

High throughput 16S rRNA gene amplicon sequencing

A fast and cheap method to study the influence of microbial community composition on activated sludge floc properties

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High throughput 16S rRNA gene amplicon sequencing: a fast and cheap method to study the influence of microbial community composition on activated sludge properties



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Introduction

A reliable and reproducible method for identification and quantification of microorganisms is important for the studies of microbial communities in activated sludge and for the demonstration of their significance for plant operation and stability.

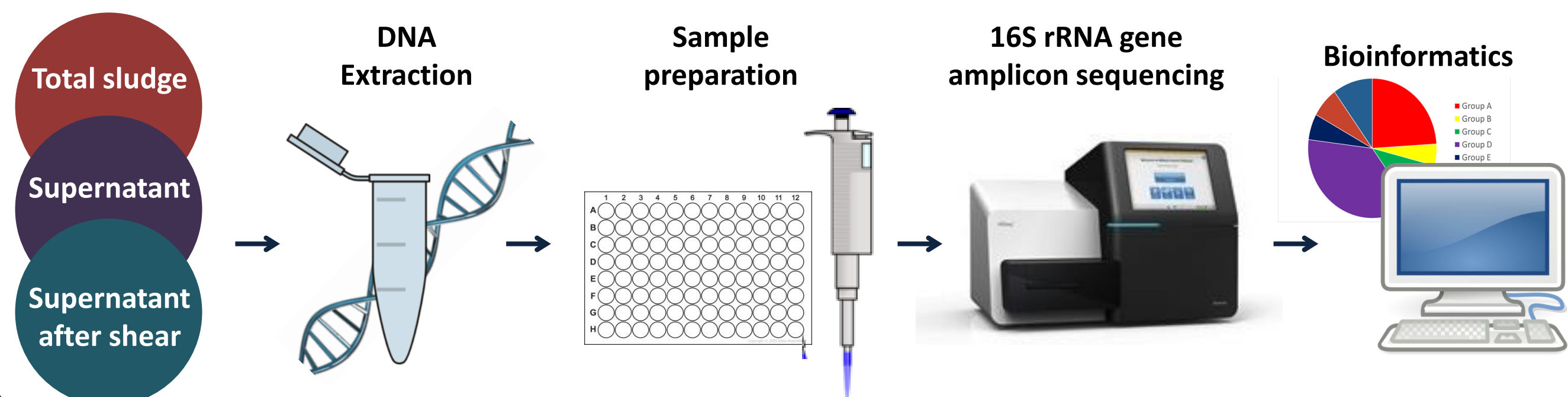
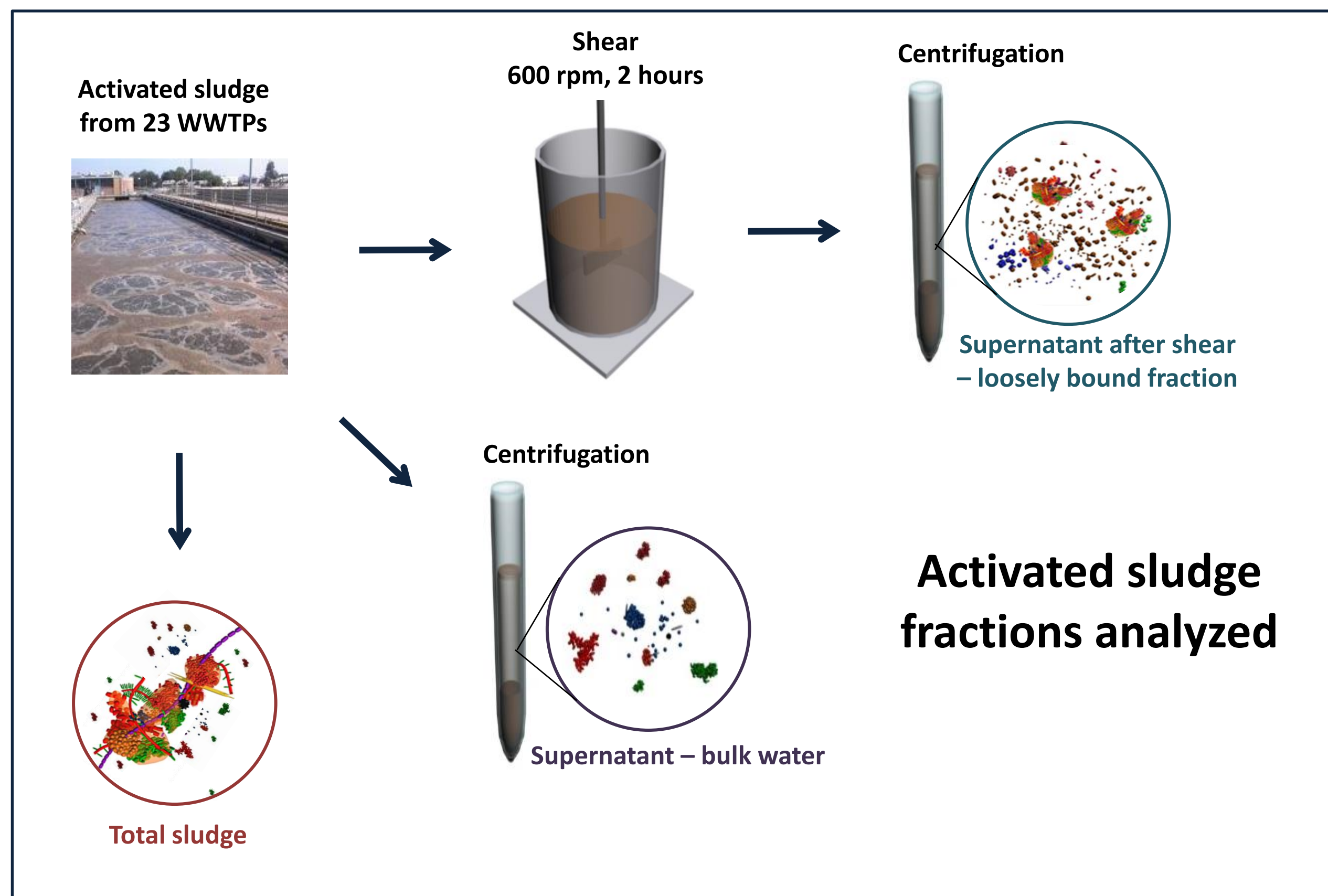
DNA-based identification of microorganisms using 16S rRNA gene amplicon sequencing has been developed over the past few years and is now ready to use for more comprehensive studies related to plant operation and optimization thanks to short analysis time, low cost, high throughput, and high taxonomic resolution.

Since bacterial morphology, mode of growth and EPS composition determine floc size, shape and strength, which in turn influence important sludge properties, the link established between the microbial community structure and physico-chemical sludge characteristics may provide a better understanding of the activated sludge process.

Objectives

- To show how 16S rRNA gene amplicon sequencing can be used to reveal factors of importance for the operation of 23 full-scale nutrient removal plants that can be related to settling problems and floc properties.
- To investigate whether the microbial community composition differs between the flocs and the supernatant (bulk water) and whether certain bacterial species are prone to detachment from the flocs.

Methods



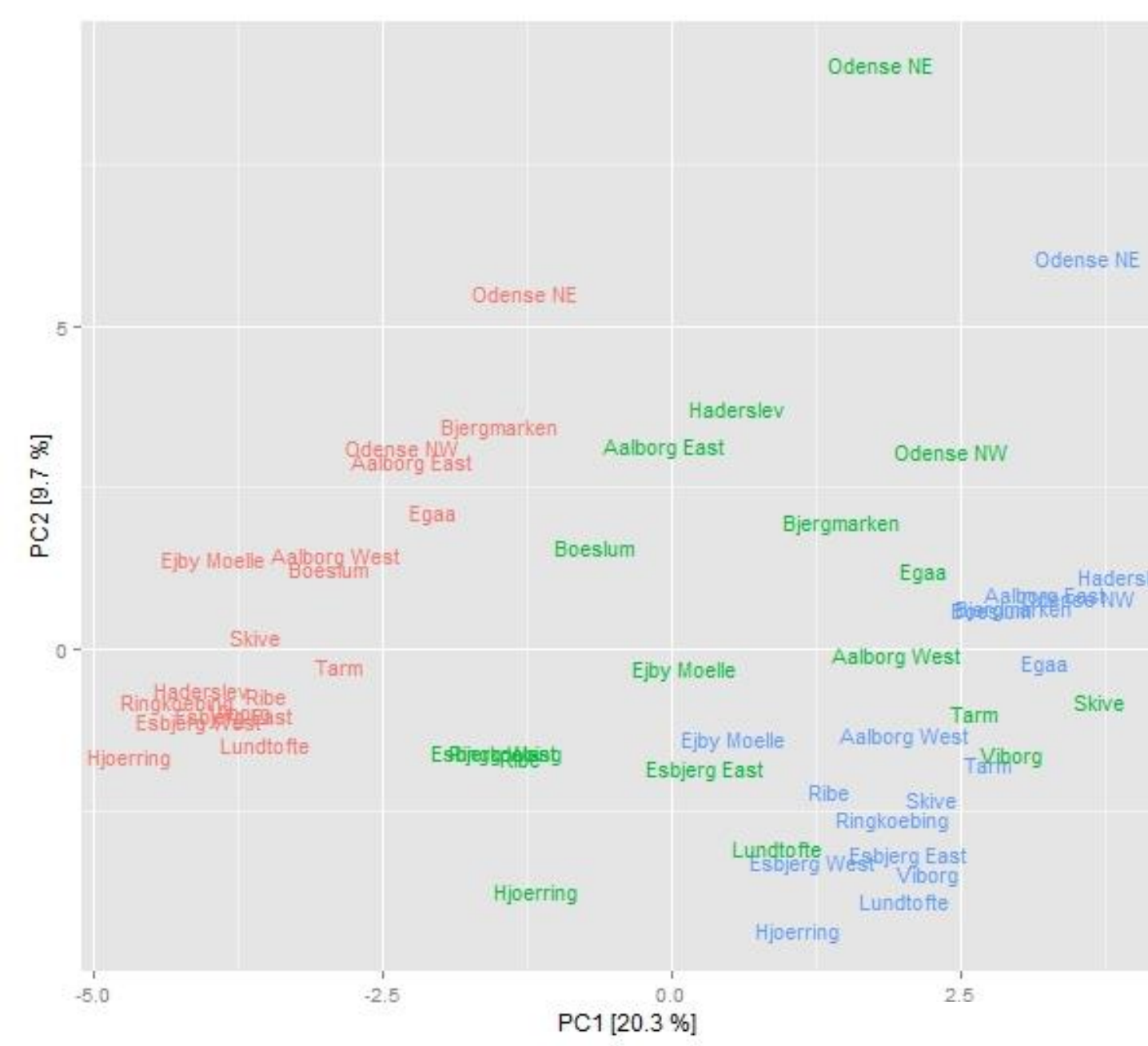
Conclusions

16S rRNA gene amplicon sequencing is suitable for comprehensive studies of WWTPs thanks to short analysis time, low cost, high throughput, and high taxonomic resolution.

A number of bacterial species can be correlated to the sludge characteristics that are important for the proper plant operation (SVI, floc strength, and EPS content).

Specific bacteria are enriched in the bulk water fraction and in the fraction loosely bound to the floc.

Results

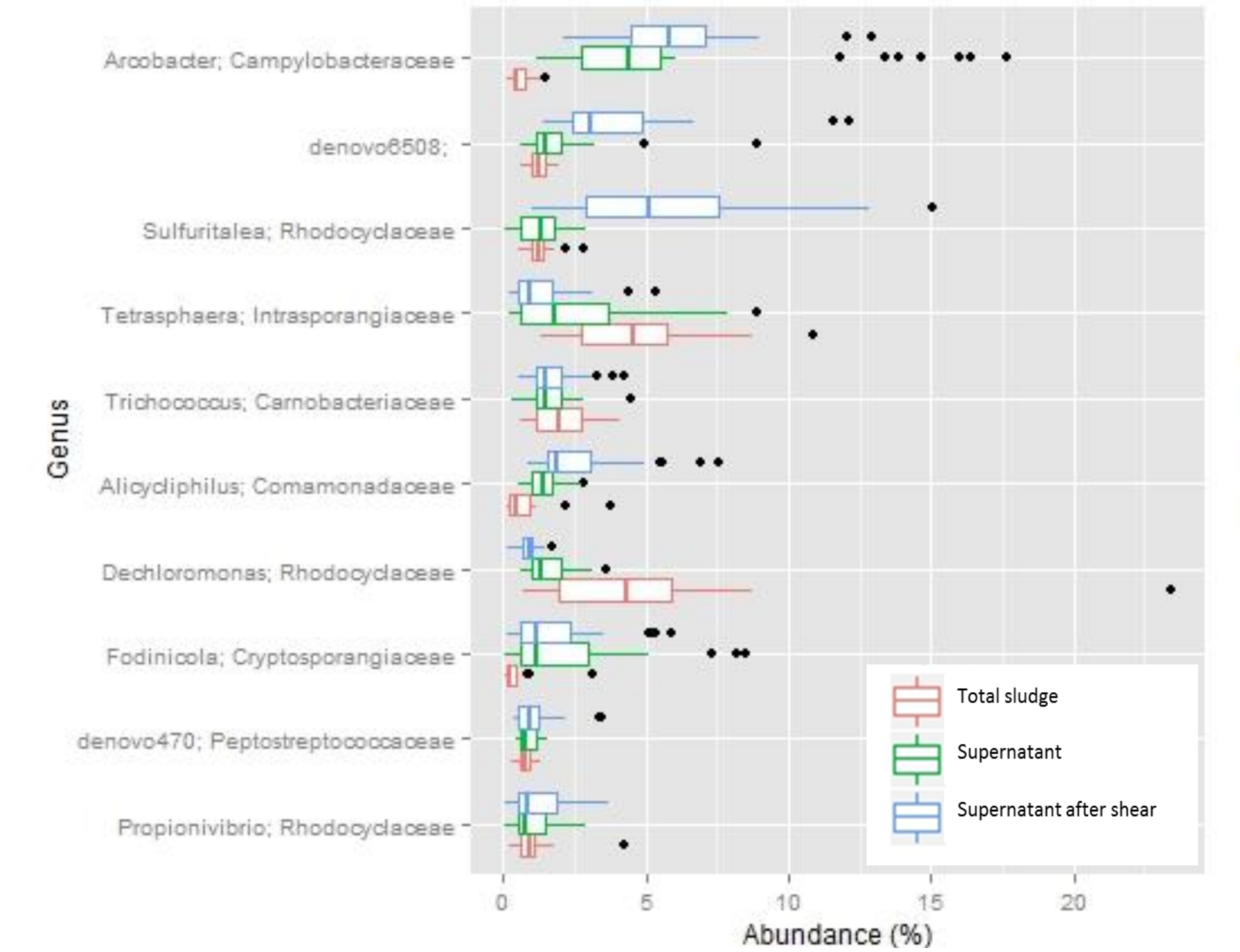


Community composition in different activated sludge fractions

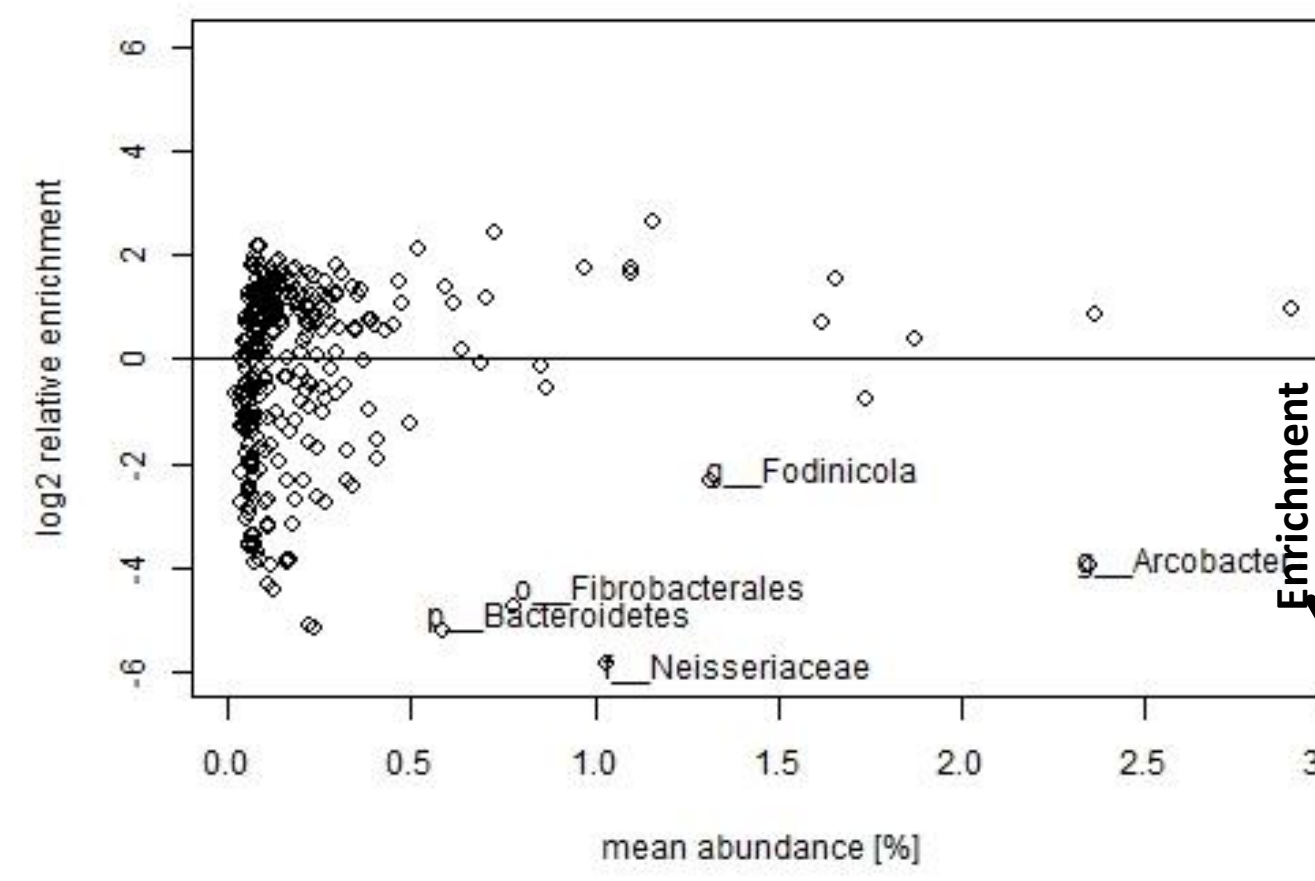
The figure shows the relationship between all samples analyzed from 23 WWTPs.

Samples grouping into individual sludge fractions (total sludge – bulk water – loosely bound fraction) can be clearly observed.

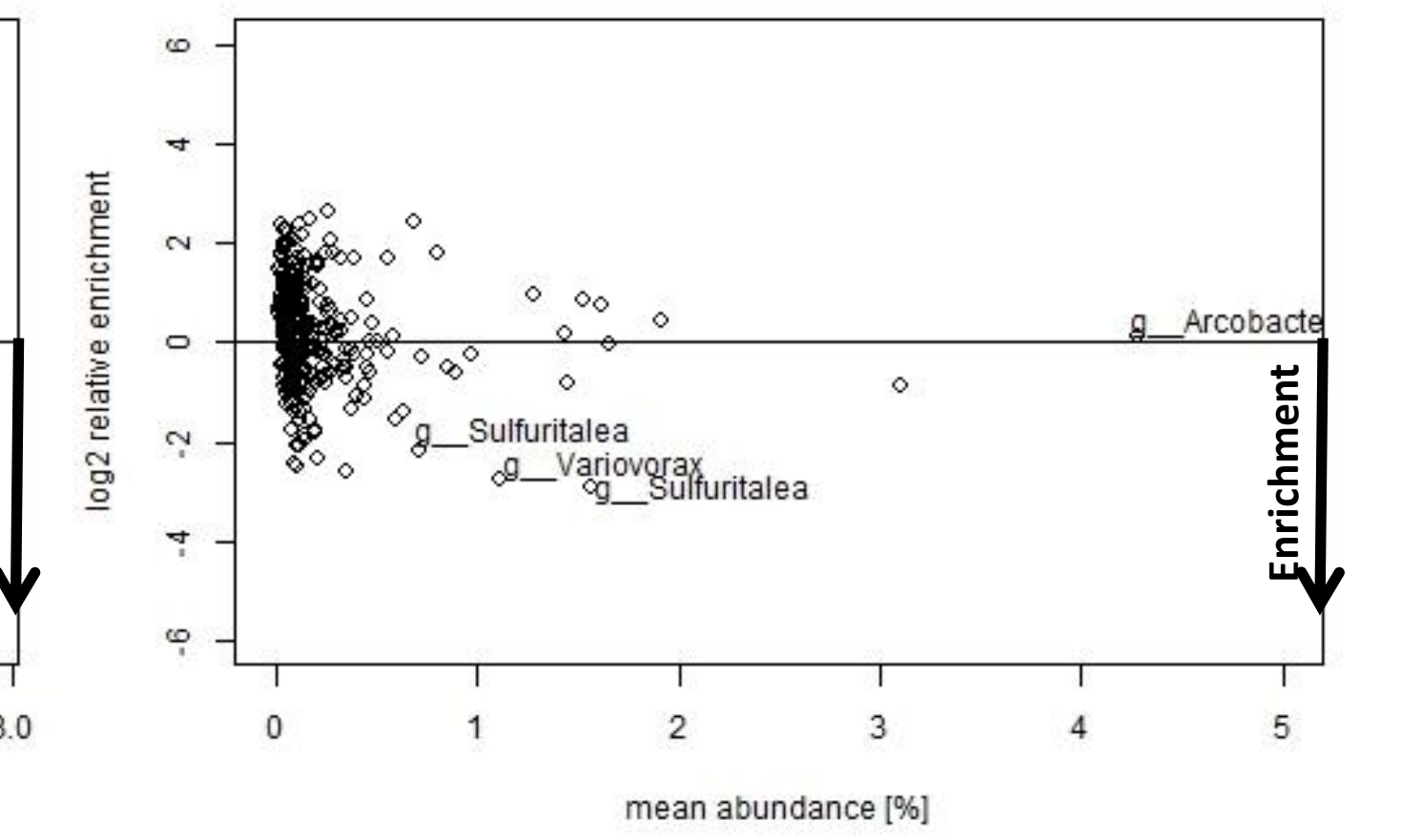
Relative abundance of 10 most frequently occurring genera compared between the different sludge fractions.
The variation between the WWTPs is captured by the boxplot width.



Bacteria enriched in the supernatant compared to total sludge



Bacteria enriched in the supernatant after shear compared to supernatant



SLUDGE VOLUME INDEX					
Total sludge		Supernatant		Supernatant after shear	
OTU	taxonomic classification	R	OTU	taxonomic classification	R
276	Lactococcus	0.8	276	Lactococcus	0.7
979	Arcobacter	0.8	632	Arcobacter	0.8
1188	Caldilineaceae	0.7	1188	Caldilineaceae	0.8
1200	Pseudorhodobacter	0.7	1200	Pseudorhodobacter	0.7
4117	Gordonia	0.7	1033	Thermomonas	0.7
8679	Fodiniibacter	0.7	203	Alicyclophus	0.7
1221	Streptococcus	0.7	1221	Streptococcus	0.6
5480	Nocardoides	0.6	1157	Variovorax	0.7
564	Ferruginibacter	0.6	6976	Cloacibacterium	0.7
790	Chitinophagaceae	0.6	424	Sph35	0.6
626	Leucobacter	0.6	6930	HF_BF35	0.7
978	env_OPS_17	0.7	1925	Unclassified	0.7
185	A0817	0.7	1925	Unclassified	0.7
14179	Sulfuritalea	0.7	5387	Geothrix	0.8
6532	Ferruginibacter	0.7	3960	Betaproteobacteria	0.7
978	Sulfuritalea	0.7	1283	Betaproteobacteria	0.8
1389	Candidatus_Epiflobacter	0.6	15181	Propionibacterium	0.6
8809	WCHB1-60	0.7	9372	PSB	0.6
1395	Caulobacteraceae	0.6	5960	Saccharomonas	0.6
5957	AKW1767	0.7	4338	Holophagaceae	0.8
6267	Rhodobates	0.7	4350	Unclassified	0.7
			10501	0319-6020	0.7

SHEAR SENSITIVITY					
Total sludge		Supernatant		Supernatant after shear	
OTU	taxonomic classification	R	OTU	taxonomic classification	R
995	Trichococcus	0.6	5179	Sulfuritalea	0.8
4117	Chitinophagaceae	0.8	1219	Agromyces	0.6
203	Alicyclophus	0.6	3387	Geothrix	0.8
1581	Propionibacterium	0.7	1581	Propionibacterium	0.7
1048	Propionibacterium	0.7	6336	OT38-B909	0.7
9913	TM214	0.7	14300	Propionibacteriaceae	0.8
1420	Propionibacterium	0.6	354	Thermomonas	0.7
184	Hydrogenobium	0.6	3235	Kinetosphaera	0.7
1012	Leucobacter	0.6	1227	Geothrix	0.8
203	Alicyclophus	0.6	1317	WCHB1-60	0.7
601	Ferruginibacter	0.6	5410	Chloroflex	0.7
1395	Caulobacteraceae	0.6	476	Rhodobacterium	0.7
389	Tetrasphaera	0.6	5551	Hydrogenobium	0.6
			1012	Leucobacter	0.6
			1395	Caulobacter	0.6

Spearman correlation of bacteria present in different sludge fractions with important sludge characteristics: Sludge Volume Index (SVI), shear sensitivity, degree of flocculation and conditions for flocculation. Bacteria that were highly correlated ($R > 0.6$) with the mentioned parameters are listed above.