottitil, S.; Fernandez-Montero, J.V.; de Mendoza, C. Advances in Hepatitis B Therapeutics. *Ther. Adv. Infect. Dis.* **2020**, *7*, 204993612096502. [CrossRef]
58. World Health Organization. Global Progress Report on HIV, Viral Hepatitis and Sexually Transmitted Infections. 2021. Available online: <https://www.who.int/publications/i/item/9789240027077> (accessed on 9 February 2023).
59. Emmanuel, B.; Wilson, E.M.; O'Brien, T.R.; Kottitil, S.; Lau, G. Shortening the Duration of Therapy for Chronic Hepatitis C Infection. *Lancet Gastroenterol. Hepatol.* **2017**, *2*, 832–836. [CrossRef] [PubMed]
60. Razavi-Shearer, D.; Gamkrelidze, I.; Nguyen, M.H.; Chen, D.S.; Van Damme, P.; Abbas, Z.; Abdulla, M.; Abou Rached, A.; Adda, D.; Aho, I.; et al. Global Prevalence, Treatment, and Prevention of Hepatitis B Virus Infection in 2016: A Modelling Study. *Lancet Gastroenterol. Hepatol.* **2018**, *3*, 383–403. [CrossRef] [PubMed]
61. Rehman, S.; Ashfaq, U.A.; Ijaz, B.; Riazuddin, S. Anti-Hepatitis C Virus Activity and Synergistic Effect of *Nymphaea Alba* Extracts and Bioactive Constituents in Liver Infected Cells. *Microb. Pathog.* **2018**, *121*, 198–209. [CrossRef]
62. Erken, R.; Loukachov, V.; van Dort, K.; van den Hurk, A.; Takkenberg, R.B.; de Niet, A.; Jansen, L.; Willemse, S.; Reesink, H.; Kootstra, N. Quantified Integrated Hepatitis B Virus Is Related to Viral Activity in Patients with Chronic Hepatitis B. *Hepatology* **2022**, *76*, 196–206. [CrossRef]
63. Ghany, M.G.; Doo, E.C. Antiviral Resistance and Hepatitis B Therapy. *Hepatology* **2009**, *49*, S174–S184. [CrossRef]
64. Marino, A.; Cosentino, F.; Ceccarelli, M.; Moscatt, V.; Pampaloni, A.; Scuderi, D.; D'andrea, F.; Venanzi Rullo, E.; Nunnari, G.; Benanti, F.; et al. Entecavir Resistance in a Patient with Treatment-Naïve Hbv: A Case Report. *Mol. Clin. Oncol.* **2021**, *14*, 113. [CrossRef] [PubMed]
65. Liu, A.L.; Wang, H.-D.; Lee, S.M.Y.; Wang, Y.T.; Du, G.H. Structure-Activity Relationship of Flavonoids as Influenza Virus Neuraminidase Inhibitors and Their in Vitro Anti-Viral Activities. *Bioorganic Med. Chem.* **2008**, *16*, 7141–7147. [CrossRef] [PubMed]
66. Parvez, M.K.; Al-Dosari, M.S.; Arbab, A.H.; Al-Rehaily, A.J.; Abdelwahid, M.A.S. Bioassay-Guided Isolation of Anti-Hepatitis B Virus Flavonoid Myricetin-3-O-Rhamnoside along with Quercetin from *Guiera Senegalensis* Leaves. *Saudi Pharm. J.* **2020**, *28*, 550–559. [CrossRef] [PubMed]
67. Bachmetov, L.; Gal-Tanamy, M.; Shapira, A.; Vorobeychik, M.; Giterman-Galam, T.; Sathiyamoorthy, P.; Golan-Goldhirsh, A.; Benhar, I.; Tur-Kaspa, R.; Zemel, R. Suppression of Hepatitis C Virus by the Flavonoid Quercetin Is Mediated by Inhibition of NS3 Protease Activity. *J. Viral Hepat.* **2012**, *19*, e81–e88. [CrossRef]
68. Khachatoorian, R.; Arumugaswami, V.; Raychaudhuri, S.; Yeh, G.K.; Maloney, E.M.; Wang, J.; Dasgupta, A.; French, S.W. Divergent Antiviral Effects of Bioflavonoids on the Hepatitis C Virus Life Cycle. *Virology* **2012**, *433*, 346–355. [CrossRef]
69. Ono, K.; Nakane, H.; Fukushima, M.; Chermann, J.-C.; Barre-Sinoussi, F.B. Differential Inhibitory Effects of Various Flavonoids on the Activities of Reverse Transcriptase and Cellular DNA and RNA Polymerases. *Eur. J. Biochem.* **1990**, *190*, 469–476. [CrossRef]
70. Xu, J.; Xu, Z.; Zheng, W. A Review of the Antiviral Role of Green Tea Catechins. *Molecules* **2017**, *22*, 1337. [CrossRef]
71. Besednova, N.N.; Andryukov, B.G.; Zaporozhets, T.S.; Kryzhanovsky, S.P.; Fedyanina, L.N.; Kuznetsova, T.A.; Zvyagintseva, T.N.; Shchelkanov, M.Y. Antiviral Effects of Polyphenols from Marine Algae. *Biomedicines* **2021**, *9*, 200. [CrossRef]
72. Loaiza-Cano, V.; Monsalve-Escudero, L.M.; Filho, C.d.S.M.B.; Martinez-Gutierrez, M.; de Sousa, D.P. Antiviral Role of Phenolic Compounds against Dengue Virus: A Review. *Biomolecules* **2021**, *11*, 11. [CrossRef]
73. Badshah, S.L.; Faisal, S.; Muhammad, A.; Poulson, B.G.; Emwas, A.H.; Jaremko, M. Antiviral Activities of Flavonoids. *Biomed. Pharmacother.* **2021**, *140*, 111596. [CrossRef]
74. Nagarajan, S. Polyphenolic Compounds-A Promising Leads for Antiviral Therapy Flavonoids from Plants View Project. *Pharmacophore* **2013**, *12*, 119–130. [CrossRef]
75. Chen, L.; Gnanaraj, C.; Arulselvan, P.; El-Seedi, H.; Teng, H. A Review on Advanced Microencapsulation Technology to Enhance Bioavailability of Phenolic Compounds: Based on Its Activity in the Treatment of Type 2 Diabetes. *Trends Food Sci. Technol.* **2019**, *85*, 149–162. [CrossRef]
76. Jakobek, L.; Matić, P. Non-Covalent Dietary Fiber-Polyphenol Interactions and Their Influence on Polyphenol Bioaccessibility. *Trends Food Sci. Technol.* **2019**, *83*, 235–247. [CrossRef]
77. Tomas, M.; Rocchetti, G.; Ghisoni, S.; Giuberti, G.; Capanoglu, E.; Lucini, L. Effect of Different Soluble Dietary Fibres on the Phenolic Profile of Blackberry Puree Subjected to in Vitro Gastrointestinal Digestion and Large Intestine Fermentation. *Food Res. Int.* **2020**, *130*, 108954. [CrossRef] [PubMed]

78. Gan, C.; Liu, L.; Du, Y.; Wang, L.; Gao, M.; Wu, L.; Yang, C. Simultaneous Determination and Pharmacokinetic Study of Four Phenol Compounds in Rat Plasma by Ultra-High Performance Liquid Chromatography with Tandem Mass Spectrometry after Oral Administration of *Echinacea purpurea* Extract. *J. Sep. Sci.* **2016**, *39*, 1628–1637. [\[CrossRef\]](#) [\[PubMed\]](#)
79. Serra, A.; Rubió, L.; Borràs, X.; Macià, A.; Romero, M.P.; Motilva, M.J. Distribution of Olive Oil Phenolic Compounds in Rat Tissues after Administration of a Phenolic Extract from Olive Cake. *Mol. Nutr. Food Res.* **2012**, *56*, 486–496. [\[CrossRef\]](#)
80. Wu, X.; Wu, X.; Sun, Q.; Zhang, C.; Yang, S.Y.; Li, L.; Jia, Z. Progress of Small Molecular Inhibitors in the Development of Anti-Influenza Virus Agents. *Theranostics* **2017**, *7*, 826–845. [\[CrossRef\]](#)
81. Cho, W.K.; Yang, H.J.; Ma, J.Y. Lotus (*Nelumbo Nucifera* Gaertn.) Leaf Water Extracts Suppress Influenza A Viral Infection via Inhibition of Neuraminidase and Hemagglutinin. *J. Funct. Foods* **2022**, *91*, 105019. [\[CrossRef\]](#)
82. Cho, W.-K.; Lee, M.-M.; Ma, J.Y. Antiviral Effect of Isoquercitrin against Influenza A Viral Infection via Modulating Hemagglutinin and Neuraminidase. *Int. J. Mol. Sci.* **2022**, *23*, 13112. [\[CrossRef\]](#)
83. Nile, S.H.; Kim, D.H.; Nile, A.; Park, G.S.; Gansukh, E.; Kai, G. Probing the Effect of Quercetin 3-Glucoside from *Dianthus superbus* L. against Influenza Virus Infection—In Vitro and in Silico Biochemical and Toxicological Screening. *Food Chem. Toxicol.* **2020**, *135*, 110985. [\[CrossRef\]](#)
84. Hariono, M.; Abdullah, N.; Damodaran, K.V.; Kamarulzaman, E.E.; Mohamed, N.; Hassan, S.S.; Shamsuddin, S.; Wahab, H.A. Potential New H1N1 Neuraminidase Inhibitors from Ferulic Acid and Vanillin: Molecular Modelling, Synthesis and in Vitro Assay. *Sci. Rep.* **2016**, *6*, 38692. [\[CrossRef\]](#) [\[PubMed\]](#)
85. Ma, Z.; Hong, Q.; Wang, Y.; Liang, Q.; Tan, H.; Xiao, C.; Tang, X.; Shao, S.; Zhou, S.; Gao, Y. Ferulic Acid Induces Heme Oxygenase-1 via Activation of ERK and Nrf2. *Drug Discov. Ther.* **2011**, *5*, 299–305. [\[CrossRef\]](#) [\[PubMed\]](#)
86. Wang, X.; Stavchansky, S.; Kerwin, S.M.; Bowman, P.D. Structure-Activity Relationships in the Cytoprotective Effect of Caffeic Acid Phenethyl Ester (CAPE) and Fluorinated Derivatives: Effects on Heme Oxygenase-1 Induction and Antioxidant Activities. *Eur. J. Pharmacol.* **2010**, *635*, 16–22. [\[CrossRef\]](#) [\[PubMed\]](#)
87. Zhu, Y.; Shao, Y.; Qu, X.; Guo, J.; Yang, J.; Zhou, Z.; Wang, S. Sodium Ferulate Protects against Influenza Virus Infection by Activation of the TLR7/9-MyD88-IRF7 Signaling Pathway and Inhibition of the NF-KB Signaling Pathway. *Biochem. Biophys. Res. Commun.* **2019**, *512*, 793–798. [\[CrossRef\]](#)
88. Enkhtaivan, G.; Maria John, K.M.; Ayyanar, M.; Sekar, T.; Jin, K.J.; Kim, D.H. Anti-Influenza (H1N1) Potential of Leaf and Stem Bark Extracts of Selected Medicinal Plants of South India. *Saudi J. Biol. Sci.* **2015**, *22*, 532–538. [\[CrossRef\]](#)
89. Mani, J.S.; Johnson, J.B.; Steel, J.C.; Broszczak, D.A.; Neilsen, P.M.; Walsh, K.B.; Naiker, M. Natural Product-Derived Phytochemicals as Potential Agents against Coronaviruses: A Review. *Virus Res.* **2020**, *284*, 197989. [\[CrossRef\]](#)
90. Kim, H.Y.; Shin, H.S.; Park, H.; Kim, Y.C.; Yun, Y.G.; Park, S.; Shin, H.J.; Kim, K. In Vitro Inhibition of Coronavirus Replications by the Traditionally Used Medicinal Herbal Extracts, *Cimicifuga Rhizoma*, *Melaleuca Cortex*, *Coptidis Rhizoma*, and *Phellodendron Cortex*. *J. Clin. Virol.* **2008**, *41*, 122–128. [\[CrossRef\]](#)
91. Spasova, M.; Philipov, S.; Nikolaeva-Glomb, L.; Galabov, A.S.; Milkova, T. Cinnamoyl- and Hydroxycinnamoyl Amides of Glauconic and Their Antioxidative and Antiviral Activities. *Bioorganic Med. Chem.* **2008**, *16*, 7457–7461. [\[CrossRef\]](#)
92. Antonopoulou, I.; Sapountzaki, E.; Rova, U.; Christakopoulos, P. Inhibition of the Main Protease of SARS-CoV-2 (Mpro) by Repurposing/Designing Drug-like Substances and Utilizing Nature's Toolbox of Bioactive Compounds. *Comput. Struct. Biotechnol. J.* **2022**, *20*, 1306–1344. [\[CrossRef\]](#)
93. Antonopoulou, I.; Sapountzaki, E.; Rova, U.; Christakopoulos, P. Ferulic Acid From Plant Biomass: A Phytochemical With Promising Antiviral Properties. *Front. Nutr.* **2022**, *8*, 1358. [\[CrossRef\]](#)
94. Salman, S.; Shah, F.H.; Idrees, J.; Idrees, F.; Velagala, S.; Ali, J.; Khan, A.A. Virtual Screening of Immunomodulatory Medicinal Compounds as Promising Anti-SARS-CoV-2 Inhibitors. *Future Virol.* **2020**, *15*, 267–275. [\[CrossRef\]](#)
95. Appiah-Opong, R.; Commandeur, J.N.M.; van Vugt-Lussenburg, B.; Vermeulen, N.P.E. Inhibition of Human Recombinant Cytochrome P450s by Curcumin and Curcumin Decomposition Products. *Toxicology* **2007**, *235*, 83–91. [\[CrossRef\]](#) [\[PubMed\]](#)
96. Gong, L.; Wang, H.; Wang, T.; Liu, Y.; Wang, J.; Sun, B. Feruloylated Oligosaccharides Modulate the Gut Microbiota in Vitro via the Combined Actions of Oligosaccharides and Ferulic Acid. *J. Funct. Foods* **2019**, *60*, 103453. [\[CrossRef\]](#)
97. Wang, X.; Zhang, M.; Zhao, Y.; Wang, H.; Liu, T.; Xin, Z. Pentadecyl Ferulate, a Potent Antioxidant and Antiproliferative Agent from the Halophyte *Salicornia herbacea*. *Food Chem.* **2013**, *141*, 2066–2074. [\[CrossRef\]](#)
98. McCarty, M.F.; DiNicolantonio, J.J. Nutraceuticals Have Potential for Boosting the Type 1 Interferon Response to RNA Viruses Including Influenza and Coronavirus. *Prog. Cardiovasc. Dis.* **2020**, *63*, 383–385. [\[CrossRef\]](#) [\[PubMed\]](#)
99. Ding, Y.; Cao, Z.; Cao, L.; Ding, G.; Wang, Z.; Xiao, W. Antiviral Activity of Chlorogenic Acid against Influenza A (H1N1/H3N2) Virus and Its Inhibition of Neuraminidase. *Sci. Rep.* **2017**, *7*, 45723. [\[CrossRef\]](#)
100. Dayem, A.A.; Choi, H.Y.; Kim, Y.B.; Cho, S.-G. Antiviral Effect of Methylated Flavonol Isorhamnetin against Influenza. *PLoS ONE* **2015**, *10*, e0121610. [\[CrossRef\]](#)
101. Kanazawa, R.; Morimoto, R.; Horio, Y.; Sumitani, H.; Isegawa, Y. Inhibition of Influenza Virus Replication by Apiaceae Plants, with Special Reference to *Peucedanum japonicum* (Sacna) Constituents. *J. Ethnopharmacol.* **2022**, *292*, 115243. [\[CrossRef\]](#)
102. Kai, H.; Obuchi, M.; Yoshida, H.; Watanabe, W.; Tsutsumi, S.; Park, Y.K.; Matsuno, K.; Yasukawa, K.; Kurokawa, M. In Vitro and in Vivo Anti-Influenza Virus Activities of Flavonoids and Related Compounds as Components of Brazilian Propolis (AF-08). *J. Funct. Foods* **2014**, *8*, 214–223. [\[CrossRef\]](#)

103. Sadati, S.M.; Gheibi, N.; Ranjbar, S.; Hashemzadeh, M.S. Docking Study of Flavonoid Derivatives as Potent Inhibitors of Influenza H1N1 Virus Neuraminidase. *Biomed. Rep.* **2019**, *10*, 33–38. [\[CrossRef\]](#)
104. Kaihatsu, K. Potential Anti-Influenza Virus Agents Based on Coffee Ingredients and Natural Flavonols. *Nat. Prod. Chem. Res.* **2014**, *2*, 1000129. [\[CrossRef\]](#)
105. Albeshri, A.; Baeshen, N.A.; Bouback, T.A.; Aljaddawi, A.A. Evaluation of Cytotoxicity and Antiviral Activity of *Rhazya Stricta* Decne Leaves Extract against Influenza A/PR/8/34 (H1N1). *Saudi J. Biol. Sci.* **2022**, *29*, 103375. [\[CrossRef\]](#)
106. Motlhlatlego, K.E.; Mehrbod, P.; Fotouhi, F.; Abdalla, M.A.; Eloff, J.N.; McGaw, L.J. Anti-Influenza A Virus Activity of Two *Newtonia* Species and the Isolated Compound Myricetin-3-o-Rhamnoside. *BMC Complement. Med. Ther.* **2021**, *21*, 92. [\[CrossRef\]](#) [\[PubMed\]](#)
107. Patil, V.S.; Harish, D.R.; Vetrivel, U.; Deshpande, S.H.; Khanal, P.; Hegde, H.V.; Roy, S.; Jalalpure, S.S. Pharmacoinformatics Analysis Reveals Flavonoids and Diterpenoids from *Andrographis Paniculata* and *Thespesia Populnea* to Target Hepatocellular Carcinoma Induced by Hepatitis B Virus. *Appl. Sci.* **2022**, *12*, 10691. [\[CrossRef\]](#)
108. Shin, M.S.; Kang, E.H.; Lee, Y.I. A Flavonoid from Medicinal Plants Blocks Hepatitis B Virus-e Antigen Secretion in HBV-Infected Hepatocytes. *Antivir. Res.* **2005**, *67*, 163–168. [\[CrossRef\]](#) [\[PubMed\]](#)
109. Abdel-Hamid, N.M. A Survey on Herbal Management of Hepatocellular Carcinoma. *World J. Hepatol.* **2011**, *3*, 175. [\[CrossRef\]](#) [\[PubMed\]](#)
110. Rehman, S.; Ijaz, B.; Fatima, N.; Muhammad, S.A.; Riazuddin, S. Therapeutic Potential of *Taraxacum Officinale* against HCV NS5B Polymerase: In-Vitro and In Silico Study. *Biomed. Pharmacother.* **2016**, *83*, 881–891. [\[CrossRef\]](#)
111. Lam, B.; Henry, L.; Younossi, Z. Sofosbuvir (Sovaldi) for the Treatment of Hepatitis C. *Expert Rev. Clin. Pharmacol.* **2014**, *7*, 555–566. [\[CrossRef\]](#)
112. Watashi, K.; Urban, S.; Li, W.; Wakita, T. NTCP and Beyond: Opening the Door to Unveil Hepatitis B Virus Entry. *Int. J. Mol. Sci.* **2014**, *15*, 2892–2905. [\[CrossRef\]](#)
113. Lv, H.; An, B.; Yu, Q.; Cao, Y.; Liu, Y.; Li, S. The Hepatoprotective Effect of Myricetin against Lipopolysaccharide and D-Galactosamine-Induced Fulminant Hepatitis. *Int. J. Biol. Macromol.* **2020**, *155*, 1092–1104. [\[CrossRef\]](#)
114. Brentnall, M.; Rodriguez-Menocal, L.; De Guevara, R.L.; Cepero, E.; Boise, L.H. Caspase-9, Caspase-3 and Caspase-7 Have Distinct Roles during Intrinsic Apoptosis. *BMC Cell Biol.* **2013**, *14*, 32. [\[CrossRef\]](#) [\[PubMed\]](#)
115. Truant, R.; Antunovic, J.; Greenblatt, J.; Prives, C.; Cromlish, J.A. Direct Interaction of the Hepatitis B Virus HBx Protein with P53 Leads to Inhibition by HBx of P53 Response Element-Directed Transactivation. *J. Virol.* **1995**, *69*, 1851–1859. [\[CrossRef\]](#) [\[PubMed\]](#)
116. Gottlob, K.; Fulco, M.; Levrero, M.; Graessmann, A. The Hepatitis B Virus HBx Protein Inhibits Caspase 3 Activity. *J. Biol. Chem.* **1998**, *273*, 33347–33353. [\[CrossRef\]](#)
117. Chung, T.; Lee, Y.; Kim, C. Hepatitis B Viral HBx Induces Matrix Metalloproteinase-9 Gene Expression through Activation of ERKs and PI-3K/AKT Pathways: Involvement of Invasive Potential. *FASEB J.* **2004**, *18*, 1123–1125. [\[CrossRef\]](#)
118. Torresi, J.; Tran, B.M.; Christiansen, D.; Earnest-Silveira, L.; Schwab, R.H.M.; Vincan, E. HBV-Related Hepatocarcinogenesis: The Role of Signalling Pathways and Innovative Ex Vivo Research Models. *BMC Cancer* **2019**, *19*, 707. [\[CrossRef\]](#) [\[PubMed\]](#)
119. Kang-Park, S.; Im, J.H.; Lee, J.H.; Lee, Y.I. PTEN Modulates Hepatitis B Virus-X Protein Induced Survival Signaling in Chang Liver Cells. *Virus Res.* **2006**, *122*, 53–60. [\[CrossRef\]](#)
120. Lin, H.J.; Ku, K.L.; Lin, I.H.; Yeh, C.C. Naringenin Attenuates Hepatitis B Virus X Protein-Induced Hepatic Steatosis. *BMC Complement. Altern. Med.* **2017**, *17*, 505. [\[CrossRef\]](#) [\[PubMed\]](#)
121. Zhang, J.; Liu, X.; Zhou, W.; Cheng, G.; Wu, J.; Guo, S.; Jia, S.; Liu, Y.; Li, B.; Zhang, X.; et al. A Bioinformatics Investigation into Molecular Mechanism of Yinzhihuang Granules for Treating Hepatitis B by Network Pharmacology and Molecular Docking Verification. *Sci. Rep.* **2020**, *10*, 11448. [\[CrossRef\]](#) [\[PubMed\]](#)
122. Rehmann, B.; Nascimbeni, M. Immunology of Hepatitis B Virus and Hepatitis C Virus Infection. *Nat. Rev. Immunol.* **2005**, *5*, 215–229. [\[CrossRef\]](#)
123. Lin, Y.-T.; Wu, Y.-H.; Tseng, C.-K.; Lin, C.-K.; Chen, W.-C.; Hsu, Y.-C.; Lee, J.-C. Green Tea Phenolic Epicatechins Inhibit Hepatitis C Virus Replication via Cyclooxygenase-2 and Attenuate Virus-Induced Inflammation. *PLoS ONE* **2013**, *8*, e54466. [\[CrossRef\]](#)
124. Ma, J.Q.; Li, Z.; Xie, W.R.; Liu, C.M.; Liu, S.S. Quercetin Protects Mouse Liver against CCl₄-Induced Inflammation by the TLR2/4 and MAPK/NF-KB Pathway. *Int. Immunopharmacol.* **2015**, *28*, 531–539. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Berzsenyi, M.D.; Roberts, S.K.; Preiss, S.; Woollard, D.J.; Beard, M.R.; Skinner, N.A.; Bowden, D.S.; Visvanathan, K. Hepatic TLR2 & TLR4 Expression Correlates with Hepatic Inflammation and TNF- α in HCV & HCV/HIV Infection. *J. Viral Hepat.* **2011**, *18*, 852–860. [\[CrossRef\]](#) [\[PubMed\]](#)
126. Ivanov, A.V.; Valuev-Elliston, V.T.; Tyurina, D.A.; Ivanova, O.N.; Kochetkov, S.N.; Bartosch, B.; Isagulants, M.G. Oxidative Stress, a Trigger of Hepatitis C and B Virus-Induced Liver Carcinogenesis. *Oncotarget* **2017**, *8*, 3895–3932. [\[CrossRef\]](#) [\[PubMed\]](#)
127. Valgimigli, M.; Valgimigli, L.; Trerè, D.; Gaiani, S.; Pedulli, G.F.; Gramantieri, L.; Bolondi, L. Oxidative Stress EPR Measurement in Human Liver by Radical-Probe Technique. Correlation with Etiology, Histology and Cell Proliferation. *Free Radic. Res.* **2002**, *36*, 939–948. [\[CrossRef\]](#) [\[PubMed\]](#)
128. Higgs, M.R.; Chouteau, P.; Lerat, H. “Liver Let Die”: Oxidative DNA Damage and Hepatotropic Viruses. *J. Gen. Virol.* **2014**, *95*, 991–1004. [\[CrossRef\]](#)
129. Yu, Y.; Cui, Y.; Niedernhofer, L.J.; Wang, Y. Occurrence, Biological Consequences, and Human Health Relevance of Oxidative Stress-Induced DNA Damage. *Chem. Res. Toxicol.* **2016**, *29*, 2008–2039. [\[CrossRef\]](#)

130. Van Hung, P. Phenolic Compounds of Cereals and Their Antioxidant Capacity. *Crit. Rev. Food Sci. Nutr.* **2016**, *56*, 25–35. [\[CrossRef\]](#)
131. Sanchez-Valle, V.; Chavez-Tapia, N.C.; Uribe, M.; Mendez-Sanchez, N. Role of Oxidative Stress and Molecular Changes in Liver Fibrosis: A Review. *Curr. Med. Chem.* **2012**, *19*, 4850–4860. [\[CrossRef\]](#)
132. Shen, J.; Wang, G.; Zuo, J. Caffeic Acid Inhibits HCV Replication via Induction of IFN α Antiviral Response through P62-Mediated Keap1/Nrf2 Signaling Pathway. *Antivir. Res.* **2018**, *154*, 166–173. [\[CrossRef\]](#)
133. Wang, G.F.; Shi, L.P.; Ren, Y.D.; Liu, Q.F.; Liu, H.F.; Zhang, R.J.; Li, Z.; Zhu, F.H.; He, P.L.; Tang, W.; et al. Anti-Hepatitis B Virus Activity of Chlorogenic Acid, Quinic Acid and Caffeic Acid in Vivo and in Vitro. *Antivir. Res.* **2009**, *83*, 186–190. [\[CrossRef\]](#)
134. Chiang, L.-C.; Ng, L.-T.; Cheng, P.-W.; Chiang, W.; Lin, C.-C. Antiviral Activities of Extracts and Selected Pure Constituents of *Ocimum Basilicum*. *Clin. Exp. Pharmacol. Physiol.* **2005**, *32*, 811–816. [\[CrossRef\]](#) [\[PubMed\]](#)
135. Yu, Y.B.; Miyashiro, H.; Nakamura, N.; Hattori, M.; Jong, C.P. Effects of Triterpenoids and Flavonoids Isolated from *Alnus Firma* on HIV-1 Viral Enzymes. *Arch. Pharm. Res.* **2007**, *30*, 820–826. [\[CrossRef\]](#) [\[PubMed\]](#)
136. Wang, P.; Liu, C.; Sanches, T.; Zhong, Y.; Liu, B.; Xiong, J.; Neamati, N.; Zhao, G. Design and Synthesis of Novel Nitrogen-Containing Polyhydroxylated Aromatics as HIV-1 Integrase Inhibitors from Caffeic Acid Phenethyl Ester. *Bioorganic Med. Chem. Lett.* **2009**, *19*, 4574–4578. [\[CrossRef\]](#) [\[PubMed\]](#)
137. Tchertanov, L.; Mouscadet, J.F. Target Recognition by Catechols and β -Ketoenols: Potential Contribution of Hydrogen Bonding and Mn/Mg Chelation to HIV-1 Integrase Inhibition. *J. Med. Chem.* **2007**, *50*, 1133–1145. [\[CrossRef\]](#) [\[PubMed\]](#)
138. Bailly, F.; Cotellet, P. Anti-HIV Activities of Natural Antioxidant Caffeic Acid Derivatives: Toward an Antiviral Supplementation Diet. *Curr. Med. Chem.* **2005**, *12*, 1811–1818. [\[CrossRef\]](#)
139. Wyatt, R.; Moore, J.; Accola, M.; Desjardin, E.; Robinson, J.; Sodroski, J. Involvement of the V1/V2 Variable Loop Structure in the Exposure of Human Immunodeficiency Virus Type 1 Gp120 Epitopes Induced by Receptor Binding. *J. Virol.* **1995**, *69*, 5723–5733. [\[CrossRef\]](#)
140. Delelis, O.; Malet, I.; Li, N.; Tchertanov, L.; Calvez, V.; Marcelin, A.-G.; Subra, F.; Deprez, E.; Mouscadet, J.-F. The G140S Mutation in HIV Integrases from Raltegravir-Resistant Patients Rescues Catalytic Defect Due to the Resistance Q148H Mutation. *Nucleic Acids Res.* **2009**, *37*, 1193–1201. [\[CrossRef\]](#)
141. King, P.J.; Lee, D.J.; Reinke, R.A.; Victoria, J.G.; Beale, K.; Robinson, W.E. Human Immunodeficiency Virus Type-1 Integrase Containing a Glycine to Serine Mutation at Position 140 Is Attenuated for Catalysis and Resistant to Integrase Inhibitors. *Virology* **2003**, *306*, 147–161. [\[CrossRef\]](#)
142. Singh, S.B.; Jayasuriya, H.; Dewey, R.; Polishook, J.D.; Dombrowski, A.W.; Zink, D.L.; Guan, Z.; Collado, J.; Platas, G.; Pelaez, F.; et al. Isolation, Structure, and HIV-1-Integrase Inhibitory Activity of Structurally Diverse Fungal Metabolites. *J. Ind. Microbiol. Biotechnol.* **2003**, *30*, 721–731. [\[CrossRef\]](#)
143. Abraham, R.T.; Weiss, A. Jurkat T Cells and Development of the T-Cell Receptor Signalling Paradigm. *Nat. Rev. Immunol.* **2004**, *4*, 301–308. [\[CrossRef\]](#)
144. Yang, X.; Zhu, X.; Ji, H.; Deng, J.; Lu, P.; Jiang, Z.; Li, X.; Wang, Y.; Wang, C.; Zhao, J.; et al. Quercetin Synergistically Reactivates Human Immunodeficiency Virus Type 1 Latency by Activating Nuclear Factor-KB. *Mol. Med. Rep.* **2018**, *17*, 2501–2508. [\[CrossRef\]](#) [\[PubMed\]](#)
145. Ono, K.; Nakane, H. Mechanisms of Inhibition of Various Cellular DNA and RNA Polymerases by Several Flavonoids. *J. Biochem.* **1990**, *108*, 609–613. [\[CrossRef\]](#) [\[PubMed\]](#)
146. Fesen, M.R.; Pommier, Y.; Leteurtre, F.; Hiroguchi, S.; Yung, J.; Kohn, K.W. Inhibition of HIV-1 Integrase by Flavones, Caffeic Acid Phenethyl Ester (CAPE) and Related Compounds. *Biochem. Pharmacol.* **1994**, *48*, 595–608. [\[CrossRef\]](#) [\[PubMed\]](#)
147. Pasetto, S.; Pardi, V.; Murata, R.M. Anti-HIV-1 Activity of Flavonoid Myricetin on HIV-1 Infection in a Dual-Chamber In Vitro Model. *PLoS ONE* **2014**, *9*, e115323. [\[CrossRef\]](#)
148. Critchfield, J.W.; Butera, S.T.; Folks, T.M. Inhibition of HIV Activation in Latently Infected Cells by Flavonoid Compounds. *AIDS Res. Hum. Retroviruses* **2009**, *12*, 39–46. [\[CrossRef\]](#)
149. Tamura, H.; Akioka, T.; Ueno, K.; Chujyo, T.; Okazaki, K.I.; King, P.J.; Robinson, W.E. Anti-Human Immunodeficiency Virus Activity of 3,4,5-Tricaffeoylquinic Acid in Cultured Cells of Lettuce Leaves. *Mol. Nutr. Food Res.* **2006**, *50*, 396–400. [\[CrossRef\]](#)
150. Kreis, W.; Kaplan, M.H.; Freeman, J.; Sun, D.K.; Sarin, P.S. Inhibition of HIV Replication by Hyssop Officinalis Extracts. *Antivir. Res.* **1990**, *14*, 323–337. [\[CrossRef\]](#)
151. Sonar, V.P.; Corona, A.; Distinto, S.; Maccioni, E.; Meleddu, R.; Fois, B.; Floris, C.; Malpure, N.V.; Alcaro, S.; Tramontano, E.; et al. Natural Product-Inspired Esters and Amides of Ferulic and Caffeic Acid as Dual Inhibitors of HIV-1 Reverse Transcriptase. *Eur. J. Med. Chem.* **2017**, *130*, 248–260. [\[CrossRef\]](#)
152. Sanna, C.; Rigano, D.; Corona, A.; Piano, D.; Formisano, C.; Farci, D.; Franzini, G.; Ballero, M.; Chianese, G.; Tramontano, E.; et al. Dual HIV-1 Reverse Transcriptase and Integrase Inhibitors from *Limonium Morisianum* Arrigoni, an Endemic Species of Sardinia (Italy). *Nat. Prod. Res.* **2019**, *33*, 1798–1803. [\[CrossRef\]](#)
153. Tamayose, C.I.; Torres, P.B.; Roque, N.; Ferreira, M.J.P. HIV-1 Reverse Transcriptase Inhibitory Activity of Flavones and Chlorogenic Acid Derivatives from *Moquiniastrum floribundum* (Asteraceae). *S. Afr. J. Bot.* **2019**, *123*, 142–146. [\[CrossRef\]](#)
154. Wu, T.S.; Tsang, Z.J.; Wu, P.L.; Lin, F.W.; Li, C.Y.; Teng, C.M.; Lee, K.H. New Constituents and Antiplatelet Aggregation and Anti-HIV Principles of *Artemisia Capillaris*. *Bioorganic Med. Chem.* **2001**, *9*, 77–83. [\[CrossRef\]](#) [\[PubMed\]](#)
155. Da Silva, C.C.F.; Salatino, A.; da Motta, L.B.; Negri, G.; Salatino, M.L.F. Chemical Characterization, Antioxidant and Anti-HIV Activities of a Brazilian Propolis from Ceará State. *Rev. Bras. Farmacogn.* **2019**, *29*, 309–318. [\[CrossRef\]](#)

156. Ortega, J.T.; Suárez, A.I.; Serrano, M.L.; Baptista, J.; Pujol, F.H.; Rangel, H.R. The Role of the Glycosyl Moiety of Myricetin Derivatives in Anti-HIV-1 Activity in Vitro. *AIDS Res. Ther.* **2017**, *14*, 57. [[CrossRef](#)] [[PubMed](#)]
157. Nakabori, T.; Hikita, H.; Murai, K.; Nozaki, Y.; Kai, Y.; Makino, Y.; Saito, Y.; Tanaka, S.; Wada, H.; Eguchi, H.; et al. Sodium Taurocholate Cotransporting Polypeptide Inhibition Efficiently Blocks Hepatitis B Virus Spread in Mice with a Humanized Liver. *Sci. Rep.* **2016**, *6*, 27782. [[CrossRef](#)]
158. Ross, J.A.; Potter, J.D.; Robison, L.L. Infant Leukemia, Topoisomerase II Inhibitors, and the MLL Gene. *J. Natl. Cancer Inst.* **1994**, *86*, 1678–1680. [[CrossRef](#)]
159. Ross, J.A.; Potter, J.D.; Reaman, G.H.; Pendergrass, T.W.; Robison, L.L. Maternal Exposure to Potential Inhibitors of DNA Topoisomerase II and Infant Leukemia (United States): Report from the Children’s Cancer Group. *Cancer Causes Control* **1996**, *7*, 581–590. [[CrossRef](#)]
160. Shi, H.; Li, X.-Y.; Chen, Y.; Zhang, X.; Wu, Y.; Wang, Z.-X.; Chen, P.-H.; Dai, H.-Q.; Feng, J.; Chatterjee, S.; et al. Quercetin Induces Apoptosis via Downregulation of Vascular Endothelial Growth Factor/Akt Signaling Pathway in Acute Myeloid Leukemia Cells. *Front. Pharmacol.* **2020**, *11*, 1972. [[CrossRef](#)]
161. Bonkovsky, H.L. Hepatotoxicity Associated with Supplements Containing Chinese Green Tea (*Camellia sinensis*). *Ann. Intern. Med.* **2006**, *144*, 68–71. [[CrossRef](#)]
162. Kassem, M.E.S.; Ibrahim, L.F.; Hussein, S.R.; El-Sharawy, R.; El-Ansari, M.A.; Hassanane, M.M.; Booles, H.F. Myricitrin and Bioactive Extract of *Albizia amara* Leaves: DNA Protection and Modulation of Fertility and Antioxidant-Related Genes Expression. *Pharm. Biol.* **2016**, *54*, 2404–2409. [[CrossRef](#)]
163. Yan, L.J.; Yang, H.T.; Duan, H.Y.; Wu, J.T.; Qian, P.; Fan, X.W.; Wang, S. Myricitrin Inhibits Vascular Adhesion Molecule Expression in TNF- α -Stimulated Vascular Smooth Muscle Cells. *Mol. Med. Rep.* **2017**, *16*, 6354–6359. [[CrossRef](#)]
164. Córdova, M.M.; de Paula Werner, M.F.; da Silva, M.D.; Ruani, A.P.; Pizzolatti, M.G.; Santos, A.R.S. Further Antinociceptive Effects of Myricitrin in Chemical Models of Overt Nociception in Mice. *Neurosci. Lett.* **2011**, *495*, 173–177. [[CrossRef](#)] [[PubMed](#)]
165. Xu, R.; Zhang, Y.; Ye, X.; Xue, S.; Shi, J.; Pan, J.; Chen, Q. Inhibition Effects and Induction of Apoptosis of Flavonoids on the Prostate Cancer Cell Line PC-3 In Vitro. *Food Chem.* **2013**, *138*, 48–53. [[CrossRef](#)] [[PubMed](#)]
166. Huang, Q.; Gao, B.; Wang, L.; Hu, Y.Q.; Lu, W.G.; Yang, L.; Luo, Z.J.; Liu, J. Protective Effects of Myricitrin against Osteoporosis via Reducing Reactive Oxygen Species and Bone-Resorbing Cytokines. *Toxicol. Appl. Pharmacol.* **2014**, *280*, 550–560. [[CrossRef](#)] [[PubMed](#)]
167. Manach, C.; Morand, C.; Demigné, C.; Texier, O.; Régéat, F.; Rémésy, C. Bioavailability of Rutin and Quercetin in Rats. *FEBS Lett.* **1997**, *409*, 12–16. [[CrossRef](#)]
168. Faridi Esfanjani, A.; Jafari, S.M. Biopolymer Nano-Particles and Natural Nano-Carriers for Nano-Encapsulation of Phenolic Compounds. *Colloids Surf. B Biointerfaces* **2016**, *146*, 532–543. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.